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- (71) Applicant: ST. ELIZABETH'S MEDICAL CENTER, INC. [US/US]; 736 Cambridge Street, Boston, MA 02135 (US).
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- (72) Inventors: CHISHTI, Athar, H.; 55 Powder Mill Road, Sudbury, MA 01776 (US). OH, S., Steven; 6 Winthrop

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(54) Title: BAND 3 ANTIGENIC PEPTIDES, MALARIA POLYPEPTIDES AND USES THEREOF

(57) Abstract: The invention provides peptides derived from erythroid Band 3 protein, which selectively bind to merozoite surface protein-1 (MSP-1), and/or one or more of the malaria polypeptides: BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and ABRA and prevent infection by the parasite of a Band 3-expressing cell, such as an erythrocyte. The invention also provides the isolated polypeptides BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA as well as peptides derived from MSP-1, which selectively bind to erythroid Band 3 protein and prevent parasite invasion into a Band 3-expressing cell, and prevent *Plasmodium* infection. Methods of using the malaria and MSP1 polypeptides of the invention for malaria prevention and/or treatment (e.g. in vaccines) are also provided. Antibodies that bind to the Band 3 polypeptides and anti-idiotypic antibodies thereto also are provided. Methods for selecting agents which inhibit Band 3-mediated parasite entry into target cells and methods of treatment which involve the polypeptides, antibodies, and anti-idiotypic antibodies also are provided.

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BAND 3 ANTIGENIC PEPTIDES, MALARIA POLYPEPTIDES
AND USES THEREOF

Related Applications

5 This application claims priority under 35 U.S.C. §119 to U.S. 60/272,930, filed March 2, 2001, the entire contents of which is hereby incorporated by reference.

Government Support

10 This invention was made in part with government support under grant number HL60961 and HL60755 from the National Institutes of Health (NIH). The government may have certain rights in this invention.

Field of the Invention

15 This invention relates to polypeptides derived from erythroid Band 3 protein and nucleic acid molecules encoding same. The polypeptides selectively bind to merozoite surface protein-1 (MSP-1) and/or to one or more of the polypeptides: BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and ABRA. The invention also relates in part to nucleic acids that encode the polypeptides BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and ABRA. The invention also relates to
20 polypeptides derived from MSP-1 which selectively bind to Band 3 protein and nucleic acid molecules encoding same. The nucleic acid molecules and encoded polypeptides are useful in, *inter alia*, research, diagnostic and therapeutic contexts, particularly for the development of antibodies and anti-idiotypic antibodies for treating malaria infection.

25

Background of the Invention

 The World Health Organization estimates that 300-500 million people are infected by malaria annually and over 2 million people, mostly women and children under the age of five die of the malaria disease each year. The disease has been
30 classified as an "emerging infection" by many national and international health authorities in recent years, due to its dramatic comeback in regions where the disease is once eliminated or suppressed. Conventional method of control for malaria disease

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mainly relies on the use of antimalarial drugs. Due to a rapid rise and spread of drug resistance to most affordable and widely used drugs in recent years, however, there is unfortunately limited means of treatment for the disease. At present, a malaria vaccine is not available.

- 5 In view of the foregoing, a need exists to develop new and improved methods and compositions for treating malaria infection. Preferably such methods and compositions are based upon inhibiting the particular interactions between the malaria parasite and a cognate molecule present in the host, thereby minimizing harmful side effects that may be due to non-specific therapeutic approaches.

10

Summary of the Invention

- The invention is based, in part, on our discovery that the erythrocyte Band 3 protein is an important receptor for malaria parasite invasion into host erythrocytes. Important regions of the Band 3 protein that form the receptor in human erythrocytes
15 are defined as amino acid residues 720-761 in the ectoplasmic domain 5 and residues 807-826 in the ectoplasmic domain 6. These two ectoplasmic domains of the erythroid Band 3 protein appear to be an important part of the erythrocyte receptor or receptor complex required for the *P. falciparum* invasion of the erythrocytes.

- It now has been discovered that particular sequences within the erythroid Band
20 3 protein (also known as Anion Exchanger 1 or AE1) selectively interact with merozoite surface protein-1 (MSP-1) protein, resulting in entry of the malaria parasite into the erythrocyte host cell. In addition, other polypeptides that selectively interact with the Band-3 derived sequences have been identified. These polypeptides include: BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and ABRA. Accordingly,
25 these particular Band 3-derived sequences are useful for further defining the nature of the interaction(s) between the parasite and the erythrocyte which result in infection, as well as for developing diagnostic and therapeutic agents which are useful for detecting and treating malaria infection. The knowledge of the particular sequences of the Band 3 protein which are important to malaria infection also permits the
30 development of novel anti-idiotypic agents for treating malaria infection. These aspects of the invention are summarized below.

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The invention also is based, in part, on the discovery that the polypeptides BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, ABRA, and particular sequences within MSP-1 selectively interact with Band 3 protein. Accordingly, the polypeptides BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, ABRA and these particular MSP-1 sequences are useful as targets for developing diagnostic and therapeutic agents for detecting and treating malaria infection. These aspects of the invention are summarized below.

In view of the foregoing discoveries, the invention embraces methods for inhibiting the selective interaction between the Band 3 protein and one or more of the polypeptides BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, ABRA, and MSP-1, as well as related compositions. Such methods are useful for identifying compounds for therapeutic use (e.g., screening assays), as well as for diagnosing and/or treating a malaria infection.

In addition, the invention also relates in part to nucleic acids that encode the polypeptides BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, ABRA and MSP-1.

According to one aspect of the invention, isolated Band 3 polypeptides are provided. The isolated Band 3 polypeptides include amino acid sequences selected from the group consisting of SEQ ID NO. 1, 2, 3, and 4 as shown herein: SEQ ID NO:1: GMPWLSATTVRSVTHANALT (also referred to herein as sequence B3_{5A}); SEQ ID NO:2: SVTHANALTVMGKASTPGAA (also referred to herein as sequence B3_{5B}); SEQ ID NO:3: GKASTPGAAQIQEVKEQRI (also referred to herein as sequence B3_{5C}); SEQ ID NO:4: DRILLFLKPPKYHPDVPYVK (also referred to herein as sequence B3_{6A}); and unique fragments thereof, wherein the unique fragments (1) bind to an MSP-1 polypeptide and (2) exclude the sequences set forth in Table 4: Band 3 Blast Homology Sequences. According to another aspect of the invention, isolated nucleic acid molecules that encode the foregoing polypeptides are provided. According to yet another aspect of the invention, expression vectors that include the foregoing nucleic acid molecules operably linked to a promoter are provided. In another aspect of the invention host cells transfected or transformed with the expression vector are provided.

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According to another aspect of the invention, immunogenic compositions are provided. The compositions include one or more of the foregoing isolated Band 3 polypeptides; and a pharmaceutically acceptable carrier; wherein the polypeptides are present in an effective amount to induce an immune system response. In some
5 embodiments, the compositions also include an adjuvant. According to another aspect of the invention, methods of making a medicament are provided. The methods include placing one or more of the foregoing isolated Band 3 polypeptides in a pharmaceutically acceptable carrier.

According to yet another aspect of the invention, methods for identifying a
10 candidate mimetic of the foregoing isolated Band 3 polypeptides are provided. The methods include providing an MSP-1 polypeptide which binds a foregoing Band 3 polypeptide, contacting the MSP-1 polypeptide with a test molecule, and determining the binding of the test molecule to the MSP-1 polypeptide, wherein a test molecule which binds to the MSP-1 polypeptide and inhibits binding of the MSP-1 polypeptide
15 to the foregoing isolated Band 3 polypeptide is a candidate mimetic of the foregoing isolated Band 3 polypeptide.

According to another aspect of the invention, protein microarrays are provided. The microarrays include at least one isolated Band 3 polypeptide selected from the group consisting of SEQ ID NOS. 1, 2, 3, and 4.

According to yet another aspect of the invention, anti-Band 3 antibodies or
20 fragments thereof are provided. The anti-Band 3 antibodies or fragments thereof selectively bind to a foregoing isolated Band 3 polypeptide, wherein the antibody inhibits infection of cells by *P. falciparum* merozoite malaria parasite. Preferably, the antibody is a monoclonal antibody, and, more preferably, a humanized monoclonal
25 antibody.

According to another aspect of the invention, anti-idiotypic antibodies which selectively bind to an idiotype of the foregoing Band 3 antibodies are provided. As used herein, an idiotype refers to a specific binding site of an antibody that binds to the peptide antigen. In accordance with the present invention, the anti-idiotypic
30 antibody blocks penetration of malaria parasite into human red blood cells, presumably by virtue of blocking the malarial parasite ligand that binds to the erythroid Band 3 protein receptor. According to a related aspect of the invention,

methods for making an anti-idiotypic antibody are provided. The methods include immunizing an animal with a foregoing Band 3 antibody under conditions to elicit an immune system response to an idiotype of said foregoing antibody.

According to some aspects of the invention, methods for treating a malaria infection, are provided. The methods include administering to a subject in need of such treatment, an effective amount of a foregoing anti-Band 3 antibody to treat the malaria infection. This method of treatment is referred to herein as "passive immunity".

According to another aspect of the invention, methods for inducing an immune system response to treat a malaria infection are provided. The methods include administering to a subject in need of such treatment, an effective amount of a foregoing anti-Band 3 antibody under conditions to induce an anti-idiotypic immune response to the anti-Band 3 antibody idiotype. This method of treatment is referred to herein as "active immunity".

The invention also is based, in part, on the discovery of the particular portion of MSP-1 (alternatively referred to herein as "MSP1") that selectively binds to Band 3 protein. Thus the invention embraces various compositions containing such MSP-1 peptides for use, e.g., in screening assays to detect the specific interaction between MSP-1 and Band 3 protein, as well as for use in diagnostic and therapeutic applications for detecting and treating, respectively, malaria infection. In general, such compositions contain components (or are contained in kits which contain additional components) which are selected to detect the specific interaction between MSP-1 and Band 3 protein (particularly the Band 3 polypeptides disclosed herein).

According to another aspect of the invention, methods for identifying a candidate mimetic of a MSP-1 polypeptide are provided. The methods include providing an isolated Band 3 polypeptide which binds a MSP-1 polypeptide, contacting the Band 3 polypeptide with a test molecule, and determining the binding of the test molecule to the Band 3 polypeptide, wherein a test molecule which binds to the isolated Band 3 polypeptide and inhibits binding of the Band 3 polypeptide to the MSP-1 polypeptide is a candidate mimetic of the MSP-1 polypeptide. In some embodiments, the MSP-1 polypeptide has a sequence selected from the group consisting of SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:33, SEQ

ID NO. 34, and SEQ ID NO:35. In certain embodiments, the test molecule is an antibody.

According to another aspect of the invention, isolated polypeptides are provided. The polypeptides include an amino acid sequence selected from the group
5 consisting of SEQ ID NOs:11, 12, 13, 33, 34, and 35, or fragments thereof.

According to another aspect of the invention, pharmaceutical compositions are provided. The compositions include one or more of the foregoing isolated MSP-1 polypeptides and a pharmaceutically acceptable carrier; wherein the polypeptides are present in an effective amount to induce an immune system response. In some
10 embodiments, the pharmaceutical composition also includes an adjuvant. According to another aspect of the invention, methods of making a medicament are provided. The methods include placing one or more of the foregoing isolated MSP-1 polypeptides in a pharmaceutically acceptable carrier.

According to yet another aspect of the invention, methods of preventing or
15 treating a malaria infection are provided. The methods include administering a foregoing MSP-1 pharmaceutical composition to a subject in need of such treatment in an amount effective to prevent or treat the malaria infection. According to some aspects of the invention, malaria polypeptide binding polypeptides are provided. The malaria polypeptide binding polypeptides selectively binds to a foregoing isolated
20 MSP-1 polypeptide. Preferably, the binding polypeptide is an antibody or antigen-binding fragment of an antibody. Preferably, the antibody is a monoclonal antibody, and more preferably, a humanized monoclonal antibody. According to another aspect of the invention, pharmaceutical compositions that include the foregoing malaria polypeptide binding polypeptide in a pharmaceutically acceptable carrier are
25 provided.

According to another aspect of the invention, methods of preventing or treating a malaria infection are provided. The methods include administering the foregoing MSP-1 polypeptide binding polypeptide pharmaceutical composition to a subject in need of such treatment in an amount effective to prevent or treat the malaria
30 infection.

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According to another aspect of the invention, isolated nucleic acids are provided. The isolated nucleic acids include a nucleotide sequence selected from the group consisting of SEQ ID NOs:54-59, or fragments thereof.

According to another aspect of the invention, an isolated Band 3 polypeptide
5 is provided. The Band 3 polypeptide includes an amino acid sequence selected from the group consisting of SEQ ID NO. 1, 2, 3, and 4 as shown herein:

SEQ ID NO:1: GMPWLSATTVRSVTHANALT (also referred to herein as sequence B3_{5A});

SEQ ID NO:2: SVTHANALTVMGKASTPGAA (also referred to herein as sequence
10 B3_{5B});

SEQ ID NO:3: GKASTPGAAAQIQEVKEQRI (also referred to herein as sequence B3_{5C});

SEQ ID NO:4: DRILLFKPPKYHPDVPYVK (also referred to herein as sequence B3_{6A}), and unique fragments thereof, wherein the unique fragments (1) bind to an
15 isolated polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:46-53, or fragment thereof, and (2) exclude the sequences set forth in Table 4:Band 3 Blast Homology Sequences.

According to another aspect of the invention, isolated nucleic acid molecules that encode the foregoing isolated Band 3 polypeptides are provided. According to
20 another aspect of the invention, expression vectors are provided. The expression vectors include the foregoing isolated Band 3 nucleic acids operably linked to a promoter. According to another aspect of the invention, host cells transfected or transformed with the foregoing expression vectors are provided.

According to another aspect of the invention, immunogenic compositions are
25 provided. The compositions include one or more of the foregoing isolated Band 3 polypeptides and a pharmaceutically acceptable carrier; wherein the Band 3 polypeptides are present in an effective amount to induce an immune system response. In some embodiments, the compositions also include an adjuvant. According to another aspect of the invention, methods of making a medicament are
30 provided. The methods include placing one or more of the foregoing isolated Band 3 polypeptides in a pharmaceutically acceptable carrier.

According to another aspect of the invention, methods for identifying a candidate mimetic of a foregoing isolated Band 3 polypeptide are provided. The methods include providing a malaria polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:46-53, or fragment thereof that
5 binds the foregoing isolated Band 3 polypeptide or fragment thereof, contacting the malaria polypeptide or fragment thereof, with a test molecule, and determining the binding of the test molecule to the malaria polypeptide or fragment thereof, wherein a test molecule which binds to the polypeptide or fragment thereof and inhibits binding of the foregoing isolated Band 3 polypeptide to the malaria polypeptide, is a candidate
10 mimetic of the foregoing isolated Band 3 polypeptide.

The invention also is based, in part, on the discovery of BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and ABRA polypeptides that selectively binds to Band 3 protein. Thus the invention embraces various compositions containing such BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and ABRA polypeptides for
15 use, e.g., in screening assays to detect the specific interaction between the BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and ABRA polypeptides and Band 3 protein, as well as for use in diagnostic and therapeutic applications for detecting and treating, respectively, malaria infection. In general, such compositions contain components (or are contained in kits which contain additional components) which are
20 selected to detect the specific interaction between BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, or ABRA and Band 3 protein (particularly the Band 3 polypeptides disclosed herein).

According to another aspect of the invention, methods for identifying a candidate mimetic of an isolated malaria polypeptide are provided. The methods
25 include providing a Band 3 molecule which binds a malaria polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:46-53, contacting the Band 3 molecule with a test molecule, and determining the binding of the test molecule to the Band 3 molecule, wherein a test molecule which binds to the Band 3 molecule and inhibits binding of the malaria polypeptide with the Band 3
30 polypeptide is a candidate mimetic of the malaria polypeptide. In some embodiments, the test molecule is an antibody.

According to another aspect of the invention, isolated polypeptide molecules that include amino acid sequences selected from the group consisting SEQ ID NOs:46-52 are provided. According to another aspect of the invention, pharmaceutical compositions are provided. The pharmaceutical compositions include one or more of the foregoing isolated polypeptides and a pharmaceutically acceptable carrier; wherein the polypeptides are present in an effective amount to induce an immune system response. In some embodiments, the pharmaceutical composition also includes an adjuvant. According to another aspect of the invention, methods of making a medicament are provided. The methods include placing one or more of the foregoing isolated polypeptides in a pharmaceutically acceptable carrier. According to another aspect of the invention, methods of preventing or treating a malaria infection are provided. The methods include administering the foregoing pharmaceutical composition to a subject in need of such treatment in an amount effective to prevent or treat the malaria infection.

According to another aspect of the invention, malaria polypeptide binding polypeptides are provided. The malaria polypeptide-binding polypeptides selectively bind to the foregoing isolated malaria polypeptides, e.g., the binding polypeptide is an antibody or antigen-binding fragment of an antibody. Preferably, the antibody is a monoclonal antibody, and more preferably, a humanized monoclonal antibody. According to another aspect of the invention, pharmaceutical compositions that include the foregoing malaria polypeptide binding polypeptide in a pharmaceutically acceptable carrier are provided.

According to another aspect of the invention, methods of preventing or treating a malaria infection are provided. The methods include administering the foregoing pharmaceutical composition to a subject in need of such treatment in an amount effective to prevent or treat the malaria infection.

According to another aspect of the invention, isolated nucleic acid molecules are provided. The nucleic acid molecules are selected from the group consisting of:

(a) nucleic acid molecules which hybridize under stringent conditions to a nucleic acid molecule comprising a nucleotide sequence selected from the group consisting of SEQ ID NO:38-44 and which codes for a *Plasmodium* polypeptide,

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(b) deletions, additions and substitutions of the nucleic acid molecules of (a), which code for a *Plasmodium* polypeptide,

(c) nucleic acid molecules that differ from the nucleic acid molecules of (a) or (b) in codon sequence due to the degeneracy of the genetic code, and

5 (d) complements of (a), (b) or (c).

In some embodiments, the isolated nucleic acid molecule comprises a nucleotide sequence selected from the group consisting of SEQ ID NO:38-44.

According to another aspect of the invention, isolated nucleic acid molecules are provided. The nucleic acid molecules are selected from the group consisting of:

10 (a) a unique fragment of the nucleotide sequence selected from the group consisting of:

nucleotides 1-1287 of SEQ ID NO:38 between 12 and 1286 nucleotides in length,

nucleotides 1-3576 of SEQ ID NO:39 between 12 and 3557 nucleotides in length,

nucleotides 1-903 of SEQ ID NO:40 between 12 and 902 nucleotides in length,

15 nucleotides 1-1203 of SEQ ID NO:41 between 12 and 1202 nucleotides in length,

nucleotides 1-3996 of SEQ ID NO:42 between 12 and 3995 nucleotides in length, and

nucleotides 1-876 of SEQ ID NO:43 between 12 and 875 nucleotides in length, and

nucleotides 1-2712 of SEQ ID NO:44 between 12 and 2711 nucleotides in length, and

(b) complements of (a),

20 wherein the unique fragments exclude nucleic acids having nucleotide sequences that are contained within SEQ ID NO:38-44, and that are known as of the filing date of this application.

According to another aspect of the invention, expression vectors that include the foregoing isolated nucleic acid molecules operably linked to a promoter are
25 provided.

According to another aspect of the invention, isolated polypeptide molecules that include a unique fragment of amino acid sequence SEQ ID NO:53 that binds to a Band 3 polypeptide are provided.

These and other objects of the invention will be described in further detail in
30 connection with the detailed description of the invention.

Brief Description of the Drawings

The figures are illustrative only and are not required for enablement of the inventions disclosed herein.

Figure 1 is a bar graph (Fig. 1A) depicting *P. falciparum* infection of mouse
5 RBCs *in vitro* and a digitized image of a photomicrograph (Fig 1B) showing in
Giemsa-stained thin smears at the end of the 24 h culture. The arrowhead indicates a
newly infected RBC (ring stage parasite). The parasite culture was kept for 22-24 h at
37°C as described (Klotz, F.W., et al., *J Exp Med* 165:1713-1718, 1987). Rings were
counted from 5,000 RBCs in Giemsa-stained thin smears. Data analyzed as the mean
10 of triplicate experiments with standard error.

Figure 2 depicts the inhibition of *P. falciparum* invasion into RBCs by human
Band 3 peptides. Fig. 2A shows a domain map illustration in (ii)-(iv), an inclusive
boundary for each putative ectodomain was chosen as shown by vertical dotted lines.
15 Ectodomains are shown as boxes in the three models. (i) Overlapping 12 to 20-residue
peptides shown in solid bars were prepared according to the putative ectodomain
boundaries. (ii) Casey model (Fujinaga, J., et al., *J Biol Chem* 274:6626-6633, 1999).
(iii) Reithmeier model (Popov, M., et al., *J Biol Chem* 272:18325-18332, 1997). (iv)
Sherman model (Crandall, I., et al., *Parasitology* 108:257-267, 1994). Fig. 2B shows
20 a bar graph depicting an invasion inhibition assay by visual counting method, in
which the number of ring stage parasites in 1,300-1,600 RBCs in Giemsa-stained thin
smears was scored and plotted for each sample. Artemisinin (25 µM) and no peptide
samples were positive and negative controls, respectively. Fig. 2C is a bar graph
depicting of invasion inhibition assay by ³H-hypoxanthine incorporation method.
25 Effects of Band 3 peptides relative to the control sample (no peptide) are shown.
Artemisinin (50 µM) and unrelated peptide derived from dematin were used as
positive and negative control, respectively. Fig.2D is a bar graph that depicts results
of a growth inhibition assay by ³H-hypoxanthine incorporation method. Effects of
Band 3 peptides relative to the control sample (no peptide) are shown. In all
30 inhibition assays, mean parasitemia from three experiments was calculated with
standard error. DMSO background was corrected when necessary. Student's *t* test

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was used to compare with the control (no peptide). Fig. 2E is a table providing a summary of net charge and pI for Band 3 peptides.

Figure 3 shows results of binding of human Band 3 and native *P. falciparum* merozoite proteins. Fig. 3A is a digitized photomicrographic image of a Giemsa-stained smear depicting purified merozoites with malaria pigments but no contaminating RBC components. Fig. 3B is a digitized image of a merozoite protein separation. Total merozoite proteins from purified merozoites and human RBC ghost proteins were separated by SDS-PAGE (M, merozoite; G, RBC ghost). Residual human serum albumin (HSA; apparent mass 67 kDa) from the culture medium is marked with an asterisk in the Coomassie gel. Fig. 3C is a digitized image of a blot overlay assay. The 5C+6A mixture specifically bound to merozoite proteins (arrowheads) but not RBC proteins on the blot. Peptides 3A, 4A, and 2 as negative control showed non-specific bindings to HSA (asterisk) in M and α/β spectrin (240/220 kDa) in G. Fig. 3D is a digitized image of a gel depicting recombinant 5ABC and 5BC expressed in *E. coli*. Coomassie gel (lanes 1-3) and anti-GST Western blot (lanes 4-6) showed affinity purified GST-5BC (lanes 1, 4) and GST-5ABC (lanes 2, 5). GST control sample (lanes 3, 6). Fig. 3E is a digitized image demonstrating native MSP1 binding to 5ABC. Autoradiography showed radiolabeled MSP1 (full length) and MSP1₄₂ bound to GST-5ABC (lane 1) and mAb 5.2 (lane 3), but not to GST (lane 2). Results reproduced three times.

Figure 4 shows binding of recombinant human Band 3 and *P. falciparum* MSP1. Fig. 4A is a digitized image of a Coomassie gel showing recombinant MSP1 and Band 3 (lanes 1-3) and a digitized image of an anti-GST Western blot (lanes 4-6) showing affinity purified GST-MSP1₄₂ (lanes 1, 4), truncated GST-MSP1₃₈ (lanes 2, 5), and GST as control (lanes 3, 6). Truncations of GST-MSP1₃₈ appear to be at the C-terminus as anti-GST antibody reacted with all three major Bands. GST-MSP1₁₉ was co-purified with GST (lane 7, Coomassie; lane 8, anti-GST blot; lane 9, anti-mAb 5.2 blot). Fig. 4A also depicts a digitized image of an autorad showing ³²P-labeled MSP1₁₉ (lane 10), 5BC (lane 11), and 5ABC (lane 12). Figs. 4B, 4C, and 4D show line graphs and a bar graph depicting results of a solution-binding assay. Binding

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assays were performed as described in Example 5 and in Oh, S.S., et al., *Mol Biochem Parasitol* 108:237-247, 2000. ^{32}P -labeled 5ABC (10, 20, 40, 80 μM) and 5BC (21, 42, 84, 168 μM) respectively bound to GST-MSP1₄₂ (Fig. 4B) and GST-MSP1₃₈ (Fig. 4C) on beads in concentration-dependent manner. ^{32}P -labeled MSP1₁₉ bound specifically to the 5ABC domain (Fig. 4D) as statistically analyzed by Student's *t* test. Fig. 4E is a graphical summary of MSP1-Band 3 interactions. Dotted line denotes the C-terminal truncation of MSP1₃₈.

Figure 5. depicts results indicating *P. falciparum* MSP1 binds to intact RBCs in suspension. Fig. 5A is a digitized image of a 10% Coomassie gel of enzyme-treated RBCs (as described in Example 5). The gel (lanes 1-6) shows ghost membrane proteins prepared from untreated human (lane 1) and mouse (lane 4) RBCs, Nm-treated human (lane 2) and mouse (lane 5) RBCs, and ChT-treated (40 min) human (lane 3) and mouse (lane 6) RBCs. Arrowheads and arrows respectively indicate full-length and ChT-digested Band 3. The middle panel is a digitized image of a gel demonstrating PAS staining of the gel, which allowed analysis of sialic acid content in RBC ghosts prepared from untreated (lane 7), ChT-treated (lane 8), and Nm-treated (lane 9) human RBC samples. The right panel depicts a digitized image of a Western blot of ghost proteins prepared from untreated (lanes 10, 13), ChT-treated (lanes 11, 14), and Nm-treated (lane 12) human RBCs using anti-Band 3 and anti-GPA antibody are shown. Fig. 5B is a digitized image of an anti-GST Western blot showing Nm-treated (lane 1) and untreated (lane 3) human RBCs binding GST-MSP1₃₈. GST was used as control (lanes 2, 4). Fig. 5C is a digitized image depicting ^{32}P -labeled MSP1₁₉ binding to various RBC types. Assays were repeated 3-6 times. Means (\pm standard error) were plotted relative to the control (untreated wild-type RBCs) and compared using Student's *t* test.

Figure 6. is a digitized image of an invasion pathway model. In both sialic acid-independent and dependent pathways, Band 3 may function as an important RBC receptor for *P. falciparum* invasion. In the former pathway, Band 3 might be an independent receptor (open arrows) or complemented by GPA to take part in the sialic acid-dependent pathway (dotted arrow). In the latter pathway, GPA appears to be a

non-essential receptor perhaps requiring the coupling of the essential Band 3 receptor (solid black arrows).

5

Detailed Description of the Invention

For over two decades, the malaria parasite protein MSP-1 has been reported to play an important role during the parasite invasion of red blood cells based on the findings that (a) it is a major protein found on the surface of the merozoite, the
10 invasive form of the malaria parasite, (b) full-length MSP-1, a segment of MSP1(38), as well as a number of 20-mer peptides derived from MSP-1 bound to erythrocytes, (c) MSP-1 induced antibodies provided protection against malaria infection in animal models, and (d) deleting a portion of the MSP-1 gene caused destruction of the parasite *in vitro*. However, the exact function of MSP-1 has remained unknown. We
15 have identified for the first time a definitive function of MSP-1 in the malaria parasite invasion of red blood cells. This newly identified function revealed that specific proteolytic fragments of MSP-1 (i.e., MSP1(38), MSP1(42), MSP1(19)) are the parasite ligands specifically binding to the erythrocyte receptor Band 3 during invasion and that the binding interaction between the parasite MSP-1 fragments and
20 the host Band 3 receptor is important for the invasion process to proceed successfully. We have identified that MSP1(19) – the 19 kDa C-terminal fragment of MSP-1 formed by secondary proteolytic processing of MSP1(42) – is also a parasite ligand binding to the Band 3 receptor peptides, e.g., SEQ ID NOs 1, 2, 3, and 4.

The functional form of MSP-1 during malaria parasite invasion into
25 erythrocytes is not full-length MSP-1 (approximately 195-205 kDa depending upon the *Plasmodium falciparum* malaria strain) but its naturally processed proteolytic fragments generally known as MSP1(83), MSP1(30), MSP1(38), MSP1(42), and MSP1(33), and MSP1(19). The first four proteolytic fragments are formed upon primary processing of full-length MSP1. The last two are formed by secondary
30 processing of MSP1(42). These primary and secondary processing products form a non-covalent complex on the surface of merozoites (the invasive form of the parasite) during invasion. However, only MSP1(19) which is anchored to the merozoite

membrane is carried into the newly infected erythrocyte while other fragments are shed into the surrounding medium.

In view of the foregoing, the invention provides isolated Band 3 peptides which selectively bind to merozoite surface protein-1 (MSP-1). The complete nucleic acid and amino acid sequences for human Band 3 protein are described in GenBank
5 acid and amino acid sequences for human Band 3 protein are described in GenBank accession nos. X12609 and M27819 (SEQ ID NOs:5 and 6, (X12609 nucleic acid and amino acid sequences, respectively); and SEQ ID NOs:7 and 8 (M27819 nucleic acid and amino acid sequences, respectively); the complete amino acid and nucleic acid sequences for MSP-1 are described in GenBank accession no. X02919 (SEQ ID
10 NOs:9 and 10, respectively). See also, (e.g., Fujinaga, J., et al. *J Biol Chem* 274:6626-6633, 1999) which reports the topology of the membrane domain of Band 3.

The results disclosed herein suggest that these particular Band 3 peptide sequences selectively interact with MSP-1 and, thereby, facilitate malaria parasite
15 entry into erythrocytes. Such peptides are alternatively referred to herein as "Band 3 immunogenic polypeptides", "Band 3-derived MSP-1 binding peptides", and the like. Hence, one aspect of the invention is an isolated peptide selected from the group of sequences having SEQ ID Nos. 1, 2, 3, and 4, and unique fragments thereof which bind to MSP-1. The selection of these particular sequences and of the particular
20 malaria protein with which they interact, could not have been predicted based on the information presently known regarding the Band 3 structure.

Although not wishing to be bound to any particular theory or mechanism, it is believed that *P. falciparum* depends on the expression on its surface of a specific molecule, MSP-1, also (referred to herein as a ligand), stereochemically
25 complementary to the Band 3 receptor for its binding to and/or, subsequent penetration into the erythrocyte. This ligand can be inhibited in its binding to the receptor (in the specific case of *P. falciparum*, the receptor molecule corresponds to erythroid Band 3) by an antibody which selectively binds to SEQ ID NO. 1, 2, 3, and/or 4 (anti-Band 3 antibody) and which possesses the same stereochemical
30 specificity. Thus, immunization with an anti-Band 3 antibody of the invention as a vaccine is useful for eliciting an immune response to its combining site, termed an anti-idiotypic response, resulting in antibodies which will also recognize the

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immunologically identical epitope on the ligand and, hence, will protect the host against the erythrocytic forms (merozoites) of *P. falciparum*. Thus, the present invention also involves the use of monoclonal anti-Band 3 antibodies immunochemically specific for the epitope used by *P. falciparum* to penetrate into
5 host cells. These monoclonal antibodies are used according to the present invention, to generate or elicit the corresponding anti-idiotypic antibodies which, by virtue of their specificity for the parasite's ligand molecule (MSP-1) are useful in the serodiagnosis and treatment of established infection.

As used herein a "subject" shall mean a human, vertebrate, or invertebrate
10 animal including but not limited to a dog, cat, horse, cow, pig, sheep, goat, non-human primate (e.g. monkey), rabbit, rat, mouse, avian, or insect (e.g. a mosquito). The malarial parasites of the invention include: *P. falciparum*, *P. vivax*, *P. malariae*, *P. ovale*, *P. berghei*, *P. yoelii*, *P. chabaudi*, *P. vinckei*, and *P. knowlesi*, *P. cynomolgi*, and *P. coatneyi*. A preferred malarial parasite of the invention is *P. falciparum*. As
15 used herein, a "malarial infection" includes infection with a malarial parasite including: *P. falciparum*, *P. vivax*, *P. malariae*, *P. ovale*, *P. berghei*, *P. yoelii*, *P. chabaudi*, *P. vinckei*, and *P. knowlesi*, *P. cynomolgi*, and/or *P. coatneyi*. A preferred malaria infection of the invention is infection with *P. falciparum*.

As used herein, the term "cell" means a cell capable of being infected by, or
20 suspected of being exposed to a malarial parasite. This may include cells in or from a subject and cells grown in culture. A cell may also mean a cell collected from a subject such as a human or animal, for example, blood collected for purposes such as, but not limited to, transfusions. In some embodiments, a cell may be a negative control cell, which may be a cell that has not been exposed to a *Plasmodium* parasite.
25 In some embodiments, a positive control cell may be a cell that has been exposed to a *Plasmodium* parasite but is free of a pharmaceutical agent of the invention. A cell is any cell that can be infected by a *Plasmodium* parasite, which includes, but is not limited to: mammalian cells, human cells, avian cells, insect cells, arthropod cells, neuronal cells, ocular cells, erythrocytes, lymphocytes, muscle cells, and intestinal
30 cells.

One class of subjects according to the present invention are subjects having a *Plasmodium* infection. Such subjects are subjects in need of treatment with a

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Plasmodium inhibitor. This class of subjects includes subjects diagnosed with infection, exhibiting symptoms of infection, or having been exposed to a *Plasmodium* parasite. A subject at risk of developing a *plasmodium* infection is a subject in need of prevention of infection. Such subjects include those at risk of exposure to an infection-causing *Plasmodium* parasite. For instance, a subject at risk may be a subject who is planning to travel to an area where a particular type of infectious *Plasmodium* parasite is found or it may be a subject who through lifestyle, occupation, or medical procedures is exposed to bodily fluids which may contain a *Plasmodium* parasite or even any subject living in an area that a *Plasmodium* parasite has been identified. Subjects at risk of developing infection also include general populations to which a medical agency recommends preventative infectious measures for a particular infectious organism.

A subject may or may not exhibit symptoms of infection such as fever, swollen lymph glands, muscle aches, and pains. Methods to diagnose symptomatic and asymptomatic *Plasmodium* infection are known to those of ordinary skill in the medical arts and are described below herein. Some methods of diagnosis include, but are not limited to, blood tests for antibodies to the *Plasmodium* parasite and other assays such as lymph assays for *Plasmodium* parasites.

As noted above, the invention embraces functional variants, such as unique fragments, of the isolated Band 3 polypeptides of the invention which selectively bind to one or more of the peptides: MSP-1 (particularly SEQ ID NOS.:11,12,13, 33, 34, 35), and the polypeptide molecules comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:46-53. As used herein, a "functional variant" or "variant" of a Band 3 polypeptide of the invention is a molecule which contains one or more modifications to the primary amino acid sequence of the Band 3 polypeptide of the invention and retains the MSP-1 binding properties disclosed herein. Modifications which create a Band 3 immunogenic polypeptide functional variant can be made, for example, 1) to enhance a property of a Band 3 binding peptide, such as peptide stability in an expression system or the stability of protein-protein binding such as MSP-1 binding; or 2) to provide a novel activity or property to a Band 3 immunogenic polypeptide, such as addition of an antigenic epitope or addition of a detectable moiety. Modifications to a Band 3 polypeptide of the invention can be

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made to a nucleic acid which encodes the peptide, and can include deletions, point mutations, truncations, amino acid substitutions and additions of amino acids.

Alternatively, modifications can be made directly to the polypeptide, such as by cleavage, addition of a linker molecule, addition of a detectable moiety, such as
5 biotin, substitution of one amino acid for another and the like. Modifications also embrace fusion proteins comprising all or part of the Band 3 polypeptide amino acid sequence.

The amino acid sequence of Band 3 immunogenic polypeptides of the invention may be of natural or non-natural origin, that is, they may comprise a natural
10 Band 3 polypeptide molecule or may comprise a modified sequence as long as the amino acid sequence retains the property of binding to MSP-1 and/or any of the other malaria polypeptides disclosed herein. For example, Band 3 polypeptides in this context may be fusion proteins of a Band 3 polypeptide of the invention and unrelated amino acid sequences, synthetic peptides of amino acid sequences shown in SEQ ID
15 NOs:1, 2, 3, and 4, peptides isolated from cultured cells which express Band 3 peptides, and peptides coupled to nonpeptide molecules (for example in certain drug delivery systems or detectable labels).

Nonpeptide analogs of the Band 3 peptides of the invention, e.g., those which provide a stabilized structure or lessened biodegradation, are also contemplated.
20 Peptide mimetic analogs can be prepared based on a selected Band 3 peptide by replacement of one or more residues by nonpeptide moieties. Preferably, the nonpeptide moieties permit the peptide to retain its natural confirmation, or stabilize a preferred, e.g., bioactive, confirmation. One example of a method for preparation of nonpeptide mimetic analogs from peptides is described in Nachman et al., *Regul. Pept.* 57:359-370 (1995). Peptide mimetics also can be selected from libraries of
25 synthetic compounds (e.g. combinatorial libraries of small organic molecules) or natural molecules according to the MSP-1 and/or other malaria polypeptide binding properties of such molecule (i.e., ability to selectively bind to MSP-1 and/or other malaria polypeptides disclosed herein (isolated or expressed on the surface of a cell or
30 organism)) and/or inhibit parasite entry into human red blood cells. In general, the methods for selection involve determining whether the library's molecules inhibit selective binding of a Band 3 peptide of the invention (e.g., SEQ ID NOs:1, 2, 3, or 4)

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to MSP-1 and/or to other malaria polypeptides disclosed herein and/or block the malaria parasite invasion of red blood cells by for example inhibiting a natural process which merozoites use to penetrate red blood cells.

If a variant involves a change to an amino acid of a Band 3 polypeptide (e.g.,
5 SEQ ID NOs:1, 2, 3, or 4), functional variants of the Band 3 immunogenic polypeptide having conservative amino acid substitutions typically will be preferred, i.e., substitutions which retain a property of the original amino acid such as charge, hydrophobicity, conformation, etc. Examples of conservative substitutions of amino acids include substitutions made amongst amino acids within the following groups:
10 (a) M, I, L, V; (b) F, Y, W; (c) K, R, H; (d) A, G; (e) S, T; (f) Q, N; and (g) E, D.

Other methods for identifying functional variants of the Band 3 polypeptides rely upon the development of amino acid sequence motifs to which potential epitopes may be compared. (See, e.g., published PCT application of Strominger and Wucherpfennig (US/96/03182)). In general, these methods rely upon the
15 development of amino acid sequence motifs to which potential epitopes may be compared. Each motif describes a finite set of amino acid sequences in which the residues at each (relative) position may be (a) restricted to a single residue, (b) allowed to vary amongst a restricted set of residues, or (c) allowed to vary amongst all possible residues. For example, a motif might specify that the residue at a Band 3
20 peptide position may be any one of the residues valine, leucine, isoleucine, methionine, or phenylalanine; that the residue at the second position must be histidine; that the residue at the third position may be any amino acid residue; that the residue at the fourth position may be any one of the residues valine, leucine, isoleucine, methionine, phenylalanine, tyrosine or tryptophan; and that the residue at
25 the fifth position must be lysine.

Sequence motifs for the Band 3 peptide functional variants can be developed by analysis of the MSP-1 (or other malaria polypeptides of the invention) contact points of the Band 3 polypeptides disclosed herein. By providing a detailed structural analysis of the residues involved in the binding of the malaria polypeptides of the
30 invention to the Band 3 polypeptides disclosed herein, one of ordinary skill in the art is enabled to make predictions of sequence motifs for binding between such pairs of proteins.

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Using these sequence motifs as search, evaluation, or design criteria, one of ordinary skill in the art is enabled to identify classes of peptides (functional variants of the Band 3 peptides disclosed herein) which have a reasonable likelihood of binding to and of interacting with MSP-1 (or other malaria polypeptides of the invention) to inhibit parasite entry into erythrocytes. These peptides can be synthesized and tested for activity as described herein. Use of these motifs, as opposed to pure sequence homology (which excludes many peptides which are antigenically similar but quite distinct in sequence) or sequence homology with unlimited "conservative" substitutions (which admits many peptides which differ at critical highly conserved sites), represents a method by which one of ordinary skill in the art can evaluate peptides for potential application in the treatment of disease, such as malaria infection.

The binding of the variant Band 3 peptides to MSP-1 (or other malaria polypeptides of the invention) then is determined according to standard procedures. For example, the variant peptide can be contacted with the MSP-1 which binds the Band 3 peptides of the invention (e.g., SEQ ID NOs:1, 2, 3, or 4) to form a complex of the variant peptide and MSP-1. This contacting can be performed in the presence of Band-3 expressed on erythrocytes to determine whether the variant peptide of Band 3 inhibits binding of the MSP-1 (e.g., expressed by a malaria parasite) and/or entry of the parasite into the erythrocyte.

Variant Band 3 peptides include "unique fragments" of the peptides having SEQ ID NOs:1, 2, 3, and 4. As used herein, a unique fragment is one that is a 'signature' for the larger nucleic acid. It, for example, is long enough to assure that its precise sequence is not found in molecules within the human genome outside of the Band 3 nucleic acids defined above (and human alleles). Those of ordinary skill in the art may apply no more than routine procedures to determine if a fragment is unique within the human genome. Unique fragments, however, exclude fragments completely composed of the nucleotide sequences of any of GenBank accession numbers identified in Table 4 or other previously published sequences as of the filing date of the priority documents for sequences listed in a respective priority document or the filing date of this application for sequences listed for the first time in this application which overlap the sequences of the invention. Thus, unique Band 3

peptide fragments exclude the previously reported peptides identified in Table 4 (Blast results for SEQ ID NOs:1, 2, 3, or 4).

Binding of the variant peptide to the MSP-1 (or other malaria polypeptides of the invention) and/or blocking of the entry of MSP-1 (or other malaria polypeptides of the invention) (e.g., expressed by malaria parasite, containing a detectable label) into erythrocytes (or other cells expressing Band 3) indicates that the variant peptide is a functional variant. The methods also can include the step of comparing the blocking of Band 3-mediated MSP-1 (or other malaria polypeptides of the invention) (e.g., expressed on the merozoite surface) entry into erythrocytes by the Band 3 peptides or antibodies thereto (anti-Band 3 antibodies), as well as anti-idiotypic antibodies, and the blocking by the functional variant as a determination of the effectiveness of the blocking by the functional variant. By comparing the functional variant with the Band 3 peptides or other compositions of the invention disclosed herein, peptides with increased Band 3 blocking properties can be prepared.

Variants of the Band 3 peptides prepared by any of the foregoing methods can be sequenced, if necessary, to determine the amino acid sequence and thus deduce the nucleotide sequence which encodes such variants. Thus, those nucleic acid sequences which code for a Band 3 peptide or variants thereof, including allelic variants, are also a part of the invention. In screening for nucleic acids which encode a Band 3 peptide of the invention, nucleic acid hybridization such as a Southern blot or a Northern blot may be performed under stringent conditions, together with a ^{32}P probe. The term "stringent conditions" as used herein refers to parameters with which the art is familiar. Nucleic acid hybridization parameters may be found in references which compile such methods, e.g. *Molecular Cloning: A Laboratory Manual*, J. Sambrook, et al., eds., Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 1989, or *Current Protocols in Molecular Biology*, F.M. Ausubel, et al., eds., John Wiley & Sons, Inc., New York. Exemplary stringent conditions include hybridization at 65°C in hybridization buffer (3.5 x SSC, 0.02% Ficoll, 0.02% Polyvinyl pyrrolidone, 0.02% Bovine Serum Albumin, 25mM NaH_2PO_4 (pH7), 0.5% SDS, 2mM EDTA). SSC is 0.15M Sodium Chloride/0.015M Sodium Citrate, pH 7; SDS is Sodium Dodecyl Sulphate; and EDTA is Ethylene diaminetetraacetic acid. After hybridization, the membrane upon which the DNA is transferred can be washed,

for example, at 2xSSC at room temperature and then at 0.1 - 0.5x SSC/0.1 x SDS at temperatures up to 68°C. After washing the membrane to which DNA encoding a Band 3 polypeptide is finally transferred, the membrane can be placed against X-ray film to detect the radioactive signal.

5 There are other conditions, reagents, and so forth which can be used, which result in a similar degree of stringency. The skilled artisan will be familiar with such conditions, and thus they are not given here. It will be understood, however, that the skilled artisan will be able to manipulate the conditions in a manner to permit the clear identification of homologs and alleles of nucleic acids encoding the Band 3
10 peptides of the invention. The skilled artisan also is familiar with the methodology for screening cells and libraries for expression of such molecules which then are routinely isolated, followed by isolation of the pertinent nucleic acid molecule and sequencing.

 The invention also includes the use of nucleic acid sequences which include
15 alternative codons that encode the same amino acid residues of the Band 3 peptides of the invention. For example, leucine residues can be encoded by the codons CUA, CUC, CUG, CUU, UUA and UUG. Each of the six codons is equivalent for the purposes of encoding a leucine residue. Thus, it will be apparent to one of ordinary skill in the art that any of the leucine-encoding nucleotide triplets may be employed to
20 direct the protein synthesis apparatus, *in vitro* or *in vivo*, to incorporate a leucine residue. Similarly, nucleotide sequence triplets which encode other amino acid residues comprising the Band 3 peptides include: GUA, GUC, GUG and GUU (valine codons); GGU, GGA, GGG, GGC (glycine codons); UAC and UAU (tyrosine codons). Other amino acid residues may be encoded similarly by multiple nucleotide
25 sequences. Thus, the invention embraces degenerate nucleic acids that differ from the native Band 3 peptide encoding nucleic acids in codon sequence due to the degeneracy of the genetic code.

 Preferred nucleic acids encoding Band 3 polypeptides are those which preferentially express Band 3 peptides, such as those having SEQ ID NOs:1, 2, 3, or
30 4. The Band 3 nucleic acids of the invention do not encode the entire Band 3 polypeptide but do include nucleotide sequences encoding the Band 3 peptides disclosed herein.

The invention also provides modified nucleic acid molecules which include additions, substitutions and deletions of one or more nucleotides. In preferred embodiments, these modified nucleic acid molecules and/or the polypeptides they encode retain at least one activity or function of the unmodified nucleic acid molecule and/or the polypeptides, such as antigenicity, ligand binding, formation of complexes by binding of peptides to MSP-1 (or other malaria polypeptides of the invention), etc. In certain embodiments, the modified nucleic acid molecules encode modified polypeptides, preferably polypeptides having conservative amino acid substitutions as are described elsewhere herein. The modified nucleic acid molecules are structurally related to the unmodified nucleic acid molecules and in preferred embodiments are sufficiently structurally related to the unmodified nucleic acid molecules so that the modified and unmodified nucleic acid molecules hybridize under stringent conditions known to one of skill in the art.

For example, modified nucleic acid molecules which encode polypeptides having single amino acid changes can be prepared. Each of these nucleic acid molecules can have one, two or three nucleotide substitutions exclusive of nucleotide changes corresponding to the degeneracy of the genetic code as described herein. Likewise, modified nucleic acid molecules which encode polypeptides having two amino acid changes can be prepared which have, e.g., 2-6 nucleotide changes. Numerous modified nucleic acid molecules like these will be readily envisioned by one of skill in the art, including for example, substitutions of nucleotides in codons encoding amino acids 2 and 3, 2 and 4, 2 and 5, 2 and 6, and so on. In the foregoing example, each combination of two amino acids is included in the set of modified nucleic acid molecules, as well as all nucleotide substitutions which code for the amino acid substitutions. Additional nucleic acid molecules that encode polypeptides having additional substitutions (i.e., 3 or more), additions or deletions (e.g., by introduction of a stop codon or a splice site(s)) also can be prepared and are embraced by the invention as readily envisioned by one of ordinary skill in the art. Any of the foregoing nucleic acids or polypeptides can be tested by routine experimentation for retention of structural relation or activity to the nucleic acids and/or polypeptides disclosed herein.

It will also be understood that the invention embraces the use of the sequences in expression vectors, as well as to transfect host cells and cell lines, be these prokaryotic (e.g., *E. coli*), or eukaryotic (e.g., CHO cells, COS cells, yeast expression systems and recombinant baculovirus expression in insect cells). The expression
5 vectors require that the pertinent sequence, i.e., those described *supra*, be operably linked to a promoter.

Delivery of expression vectors encoding the Band 3 sequences *in vivo* and/or *in vitro* can be via the use of nucleic acid delivery systems known in the art (see, e.g., Allsopp et al., *Eur. J. Immunol.* 26(8):1951-1959, 1996). Recombinant vectors
10 including viruses selected from the group consisting of adenoviruses, adeno-associated viruses, poxviruses including vaccinia viruses and attenuated poxviruses such as NYVAC, Semliki Forest virus, Venezuelan equine encephalitis virus, retroviruses, Sindbis virus, and Ty virus-like particle, plasmids (e.g. "naked" DNA), bacteria (e.g. the bacterium Bacille Calmette Guerin, BCG), and the like can be used
15 in such delivery, for example, for use as a vaccine. Other viruses, expression vectors and the like which are useful in preparation of a vaccine are known to one of ordinary skill in the art. One can test the Band 3 delivery systems in standard model systems such as mice to determine efficacy of the delivery system. The systems also can be tested in human clinical trials.

20 As used herein, a "vector" may be any of a number of nucleic acids into which a desired sequence may be inserted by restriction and ligation for transport between different genetic environments or for expression in a host cell. Vectors are typically composed of DNA although RNA vectors are also available. Vectors include, but are not limited to, plasmids, phagemids, bacteria and virus genomes as disclosed herein,
25 such as adenovirus, poxvirus and BCG. A cloning vector is one which is able to replicate in a host cell or be replicated after its integration into the genome of a host cell, and which is further characterized by one or more endonuclease restriction sites at which the vector may be cut in a determinable fashion and into which a desired DNA sequence may be ligated such that the new recombinant vector retains its ability
30 to replicate in the host cell. In the case of plasmids, replication of the desired sequence may occur many times as the plasmid increases in copy number within the host bacterium or just a single time per host before the host reproduces by mitosis. In

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the case of phage, replication may occur actively during a lytic phase or passively during a lysogenic phase. An expression vector is one into which a desired DNA sequence may be inserted by restriction and ligation such that it is operably joined to regulatory sequences and may be expressed as an RNA transcript. Vectors may
5 further contain one or more marker sequences suitable for use in the identification of cells which have or have not been transformed or transfected with the vector.

Markers include, for example, genes encoding proteins which increase or decrease either resistance or sensitivity to antibiotics or other compounds, genes which encode enzymes whose activities are detectable by standard assays known in the art (e.g., β -galactosidase, luciferase or alkaline phosphatase), and genes which visibly affect the
10 phenotype of transformed or transfected cells, hosts, colonies or plaques (e.g., green fluorescent protein). Preferred vectors are those capable of autonomous replication and expression of the structural gene products present in the DNA segments to which they are operably joined.

15 As used herein, a coding sequence and regulatory sequences are said to be "operably" joined when they are covalently linked in such a way as to place the expression or transcription of the coding sequence under the influence or control of the regulatory sequences. If it is desired that the coding sequences be translated into a functional protein, two DNA sequences are said to be operably joined if induction of a
20 promoter in the 5' regulatory sequences results in the transcription of the coding sequence and if the nature of the linkage between the two DNA sequences does not (1) result in the introduction of a frame-shift mutation, (2) interfere with the ability of the promoter region to direct the transcription of the coding sequences, or (3) interfere with the ability of the corresponding RNA transcript to be translated into a protein.

25 Thus, a promoter region would be operably joined to a coding sequence if the promoter region were capable of effecting transcription of that DNA sequence such that the resulting transcript might be translated into the desired protein or polypeptide. As noted above, certain preferred nucleic acids express only fragments of Band 3 polypeptides which bind to the malaria peptides disclosed herein.

30 The precise nature of the regulatory sequences needed for gene expression may vary between species or cell types, but shall in general include, as necessary, 5' non-transcribed and 5' non-translated sequences involved with the initiation of

transcription and translation respectively, such as a TATA box, capping sequence, CAAT sequence, and the like. Especially, such 5' non-transcribed regulatory sequences will include a promoter region which includes a promoter sequence for transcriptional control of the operably joined gene. Regulatory sequences may also
5 include enhancer sequences or upstream activator sequences as desired. The vectors of the invention may optionally include 5' leader or signal sequences. The choice and design of an appropriate vector is within the ability and discretion of one of ordinary skill in the art.

Expression vectors containing all the necessary elements for expression are
10 commercially available and known to those skilled in the art. See, e.g., Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Second Edition, Cold Spring Harbor Laboratory Press, 1989. Cells are genetically engineered by the introduction into the cells of heterologous DNA (RNA) encoding a Band 3 peptide of the invention. That heterologous DNA (RNA) is placed under operable control of transcriptional elements
15 to permit the expression of the heterologous DNA in the host cell.

Preferred systems for mRNA expression in mammalian cells are those such as pcDNA3.1 (available from Invitrogen, Carlsbad, CA) that contain a selectable marker such as a gene that confers G418 resistance (which facilitates the selection of stably transfected cell lines) and the human cytomegalovirus (CMV) enhancer-promoter
20 sequences. Additionally, suitable for expression in primate or canine cell lines is the pCEP4 vector (Invitrogen), which contains an Epstein Barr virus (EBV) origin of replication, facilitating the maintenance of plasmid as a multicopy extrachromosomal element. Another expression vector is the pEF-BOS plasmid containing the promoter of polypeptide Elongation Factor 1 α , which stimulates efficiently transcription *in*
25 *vitro*. The plasmid is described by Mishizuma and Nagata (*Nuc. Acids Res.* 18:5322, 1990), and its use in transfection experiments is disclosed by, for example, Demoulin (*Mol. Cell. Biol.* 16:4710-4716, 1996). Still another preferred expression vector is an adenovirus, described by Stratford-Perricaudet, which is defective for E1 and E3 proteins (*J. Clin. Invest.* 90:626-630, 1992). The use of the adenovirus to express
30 proteins for immunization is disclosed by Warnier et al., in intradermal injection in mice for immunization against P1A (*Int. J. Cancer*, 67:303-310, 1996).

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The invention also embraces so-called expression kits, which allow the artisan to prepare a desired expression vector or vectors. Such expression kits include at least separate portions of at least two of the previously discussed materials. Other components may be added, as desired.

5 The invention further includes nucleic acid or protein microarrays which include Band 3 nucleic acids or peptides of the invention (preferably at least one isolated Band 3 peptide selected from the group consisting of SEQ ID NO:1, 2, 3, or 4) or nucleic acids encoding such peptides. In this aspect of the invention, standard techniques of microarray technology are utilized to assess expression of the Band 3
10 binding peptides (e.g., anti-Band 3 antibodies) and/or identify biological constituents that bind such peptides. The constituents of biological samples include antibodies, MSP-1 molecules, other of the malaria peptide disclosed herein, and the like. Microarray technology, which is also known by other names including: protein chip technology and solid-phase protein array technology, is well known to those of
15 ordinary skill in the art and is based on, but not limited to, obtaining an array of identified peptides or proteins on a fixed substrate, binding target molecules or biological constituents to the peptides, and evaluating such binding. See, e.g., G. MacBeath and S.L. Schreiber, "Printing Proteins as Microarrays for High-Throughput Function Determination," *Science* 289(5485):1760-1763, 2000.

20 Microarray substrates include but are not limited to glass, silica, aluminosilicates, borosilicates, metal oxides such as alumina and nickel oxide, various clays, nitrocellulose, or nylon. The microarray substrates may be coated with a compound to enhance synthesis of a probe (peptide or nucleic acid) on the substrate. Coupling agents or groups on the substrate can be used to covalently link the first
25 nucleotide or amino acid to the substrate. A variety of coupling agents or groups are known to those of skill in the art. Peptide or nucleic acid probes thus can be synthesized directly on the substrate in a predetermined grid. Alternatively, peptide or nucleic acid probes can be spotted on the substrate, and in such cases the substrate may be coated with a compound to enhance binding of the probe to the substrate. In
30 these embodiments, presynthesized probes are applied to the substrate in a precise, predetermined volume and grid pattern, preferably utilizing a computer-controlled robot to apply probe to the substrate in a contact-printing manner or in a non-contact

manner such as ink jet or piezo-electric delivery. Probes may be covalently linked to the substrate.

In some embodiments, one or more control peptide or nucleic acid molecules are attached to the substrate. Preferably, control nucleic acid molecules allow
5 determination of factors such as binding characteristics, reagent quality and effectiveness, hybridization success, and analysis thresholds and success.

The present invention also concerns a general method for producing immunity against an infectious microorganism based on the stereochemical complementarity between the ligand molecule on the microorganism and its receptor in the host's cell.
10 This complementarity is used according to the present invention to generate antibodies against the ligand by immunizing with monoclonal antibodies against the ligand-binding region of the receptor molecule. Such monoclonal anti-receptor antibodies, which react with the exact area of binding of the ligand molecule of the parasitic organism, bring about protective immunity because they elicit anti-idiotypic
15 antibodies that react with the stereochemically equivalent region of the ligand.

The anti-Band 3 antibodies of the inventions selectively bind to SEQ ID NOs:1, 2, 3, and/or 4 and, thereby block penetration of *P. falciparum* malaria parasite into human red blood cells by virtue of effectively blocking the site on the erythrocytic molecule (Band 3) used as a target by the *P. falciparum* malaria parasite.
20 Accordingly, the invention provides monoclonal antibodies which have a combining site that has the same stereochemical configuration as the ligand site (e.g., MSP-1) on the *P. falciparum* malaria parasite. Such anti-Band 3 antibodies and anti-idiotypic antibodies are prepared by standard methods. The selected cloned hybridomas produce as large quantities of suitable monoclonal antibodies as desired.

25 In view of the foregoing, the invention also permits the artisan to treat a subject having a malaria infection. Treatments include administering an anti-Band 3 binding peptide (e.g., an anti-Band 3 antibody) or other agent which inhibits binding of MSP-1 (or other malaria polypeptides of the invention) expressed by a malaria parasite to Band 3 expressed by an erythrocyte. Agents useful in the foregoing
30 treatments include Band 3 polypeptides and functional variants thereof, as well as the anti-Band 3 antibodies and anti-idiotypic antibodies of the invention disclosed herein.

The function or status of a pharmaceutical agent as an *Plasmodium* inhibitor, can be determined according to assays known in the art or described herein. For example, cells can be contacted with a putative pharmaceutical agent and a *Plasmodium* parasite, and standard procedures can be used to determine whether the parasite is inhibited in its ability to enter or infect the cells. Such methods may also be utilized to determine the status of analogs, variants, derivatives, and fragments as inhibitors of invasion by *Plasmodium* parasites. One method for inhibiting infection is by inhibiting entry of *Plasmodium* parasite into cells. The ability to inhibit entry of *Plasmodium* parasite into cells with a putative pharmaceutical agent can be assessed using routine screening assays, e.g. by determining the level of entry of *Plasmodium* parasite into cells with and without the presence of the putative pharmaceutical agent.

Once the pharmaceutical agents are verified as modulating *Plasmodium* parasitic infection using secondary assays as described above herein, further biochemical and molecular techniques may be used to identify the targets of these compounds and to elucidate the specific roles that these target molecules play in the process of invasion. An example, though not intended to be limiting, is that the compound(s) may be labeled and contacted with a parasite to identify the host cell proteins with which these compounds interact. Such proteins may be purified, e.g., by labeling the compound with an immunoaffinity tag and applying the protein-bound compound to an immunoaffinity column.

In addition, the status of a pharmaceutical agent as a *Plasmodium* parasite toxin can be identified by using methods provided herein to determine the presence of a functional, active *Plasmodium* parasite. The agent may for example be assayed in the context of a material, for example a water sample, before and after contact with the sample and the pharmaceutical agent.

When administered, the therapeutic compositions of the present invention are administered in pharmaceutically acceptable preparations. Such preparations may routinely contain pharmaceutically acceptable concentrations of salt, buffering agents, preservatives, compatible carriers, and optionally other therapeutic agents.

The term "pharmaceutically acceptable" means a non-toxic material that does not interfere with the effectiveness of the biological activity of the active ingredients. The characteristics of the carrier will depend on the route of administration.

The therapeutics of the invention can be administered by any conventional route, including injection or by gradual infusion over time. The administration may, for example, be oral, intravenous, intraperitoneal, intramuscular, intranasal, intracavity, subcutaneous, intradermal, or transdermal.

5 Preparations for parenteral administration include sterile aqueous or non-aqueous solutions, suspensions, and emulsions. Examples of non-aqueous solvents are propylene glycol, polyethylene glycol, vegetable oils such as olive oil, and injectable organic esters such as ethyl oleate. Aqueous carriers include water, alcoholic/aqueous solutions, emulsions or suspensions, including saline and buffered
10 media. Parenteral vehicles include sodium chloride solution, Ringer's dextrose, dextrose and sodium chloride, lactated Ringer's or fixed oils. Intravenous vehicles include fluid and nutrient replenishers, electrolyte replenishers (such as those based on Ringer's dextrose), and the like. Preservatives and other additives may also be present such as, for example, antimicrobials, anti-oxidants, chelating agents, and inert
15 gases and the like.

The preparations of the invention are administered in effective amounts. An effective amount is that amount of a pharmaceutical preparation that alone, or together with further doses, stimulates the desired response. In the case of treating an infectious disease such as a *Plasmodium* infection, the desired response is inhibiting
20 the onset, stage or progression of the disease or infection. This may involve only slowing the progression of the disease temporarily, although more preferably, it involves halting the progression of the disease permanently. An effective amount for preventing infection is that amount that reduces the incidence of active infection when the cell or subject is exposed to the parasite, with respect to that amount that would
25 occur in the absence of the active agent.

In another aspect of the invention, cell models and/or non-human animal models of *Plasmodium* infection may be produced by administering a molecule of the invention, such as an antibody, that inhibits *Plasmodium* infection. In some cases, a molecule of the invention that enhances *Plasmodium* invasion may be administered to
30 an animal or cell. Such models may be useful for testing treatment strategies, monitoring clinical features of disease, or as tools to assess prevention strategies of *Plasmodium* infection. Cells and animal models made using enhancing molecules of

the invention may also be useful for assessing the ability of lead compounds to inhibit *Plasmodium* infection. For example, a cell contacted with an enhancer of invasion of the invention may be further contacted with putative agents that are candidate or lead compounds for treating or preventing *Plasmodium* infection. The ability of the lead or candidate compound to prevent or treat the infection may be evaluated in the model cell or animal. In addition the enhancers may serve as valuable lead compounds in that if their targets (by definition functionally important) can be identified and characterized, it may subsequently be possible to rationally design new compounds that act as inhibitors of these targets. As used herein, an "effective amount of an enhancer" is that amount effective to enhance *Plasmodium* parasitic infection. Such enhancements can be determined using standard assays as described above herein. Measurements of *Plasmodium* parasitic infection, are known to those of ordinary skill in the art and may vary depending on the specific parasite.

The pharmaceutical compound dosage may be adjusted by the individual physician or veterinarian, particularly in the event of any complication. A therapeutically effective amount typically varies from 0.01 mg/kg to about 1000 mg/kg, preferably from about 0.1 mg/kg to about 200 mg/kg, and most preferably from about 0.2 mg/kg to about 20 mg/kg, in one or more dose administrations daily, for one or more days.

The absolute amount will depend upon a variety of factors, including the material selected for administration, whether the administration is in single or multiple doses, and individual patient parameters including age, physical condition, size, weight, and the stage of the disease. These factors are well known to those of ordinary skill in the art and can be addressed with no more than routine experimentation.

Diagnostic tests known to those of ordinary skill in the art may be used to assess *Plasmodium* infection status of a subject and to evaluate a therapeutically effective amount of a pharmaceutical agent administered. Examples of diagnostic tests are set forth below. A first determination of *Plasmodium* infection may be obtained using one of the methods described below (or other methods known in the art), and a subsequent determination of infection may be done. A comparison of the infection levels may be used to assess the effectiveness of administration of a

pharmaceutical agent of the invention as a prophylactic or a treatment of the *Plasmodium* infection. Absence of a *Plasmodium* infection may be an indication for prophylactic intervention by administering a pharmaceutical agent described herein to prevent *Plasmodium* infection.

5 Tests useful for diagnosis of *Plasmodium* infections are known to those of ordinary skill in the art. For example, diagnosis of malaria can be done by microscopic identification of asexual forms of the parasite in peripheral blood smears stained with Romanovsky staining, or Giemsa at pH 7.2, Wright's, Field's, or Leishman's stain. Both thin and thick blood smears may be examined. In addition, a
10 finger-prick blood test is also available, in which the presence of *P. falciparum* histidine-rich protein 2 is determined. Additional methods of diagnosis and assessment of *Plasmodium* infection are known to those of skill in the art. The level of parasitemia may be important in the prognosis and can be determined with the above-identified diagnostic tests and by other means known in the art.

15 In addition to the diagnostic tests described above, clinical features of *Plasmodium* infection can be monitored for assessment of infection. These features include, but are not limited to: normochromic, normocytic anemia, erythrocyte sedimentation rate, plasma viscosity, and platelet count may be reduced. Subjects may also have metabolic acidosis, with low plasma concentrations of glucose,
20 sodium, bicarbonate, calcium, phosphate, and albumin together with elevations in lactate, blood urea nitrogen, creatinine, urate, muscle and liver enzymes, and conjugated and unconjugated bilirubin. In adults and children with cerebral malaria, the mean opening pressure at lumbar puncture is about 160 mm cerebrospinal fluid; the cerebrospinal fluid usually is normal or has a slightly elevated total protein level
25 [<1.0 g/L (100 mg/dL)] (see Harrison's Principles of Internal Medicine, 14/e, McGraw Hill Companies, New York, 1998).

 The identification of *Plasmodium* parasites in or on an object, may be performed via standard diagnostic methods described above including microscopic examination, antibody labeling in a sample of the object, and by PCR analysis of a
30 sample.

 The pharmaceutical agents of the invention may be administered alone, in combination with each other, and/or in combination with other anti-*Plasmodium* drug

therapies. Anti-malarial agents (for treatment and/or prophylaxis) that may be administered with pharmaceutical agents of the invention include, but are not limited to: mefloquine, doxycycline, chloroquine, aminoquinolines, dihydrofolate reductase inhibitors: pyrimethamine and proguanil (chloroguanide), dapsone, quinidine
5 gluconate, quinine, artemisinin derivatives: artemether and artesunate, and primaquin. Methods of anti-*Plasmodium* treatment of the invention may also be used in combination with drugs that target sialic acid interactions.

The above-described drug therapies are known to those of ordinary skill in the art and are administered by modes known to those of skill in the art. The drug
10 therapies are administered in amounts that are effective to achieve the physiological goals (to reduce *Plasmodium* infection, and/or reduce *Plasmodium* parasite titer in a subject), in combination with the pharmaceutical agents of the invention. Thus, it is contemplated that the drug therapies may be administered in amounts which are not capable of preventing or reducing the physiological consequences of the *Plasmodium*
15 infections when the drug therapies are administered alone, but which are capable of preventing or reducing the physiological consequences of *Plasmodium* infection when administered in combination with the pharmaceutical agents of the invention.

The pharmaceutical agents of the invention may also be administered in conjunction with vaccine formulations administered to confer immunity to a subject at
20 risk of exposure to *Plasmodium* infection, which thereby prevents, reduces the severity of, or delays the onset of a subsequent *Plasmodium* infection.

The invention also provides a pharmaceutical kit comprising one or more containers comprising one or more of the pharmaceutical agents of the invention and or formulations of the invention. The kit may also include instructions for the use of
25 the one or more pharmaceutical agents or formulations of the invention for the treatment of *Plasmodium* infection.

In other aspects the invention involves preventing and/or treating *Plasmodium* contamination of materials. A "material" as used herein is any liquid or solid material including, but not limited to: blood, tissue, bodily fluids, and tissue-processing
30 equipment, including but not limited to: equipment for food processing, medical equipment, equipment for tissue transplant processing, and equipment for cell or bodily fluid processing. In some embodiments of the invention, the material is

aqueous. In some embodiments, the material is water, an example of which, although not intended to be limiting, is drinking water. The invention also involves preventing and/or treating *Plasmodium* contamination in blood, bodily fluids, cells, and tissue samples, including those from live human subjects and cadavers, as well as live
5 animals and animal tissues and cells processed as food, cosmetics, or medication. As used herein, the term "contamination" means contact between the material and a *Plasmodium* parasite.

The isolated Band 3 peptides or anti-Band 3 antibodies of the invention may be combined with materials such as adjuvants to produce vaccines to prepare,
10 respectively, anti-Band 3 antibodies that selectively bind to the portion of Band 3 involved in binding to MSP-1 and to prepare anti-idiotypic antibodies that selectively bind to the portion of MSP-1 involved in binding to Band 3. Vaccines also encompass expression vectors and naked DNA or RNA, encoding a Band 3 peptide or anti-Band 3 antibody of the invention, precursors thereof, or fusion proteins thereof,
15 which may be produced *in vitro* and administered via injection, particle bombardment, nasal aspiration and other methods. Vaccines of the "naked nucleic acid" type have been demonstrated to provoke an immunological response including generation of CTLs specific for the peptide encoded by the naked nucleic acid (*Science* 259:1745-1748, 1993).

20 In certain embodiments, the Band 3 peptides and anti-Band 3 antibodies of the invention are used to produce antibodies ("anti-Band 3 antibodies") which, in turn, may be used to produce "anti-idiotypic Band 3 antibodies", using standard techniques well known to the art. Standard reference works setting forth the general principles of antibody production include Catty, D., Antibodies, A Practical Approach, Vol. 1, IRL
25 Press, Washington DC (1988); Klein, J., Immunology: The Science of Cell-Non-Cell Discrimination, John Wiley and Sons, New York (1982); Kennett, R., et al., Monoclonal Antibodies, Hybridoma, A New Dimension In Biological Analyses, Plenum Press, New York (1980); Campbell, A., Monoclonal Antibody Technology, in Laboratory Techniques and Biochemistry and Molecular Biology, Vol. 13 (Burdon,
30 R. et al. EDS.), Elsevier Amsterdam (1984); and Eisen, H.N., Microbiology, third edition, Davis, B.D. et al. EDS. (Harper & Rowe, Philadelphia (1980). See also, U.S. patent no. 5,101,017, issued March 31, 1992 to Rubinstein, et al., entitled,

“Antibodies for providing protein against *P. vivax* malaria infection,” which also reports the preparation of anti-idiotypic antibodies for treating infectious disease.

References which report vaccine approaches for treating malaria include: U.S. patent no. 6,066,623, issued to Hoffman, et al., entitled “Polynucleotide vaccine protective against malaria, methods of protection and vector for delivering polynucleotide vaccines”; and U.S. patent no. 6,120,770, issued to Adams et al., entitled “*Plasmodium* proteins useful for preparing vaccine compositions.” Additional references which report anti-idiotypic vaccines for treating various disorders include: Bendandi, *Leukemia*, 2000, 14(8):1333-9; Bhattacharya-Chatterjee et al., *Immunol. Lett.*, 2000, 15:74(1):51-8; Maruyama et al., *Cancer Immunol. Immunother.*, 2000, 49(3):123-32; Herlyn et al., *Exp. Clin. Immunogenet.*, 1988, 5(4):165-75; Finberg et al., *Crit. Rev. Immunol.* 1987, 7(4):269-84; Nisonoff, *American Association of Immunologists*, 1991, 147(8):2429-2438; Greenspan et al., *FASEB J.*, 1993, 7:437-444; and Syrengelas et al., *The Journal of Immunology*, 1999, 162:4790-4795.

Thus, according to one aspect of the invention, an anti-Band 3 antibody (or fragment thereof) that selectively binds to a peptide having SEQ ID NO:1, 2, 3, and/or 4 and which blocks penetration of *P. falciparum* merozoite malaria parasite into human red blood cells is provided. According to yet another aspect of the invention, an anti-idiotypic antibody which selectively binds to the idiotypic of the anti-Band 3 antibodies described herein is provided. The antibodies of the present invention are prepared using any of a variety of methods, including administering the Band 3 peptides of the invention, fragments of the foregoing, antibodies selective for the foregoing, and the like to an animal to induce monoclonal or polyclonal antibodies. The production of monoclonal antibodies is according to techniques well known in the art.

Since the antibodies selective for the Band 3 epitope(s) (herein, anti-Band 3 antibodies) and the malaria polypeptide ligand(s) on the parasite recognize the same epitope, antibodies specific for the combining site on the anti-Band 3 antibodies (anti-idiotypic antibodies), can be elicited, which will react with the parasite's ligand(s). Susceptible individuals who make these anti-idiotypic antibodies will be protected against *P. falciparum* merozoites because (a) they block the ability of the parasite to

recognize the erythrocytic receptor and (b) they may induce the lysis or inactivation of the *Plasmodium* cell by fixing complement.

The monoclonal anti-Band 3 antibodies according to the invention have the following characteristics and properties:

5 (1) The antibodies can be used as the immunogenic agent in a vaccine and can be produced in virtually limitless quantity.

(2) The region of the antibody molecule that bears the immunogenic moiety (idiotype) is located in the variable region which may be further purified in order to avoid the undesired immunogenicity of the constant region of the molecule.

10 (3) If desired, the antigen combining region of the antibody can be transferred to a carrier molecule devoid of additional immunogenic properties for human subjects. As discussed below, this may be done by a number of methods that are equivalent in this regard, such as, by chemically binding the Fab fragment of the antibody molecule to an Fc fragment derived from human immunoglobulin or by
15 genetically engineering an appropriate hybrid molecule using the necessary portion of the rearranged immunoglobulin heavy and light chain genes from the monoclonal-producing hybridoma cell line into human immunoglobulin genes from which the equivalent regions have been excised. Alternatively, the idiotype-bearing portions of the protein (or the DNA fragments encoding them) may be attached to other
20 immunogenic molecules or particles (or to their respective genetic determinants in the case of the DNA fragments encoding the idiotypes).

(4) The Band 3 peptides of the invention may be used for binding to and isolating the parasite's specific recognition molecule(s) (e.g., MSP-1). The purified ligand molecule(s) from the merozoite can then be characterized and used as "blue-
25 prints" for the preparation of synthetic peptides (Band 3 functional variants) with protective immunogenic properties.

(5) The antibodies can be used to prepare anti-idiotypic monoclonal antibodies in mice. Those anti-idiotypic antibodies that additionally react with the combining site of *P. falciparum* merozoites can be used as affinity probes, to isolate the ligand as is
30 described in (4) above for the receptor on red blood cells and with the same objectives.

(6) The antibodies and the anti-idiotypic antibody can be used in the immunodiagnosis of *P. falciparum* infection. Thus, the presence of *P. falciparum* antigen in serum or other fluid may be detected and its concentration measured by its interference with the binding of the monoclonal anti-Band 3 to either the Band 3 molecule or to its monoclonal anti-idiotypic antibody. Since the parasite's ligand(s) and the anti-Band 3 antibodies will react with the same respective combining site both on the Band 3 molecule and in the monoclonal anti-idiotypic immunoglobulin, a simple competition assay can be designed using either enzyme-linked or radiolabeled reagents, or other labeling reagents.

(7) The anti-Band 3 antibodies may be used directly *in vivo* to block the red cell receptors for the parasite. This might be useful in the management of patients with particularly severe attacks of *P. falciparum* malaria, in whom the level of parasitemia may be very high. In the same type of patients, but not simultaneously, passively administered anti-idiotypic antibodies may be useful by directly binding to and destroying the parasites.

The present invention also provides a method for the detection of the presence of *P. falciparum* infection in a patient. The method employs insolubilized monoclonal antibody which identifies Band 3 and labeled, e.g., radiolabeled or enzyme labeled, monoclonal anti-idiotypic antibody to the aforesaid monoclonal antibody. Soluble *P. falciparum* in the test sample will interfere with the binding to the insolubilized monoclonal antibody of the labeled monoclonal anti-idiotypic antibody and will thus decrease the amount of the detectable label, e.g., the radioactivity or the enzyme, bound by the insolubilized antibody.

Non-limiting examples of supports for affinity-separation of antibodies, including monoclonals, include the following: activated Sepharose, activated cellulose and activated Sephadex. "Activated" refers to the creation, on the insoluble material, of reactive chemical groups that will form covalent linkages with the antibody molecules when incubated together under appropriate conditions. Typically, reactive groups are introduced into the insoluble substrate by the action of cyanogen bromide (CNBr) at high pH.

Significantly, as is well-known in the art, only a small portion of an antibody molecule, the paratope, is involved in the binding of the antibody to its epitope (see,

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in general, Clark, W.R. (1986) The Experimental Foundations of Modern Immunology Wiley & Sons, Inc., New York; Roitt, I. (1991) Essential Immunology, 7th Ed., Blackwell Scientific Publications, Oxford). The pFc' and Fc regions, for example, are effectors of the complement cascade but are not involved in antigen binding. An antibody from which the pFc' region has been enzymatically cleaved, or which has been produced without the pFc' region, designated an F(ab')₂ fragment, retains both of the antigen binding sites of an intact antibody. Similarly, an antibody from which the Fc region has been enzymatically cleaved, or which has been produced without the Fc region, designated an Fab fragment, retains one of the antigen binding sites of an intact antibody molecule. Fab fragments consist of a covalently bound antibody light chain and a portion of the antibody heavy chain denoted Fd. The Fd fragments are the major determinant of antibody specificity (a single Fd fragment may be associated with up to ten different light chains without altering antibody specificity) and Fd fragments retain epitope-binding ability in isolation.

Within the antigen-binding portion of an antibody, there are complementary determining regions (CDRs), which directly interact with the epitope of the antigen, and framework regions (FRs), which maintain the tertiary structure of the paratope (see, in general, Clark, 1986; Roitt, 1991). In both the heavy chain Fd fragment and the light chain of IgG immunoglobulins, there are four framework regions (FR1 through FR4) separated respectively by three complementarity determining regions (CDR1 through CDR3). The CDRs, and in particular the CDR3 regions, and more particularly the heavy chain CDR3, are largely responsible for antibody specificity.

It is now well-established in the art that the non-CDR regions of a mammalian antibody may be replaced with similar regions of conspecific or heterospecific antibodies while retaining the epitopic specificity of the original antibody. This is most clearly manifested in the development and use of "humanized" antibodies in which non-human CDRs are covalently joined to human FR and/or Fc/pFc' regions to produce a functional antibody. See, e.g., U.S. patents 4,816,567, 5,225,539, 5,585,089, 5,693,762 and 5,859,205. Accordingly, humanized anti-Band 3 antibodies and the use of such antibodies (e.g., to provide passive immunity to a subject) are embraced with the inventions disclosed herein.

For example, PCT International Publication Number WO 92/04381 teaches the production and use of humanized murine RSV antibodies in which at least a portion of the murine FR regions have been replaced by FR regions of human origin. Such antibodies, including fragments of intact antibodies with antigen-binding ability,
5 are often referred to as "chimeric" antibodies.

Thus, the present invention also provides for F(ab')₂, Fab, Fv and Fd fragments; chimeric antibodies in which the Fc and/or FR and/or CDR1 and/or CDR2 and/or light chain CDR3 regions have been replaced by homologous human or non-human sequences; chimeric F(ab')₂ fragment antibodies in which the FR and/or CDR1
10 and/or CDR2 and/or light chain CDR3 regions have been replaced by homologous human or non-human sequences; chimeric Fab fragment antibodies in which the FR and/or CDR1 and/or CDR2 and/or light chain CDR3 regions have been replaced by homologous human or non-human sequences; and chimeric Fd fragment antibodies in which the FR and/or CDR1 and/or CDR2 regions have been replaced by homologous
15 human or non-human sequences. The present invention also includes so-called single chain antibodies and human monoclonal antibodies, such as those produced by mice having functional human immunoglobulin gene loci.

Such antibodies also may be used to identify tissues expressing protein or to purify protein. Antibodies, particularly the anti-idiotypic antibodies of the invention,
20 also may be coupled to specific labeling agents for imaging or to anti-infectious agents, toxins such as ricin, other cytostatic or cytolytic drugs, and so forth, for therapeutic purposes.

As part of certain immunization compositions, one or more anti-Band 3 antibodies or stimulatory fragments thereof are administered with one or more
25 adjuvants to induce an immune response or to increase an immune response. An adjuvant is a substance incorporated into or administered with antigen which potentiates the immune response. Adjuvants may enhance the immunological response by providing a reservoir of antigen (extracellularly or within macrophages), activating macrophages and stimulating specific sets of lymphocytes. Adjuvants of
30 many kinds are well known in the art. Specific examples of adjuvants include monophosphoryl lipid A (MPL, SmithKline Beecham), a congener obtained after purification and acid hydrolysis of *Salmonella minnesota* Re 595 lipopolysaccharide;

saponins including QS21 (SmithKline Beecham), a pure QA-21 saponin purified from *Quillaja saponaria* extract; DQS21, described in PCT application WO96/33739 (SmithKline Beecham); QS-7, QS-17, QS-18, and QS-L1 (So et al., *Mol. Cells* 7:178-186, 1997); incomplete Freund's adjuvant; complete Freund's adjuvant; 5 montanide; immunostimulatory oligonucleotides (see e.g. CpG oligonucleotides described by Kreig et al., *Nature* 374:546-9, 1995); and various water-in-oil emulsions prepared from biodegradable oils such as squalene and/or tocopherol. Other adjuvants are known in the art and can be used in the invention (see, e.g. Goding, *Monoclonal Antibodies: Principles and Practice*, 2nd Ed., 1986). Methods 10 for the preparation of mixtures or emulsions of peptide and adjuvant are well known to those of skill in the art of vaccination.

Other agents which stimulate the immune response of the subject can also be administered to the subject. For example, other cytokines are also useful in vaccination protocols as a result of their lymphocyte regulatory properties. Many 15 other cytokines useful for such purposes will be known to one of ordinary skill in the art, including interleukin-12 (IL-12) which has been shown to enhance the protective effects of vaccines (see, e.g., *Science* 268: 1432-1434, 1995), GM-CSF and IL-18. Thus cytokines can be administered in conjunction with antigens and adjuvants to increase the immune response to the antigens.

20 There are a number of additional immune response potentiating compounds that can be used in vaccination protocols. These include costimulatory molecules provided in either protein or nucleic acid form. Such costimulatory molecules include the B7-1 and B7-2 (CD80 and CD86 respectively) molecules which are expressed on dendritic cells (DC) and interact with the CD28 molecule expressed on the T cell. 25 This interaction provides costimulation (signal 2) to an antigen/MHC/TCR stimulated (signal 1) T cell, increasing T cell proliferation and effector function. B7 also interacts with CTLA4 (CD152) on T cells and studies involving CTLA4 and B7 ligands indicate that the B7-CTLA4 interaction can enhance antitumor immunity and CTL proliferation (Zheng et al., *Proc. Nat'l Acad. Sci. USA* 95:6284-6289, 1998).

30 The invention also contemplates gene therapy. The procedure for performing *ex vivo* gene therapy is outlined in U.S. Patent 5,399,346 and in exhibits submitted in the file history of that patent, all of which are publicly available documents. In

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general, it involves introduction *in vitro* of a functional copy of a gene into a cell(s) of a subject which contains a defective copy of the gene, and returning the genetically engineered cell(s) to the subject. The functional copy of the gene is under operable control of regulatory elements which permit expression of the gene in the genetically engineered cell(s). Numerous transfection and transduction techniques as well as appropriate expression vectors are well known to those of ordinary skill in the art, some of which are described in PCT application WO95/00654. *In vivo* gene therapy using vectors such as adenovirus also is contemplated according to the invention.

Where it is desired to stimulate an immune response using a therapeutic composition of the invention, this may involve the stimulation of a humoral antibody response resulting in an increase in antibody titer in serum, a clonal expansion of cytotoxic lymphocytes, or some other desirable immunologic response. It is believed that doses of immunogens ranging from one nanogram/kilogram to 100 milligrams/kilogram, depending upon the mode of administration, would be effective. The preferred range is believed to be between 500 nanograms and 500 micrograms per kilogram. The absolute amount will depend upon a variety of factors, including the material selected for administration, whether the administration is in single or multiple doses, and individual patient parameters including age, physical condition, size, weight, and the stage of the disease. These factors are well known to those of ordinary skill in the art and can be addressed with no more than routine experimentation.

In addition to the above-described inventions which are based, in part, on the discovery of the particular Band 3 sequences which interact with MSP1 (or other malaria polypeptides of the invention), Applicants disclose herein related inventions which are based, in part, on the discovery of the particular portions of MSP1 (or other malaria polypeptides of the invention) that selectively binds to Band 3 protein. It is to be understood that enablement of the MSP1 (or other malaria polypeptides of the invention) related inventions throughout a broad scope can be accomplished in an analogous manner to that described for the Band 3 protein-related invention. Thus, the methods and definitions applied above in reference to the Band 3 molecule compositions and methods can be used in reference to the MSP1 molecule (or other malaria polypeptides of the invention) compositions and methods by substituting the

MSP1 molecule (or other malaria polypeptides of the invention) for the Band 3 molecule. For example, vectors expressing an MSP1 protein can be prepared by substituting an MSP1 nucleic acid for a Band 3 nucleic acid and inserting into an expression vector as described above. Thus, the invention embraces various
5 compositions containing such MSP-1 peptides for use, e.g., in screening assays to detect the specific interaction between MSP-1 and Band 3 protein, as well as for use in diagnostic and therapeutic applications for detecting and treating, respectively, malaria infection. In general, such compositions contain components (or are contained in kits which contain additional components) which are selected to detect
10 the specific interaction between MSP-1 (or other malaria polypeptides of the invention) and Band 3 protein (particularly, the Band 3 peptides disclosed herein).

According to one aspect of the invention, an isolated MSP1 peptide is provided. The peptide has an amino acid sequence selected from the group consisting of (Wellcome strain) SEQ ID NO:11 (MSP1(42)), SEQ ID NO:12 (MSP1(38), and
15 SEQ ID NO:13 (MSP1(38)-N-terminal domain) and (FCB1 strain) MSP1₃₈ (SEQ ID NO:33), MSP1₄₂ (SEQ ID NO:34), and MSP1₁₉ (SEQ ID NO:35), and unique fragments thereof which bind Band 3 protein. The sequences for peptides SEQ ID NOs:11-13 are based on *P. falciparum* Wellcome strain: GenBank Accession no.X02919 as follows:

20 SEQ ID NO:11 (MSP1(42)) refers to X02919 amino acids 1262 to 1639, inclusive;

SEQ ID NO:12 (MSP1(38)) refers to X02919 amino acids 902 to 1261, inclusive; and

25 SEQ ID NO:13 (MSP1(38) – N terminal region) refers to X02919 amino acids 902 to 1121, inclusive.

The sequences for peptides SEQ ID Nos:33-35 are based on *P. falciparum* FCB1 strain. The sequence of polypeptide SEQ ID NO:33 is deposited as GenBank Accession No.:AF286876, amino acids 911-1263. The sequence for polypeptide SEQ ID NO:34 is deposited as GenBank Accession No: AF325919; amino acids 1264-
30 1639. The sequence for polypeptide SEQ ID NO:35 is the C-terminal domain of MSP1(42); amino acids 1526-1639.

Such peptides may be contained in kits which detect the selective binding of the MSP1 peptide to a Band 3 protein, particularly to the Band 3 peptides disclosed herein.

According to another aspect of the invention, isolated nucleic acids encoding the foregoing MSP1 (or other malaria polypeptides of the invention) peptides are provided. The MSP1 nucleic acids have nucleotide sequences selected from the group consisting of (Wellcome strain) SEQ ID NO:54 (MSP1(42)), SEQ ID NO:55 (MSP1(38), and SEQ ID NO:56 (MSP1(38)-N-terminal domain) and (FCB1 strain) MSP1₃₈ (SEQ ID NO:57), MSP1₄₂ (SEQ ID NO:58), and MSP1₁₉ (SEQ ID NO:59), and unique fragments thereof. The sequences for nucleotide SEQ ID NOs:54-56 are based on *P. falciparum* Wellcome strain: GenBank Accession no.X02919. The sequences for nucleic acids SEQ ID Nos:57-59 are based on *P. falciparum* FCB1 strain, and are included in the nucleic acid sequences deposited as GenBank Accession Nos. AF286876 and AF325919

In certain embodiments, the nucleic acids comprise a unique fragment of the nucleotide sequence encoding the MSP1 peptides (or other malaria polypeptides of the invention) that selectively bind to Band 3. It is to be understood that all of the definitions described above in reference to the Band 3 molecules also apply to the MSP1 molecules (or other malaria polypeptides of the invention) of the invention. Thus, for example, the same definitions described above for "unique fragment", "isolated", "expression vectors", "alternate codons", and "operably joined" in reference to the Band 3 molecules of the invention apply to the MSP1 molecules (or other malaria polypeptides of the invention) of the invention. The expression vectors include the isolated foregoing MSP1 (or other malaria polypeptides of the invention) nucleic acids operably linked to a promoter. In another aspect of the invention, host cells transfected or transformed with the foregoing MSP1 (or other malaria polypeptides of the invention) nucleic acids or expression vectors are provided.

According to another aspect of the invention, compositions comprising the foregoing MSP1 (or other malaria polypeptides of the invention) peptides or nucleic acids and methods for making same are provided. The compositions are useful for inducing an immune response and include, e.g., immunogenic compositions. In general, the immunogenic compositions of the invention comprise one or more of the

foregoing isolated MSP1 (or other malaria polypeptides of the invention) polypeptides, a pharmaceutically acceptable carrier; and, optionally, an adjuvant, wherein the polypeptides are present in the composition in an effective amount to induce an immune system response. Antibodies to MSP1 peptides can be used, for example, in kits to identify agents which competitively bind to MSP1 and inhibit MSP1 binding to Band 3 protein or peptides as disclosed herein. According to a related aspect, a method of making a medicament (including, e.g., an immunogenic composition) is provided. The method involves placing one or more of the foregoing isolated MSP1 (or other malaria polypeptides of the invention) polypeptides or nucleic acids in a pharmaceutically acceptable carrier to form the medicament.

According to yet another aspect of the invention, a protein microarray comprising at least one isolated MSP-1 peptide selected from the group consisting of SEQ ID NOS. 11, 12, 13, 33, 34, and 35 is provided. Alternatively, a nucleic acid microarray comprising at least one nucleic acid encoding any of the foregoing MSP-1 peptides is provided. The arrays are useful in identifying agents which selectively bind to MSP1 molecules and inhibit MSP1 binding to Band 3 protein or peptides as disclosed herein.

According to still another embodiment, a method for identifying a candidate mimetic of an isolated MSP1 peptide is provided. The method involves: providing a Band 3 molecule (e.g., the above-described isolated Band 3 peptides) which binds an isolated MSP1 peptide (e.g., an MSP1 peptide having SEQ ID NOS:11, 12, 13, 33, 34, and/or 35); contacting the Band 3 molecule with the MSP1 peptide in the presence or absence of a test molecule; and determining the binding of the test molecule to the Band 3 molecule, wherein a test molecule which binds to the Band 3 molecule and inhibits binding of the Band 3 molecule with the MSP1 peptide is a candidate mimetic of the isolated peptide.

According to still another aspect of the invention, an antibody (or fragment thereof) (referred to herein as a "anti-MSP1 antibody") that: (1) selectively binds to any of the above-described peptides, particularly the peptides having a sequence selected from the group consisting of SEQ ID Nos. 11, 12, 13, 33, 34, and 35; and (2) inhibits binding of the MSP1 peptide to Band 3 protein (particularly the Band 3

peptides disclosed herein) is provided. Preferably, the antibody is a monoclonal antibody, and, more preferably, a humanized monoclonal antibody.

In addition to the inventions which are based, in part, on the discovery of the particular Band 3 sequences which interact with MSP1, Applicants disclose herein
5 related inventions which are based, in part, on the discovery that Band 3 sequences interact with additional *Plasmodium* polypeptide sequences comprising the amino acid sequences selected from the group consisting of SEQ ID NOs:46-53. These polypeptides are encoded by the nucleic acid molecules comprising a nucleotide sequence selected from the group consisting of SEQ ID NOs:38-45. It is to be
10 understood that enablement of the BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and ABRA-related inventions throughout a broad scope can be accomplished in an analogous manner to that described for the Band 3 protein-related invention and the MSP1 polypeptide-related invention. Thus, the methods and definitions applied above in reference to the Band 3 and MSP-1 molecule
15 compositions and methods can be used in reference to the BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA molecule compositions and methods by substituting the BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA molecule for the MSP-1 molecule. For example, vectors expressing an BBP-1 polypeptide can be prepared by substituting a BBP-1 nucleic acid for a MSP-1 nucleic
20 acid and inserting into an expression vector as described above. Thus, the invention embraces various compositions containing such BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA polypeptides for use, e.g., in screening assays to detect the specific interaction between BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA and Band 3 protein, as well as for use in diagnostic (e.g.
25 detection of the malaria polypeptides of the invention in biological samples obtained from a subject using conventional assays, such as an antibody-based assays or nucleic acid hybridization-based assays) and therapeutic applications (e.g., vaccines) for detecting and treating, respectively, malaria infection. The sequence identification for these polypeptides is provided in Table 1. In general, such compositions contain
30 components (or are contained in kits which contain additional components) which are selected to detect the specific interaction between BBP-1, BBP-2, BBP-3, BBP-4,

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BBP-5, BBP-6, RhopH3, and/or ABRA, and Band 3 protein (particularly, the Band 3 peptides disclosed herein).

Table 1. Band 3 Binding Polypeptide Identification

Polypeptide Name	Nucleic Acid SEQ ID NO.	Amino Acid SEQ ID NO.
BBP-1	38	46
BBP-2	39	47
BBP-3	40	48
BBP-4	41	49
BBP-5	42	50
BBP-6	43	51
RhopH3	44	52
ABRA	45	53

5

According to one aspect of the invention, an isolated BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA peptide is provided. The peptide has an amino acid sequence selected from the group consisting of SEQ ID NOs:46-53 and unique fragments thereof that bind Band 3 protein. The sequences for these peptides are presented herein.

10

Such peptides may be contained in kits which detect the selective binding of the BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA polypeptide to a Band 3 protein, particularly to the Band 3 peptides disclosed herein.

According to another aspect of the invention, isolated nucleic acids encoding the foregoing BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA polypeptides are provided. In certain embodiments, the nucleic acids comprise a unique fragment of the nucleotide sequence encoding the BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA polypeptides that selectively bind to Band 3 peptides. It is to be understood that all of the definitions described above in reference to the Band 3 and MSP-1 molecules also apply to the BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA molecules of the invention. Thus, for example, the same definitions described above for "unique fragment", "isolated",

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“expression vectors”, “alternate codons”, and “operably joined” in reference to the Band 3 and MSP-1 molecules of the invention apply to the BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA molecules of the invention. The expression vectors include the isolated foregoing BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA nucleic acids operably linked to a promoter. In another aspect of the invention, host cells transfected or transformed with the foregoing BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA nucleic acids or expression vectors are provided.

According to another aspect of the invention, compositions comprising the foregoing BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA polypeptides or nucleic acids and methods for making same are provided. The compositions are useful for inducing an immune response and include, e.g., immunogenic compositions. In general, the immunogenic compositions of the invention comprise one or more of the foregoing isolated BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and ABRA polypeptides, a pharmaceutically acceptable carrier; and, optionally, an adjuvant, wherein the polypeptides are present in the composition in an effective amount to induce an immune system response. Antibodies to BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and ABRA polypeptides can be used, for example, in kits to identify agents which competitively bind to BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA and inhibit BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA binding to Band 3 protein or peptides as disclosed herein. According to a related aspect, a method of making a medicament (including, e.g., an immunogenic composition) is provided. The method involves placing one or more of the foregoing isolated BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA polypeptides or nucleic acids in a pharmaceutically acceptable carrier to form the medicament.

According to yet another aspect of the invention, a protein microarray comprising at least one isolated BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA polypeptide selected from the group consisting of SEQ ID NOs:46-53 is provided. Alternatively, a nucleic acid microarray comprising at least one nucleic acid encoding any of the foregoing BBP-1, BBP-2, BBP-3, BBP-4, BBP-

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5, BBP-6, RhopH3, and/or ABRA polypeptides is provided. The arrays are useful in identifying agents which selectively bind to BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA molecules and inhibit BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA binding to Band 3 protein or peptides as disclosed herein.

According to still another embodiment, a method for identifying a candidate mimetic of an isolated BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA polypeptide is provided. The method involves: providing a Band 3 molecule (e.g., the above-described isolated Band 3 peptides) which binds an isolated BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA polypeptide (e.g., a BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA polypeptide having SEQ ID NOs:46-53); contacting the Band 3 molecule with the BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA polypeptide in the presence or absence of a test molecule; and determining the binding of the test molecule to the Band 3 molecule, wherein a test molecule which binds to the Band 3 molecule and inhibits binding of the Band 3 molecule with the isolated peptide is a candidate mimetic of the isolated peptide.

According to still another aspect of the invention, an antibody (or fragment thereof) (referred to herein as a "anti-BBP-1, anti-BBP-2, anti-BBP-3, anti-BBP-4, anti-BBP-5, anti-BBP-6, anti-RhopH3, and/or anti-ABRA antibody") that: (1) selectively binds to any of the above-described peptides, particularly the peptides having an amino acid sequence selected from the group consisting of SEQ ID Nos. 46-53; and (2) inhibits binding of the BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA polypeptide to Band 3 protein (particularly the Band 3 peptides disclosed herein) is provided. Preferably, the antibody is a monoclonal antibody, and, more preferably, a humanized monoclonal antibody.

The present invention also, in some aspects, involves the identification of cDNAs that encode *Plasmodium* polypeptides BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, and BBP-6. The sequence of the coding portion of the *Plasmodium* gene for each is presented as SEQ ID NOs:38-43, and the predicted amino acid sequences of these genes' protein products are presented as SEQ ID NOs:46-51.

The invention thus involves in one aspect *Plasmodium* BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, and BBP-6 polypeptides, nucleic acid molecules encoding those proteins, functional modifications and variants of the foregoing, useful fragments of the foregoing, as well as therapeutic and diagnostic products (including antibodies),
5 non-human animal models, and methods relating thereto.

According to one aspect of the invention, isolated nucleic acid molecules are provided. The isolated nucleic acid molecule is selected from the group consisting of:
An isolated nucleic acid molecule selected from the group consisting of:

- (a) nucleic acid molecules which hybridize under stringent conditions to a
10 nucleic acid molecule comprising a nucleotide sequence selected from the group consisting of SEQ ID NOs:38-43 and which codes for a *Plasmodium* polypeptide,
- (b) deletions, additions and substitutions of the nucleic acid molecules of (a), which code for a *Plasmodium* polypeptide,
- (c) nucleic acid molecules that differ from the nucleic acid molecules of (a) or
15 (b) in codon sequence due to the degeneracy of the genetic code, and
- (d) complements of (a), (b) or (c).

The preferred isolated nucleic acids of the invention are *Plasmodium* nucleic acid molecules which encode a *Plasmodium* polypeptide. As used herein, a
20 *Plasmodium* polypeptide refers to a protein that is encoded by a nucleic acid having SEQ ID NOs:38-45 and 54-59 or a functional fragment thereof, or a functional equivalent thereof (e.g., a nucleic acid sequence encoding the same protein as encoded by SEQ ID NOs:38-45 and 54-59), provided that the functional fragment or equivalent encodes a protein which exhibits a *Plasmodium* polypeptide functional
25 activity. As used herein, a *Plasmodium* functional activity refers to the ability of a *Plasmodium* polypeptide of the invention to interact with a Band 3 molecule of the invention.

In the preferred embodiments, the isolated nucleic acid molecule is selected from the group consisting of SEQ ID NOs:38-45 and 54-59.

30 The invention provides isolated nucleic acid molecules which code for *Plasmodium* proteins and which hybridize under stringent conditions to a nucleic acid molecule consisting of the nucleotide selected from the group consisting of SEQ ID

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NOs:38-45 and 54-59. Such nucleic acids may be DNA, RNA, composed of mixed deoxyribonucleotides and ribonucleotides, or may incorporate synthetic non-natural nucleotides. Various methods for determining the expression of a nucleic acid and/or a polypeptide in cells are known to those of skill in the art and are described further below. As used herein, the term protein is meant to include large molecular weight proteins and peptides and low molecular weight peptides or fragments thereof.

The term "stringent conditions" as used herein refers to parameters with which the art is familiar. Nucleic acid hybridization parameters may be found in references which compile such methods, e.g. *Molecular Cloning: A Laboratory Manual*, J. Sambrook, et al., eds., Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 1989, or *Current Protocols in Molecular Biology*, F.M. Ausubel, et al., eds., John Wiley & Sons, Inc., New York. More specifically, stringent conditions, as used herein, refers, for example, to hybridization at 65°C in hybridization buffer (3.5 x SSC, 0.02% Ficoll, 0.02% polyvinyl pyrrolidone, 0.02% Bovine Serum Albumin (BSA), 2.5mM NaH₂PO₄ (pH 7), 0.5% SDS, 2mM EDTA). SSC is 0.15M sodium chloride/0.15M sodium citrate, pH 7; SDS is sodium dodecyl sulphate; and EDTA is ethylenediaminetetracetic acid. After hybridization, the membrane upon which the DNA is transferred is washed at 2x SSC at room temperature, and then at 0.1x SSC/0.1 x SDS at temperatures up to 68°C.

The foregoing set of hybridization conditions is but one example of stringent hybridization conditions known to one of ordinary skill in the art. There are other conditions, reagents, and so forth which can be used, which result in a stringent hybridization. The skilled artisan will be familiar with such conditions, and thus they are not given here. It will be understood, however, that the skilled artisan will be able to manipulate the conditions in a manner to permit the clear identification of homologs and alleles of *Plasmodium* nucleic acid molecules and Band 3 nucleic acid molecules of the invention. The skilled artisan also is familiar with the methodology for screening cells and libraries for expression of such molecules which then are routinely isolated, followed by isolation of the pertinent nucleic acid molecule and sequencing.

In general homologs and alleles of the malaria polypeptides typically will share at least 40% nucleotide identity and/or at least 50% amino acid identity to SEQ

ID NOs:38-45 and 54-59 and SEQ ID NOs:46-53, 11-13, and 33-35, respectively, in some instances will share at least 50% nucleotide identity and/or at least 65% amino acid identity and in still other instances will share at least 60% nucleotide identity and/or at least 75% amino acid identity. Preferred homologs and alleles share
5 nucleotide and amino acid identities with SEQ ID NOs:38-45, 54-59 and SEQ ID NOs:46-53, 11-13, and 33-35, respectively, and encode polypeptides of greater than 80%, more preferably greater than 90%, still more preferably greater than 95% and most preferably greater than 99% identity. The percent identity can be calculated using various publicly available software tools developed by NCBI (Bethesda,
10 Maryland) that can be obtained through the internet (<ftp://ncbi.nlm.nih.gov/pub/>). Exemplary tools include the BLAST system available at <http://www.ncbi.nlm.nih.gov>, which uses algorithms developed by Altschul et al. (*Nucleic Acids Res.* 25:3389-3402, 1997). Pairwise and ClustalW alignments (BLOSUM30 matrix setting) as well as Kyte-Doolittle hydropathic analysis can be
15 obtained using the MacVector sequence analysis software (Oxford Molecular Group). Watson-Crick complements of the foregoing nucleic acid molecules also are embraced by the invention.

In screening for *Plasmodium* genes, a Southern blot may be performed using the foregoing conditions, together with a detectably labeled probe (e.g. radioactive or
20 chemiluminescent probes). After washing the membrane to which the DNA is finally transferred, the membrane can be placed against X-ray film or a phosphorimager to detect the radioactive or chemiluminescent signal. In screening for the expression of *Plasmodium* RNA, Northern blot hybridizations using the foregoing conditions can be performed on samples taken from cells or subjects suspected of having a *Plasmodium*
25 infection. Amplification protocols such as PCR using primers that hybridize to the sequences presented also can be used for detection of the *Plasmodium* genes or expression thereof.

Identification of related sequences can be achieved using PCR and other amplification techniques suitable for cloning related nucleic acid sequences.
30 Preferably, PCR primers are selected to amplify portions of a nucleic acid sequence believed to be conserved (e.g., a binding domain, etc.). Again, nucleic acids are preferably amplified from a *Plasmodium* library.

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The invention also includes degenerate nucleic acid molecules which include alternative codons to those present in the native materials. For example, serine residues are encoded by the codons TCA, AGT, TCC, TCG, TCT and AGC. Each of the six codons is equivalent for the purposes of encoding a serine residue. Thus, it will be apparent to one of ordinary skill in the art that any of the serine-encoding nucleotide triplets may be employed to direct the protein synthesis apparatus, *in vitro* or *in vivo*, to incorporate a serine residue into an elongating *Plasmodium* polypeptide. Similarly, nucleotide sequence triplets that encode other amino acid residues include, but are not limited to: CCA, CCC, CCG and CCT (proline codons); CGA, CGC, CGG, CGT, AGA and AGG (arginine codons); ACA, ACC, ACG and ACT (threonine codons); AAC and AAT (asparagine codons); and ATA, ATC and ATT (isoleucine codons). Other amino acid residues may be encoded similarly by multiple nucleotide sequences. Thus, the invention embraces degenerate nucleic acids that differ from the biologically isolated nucleic acids in codon sequence due to the degeneracy of the genetic code.

According to another aspect of the invention, further isolated nucleic acid molecules that are based on the above-noted *Plasmodium* nucleic acid molecules are provided. In this aspect, the isolated nucleic acid molecules are selected from the group consisting of:

- (a) a unique fragment of the nucleotide sequence selected from the group consisting of:
- nucleotides 1-1287 of SEQ ID NO:38 between 12 and 1286 nucleotides in length,
 - nucleotides 1-3576 of SEQ ID NO:39 between 12 and 3557 nucleotides in length,
 - nucleotides 1-903 of SEQ ID NO:40 between 12 and 902 nucleotides in length,
 - nucleotides 1-1203 of SEQ ID NO:41 between 12 and 1202 nucleotides in length,
 - nucleotides 1-3996 of SEQ ID NO:42 between 12 and 3995 nucleotides in length, and
 - nucleotides 1-876 of SEQ ID NO:43 between 12 and 875 nucleotides in length, and
 - nucleotides 1-2712 of SEQ ID NO:44 between 12 and 2711 nucleotides in length, and
 - nucleotides 1-2232 of SEQ ID NO:45 between 12 and 2231 nucleotides in length,
 - and nucleotides 1-1134 of SEQ ID NO:54 between 12 and 1133 nucleotides in length,
 - and nucleotides 1-1080 of SEQ ID NO:55 between 12 and 1079 nucleotides in length,
 - and nucleotides 1-660 of SEQ ID NO:56 between 12 and 659 nucleotides in length,

and nucleotides 1-1080 of SEQ ID NO:57 between 12 and 1079 nucleotides in length,
and nucleotides 1-1131 of SEQ ID NO:58 between 12 and 1130 nucleotides in length,
and nucleotides 1-343 of SEQ ID NO:59 between 12 and 342 nucleotides in length,
and

5 (b) complements of (a),

wherein the unique fragments exclude nucleic acids having nucleotide
sequences that are contained within SEQ ID NOs:38-45, 54-59, and that are known as
of the filing date of this application.

The invention also provides isolated unique fragments of SEQ ID NOs:38-44
10 or complements of SEQ ID NOs:38-45, 54-59. A unique fragment is one that is a
'signature' for the larger nucleic acid. It, for example, is long enough to assure that
its precise sequence is not found in molecules outside of the *Plasmodium* nucleic acid
molecules defined above. Those of ordinary skill in the art may apply no more than
routine procedures to determine if a fragment is unique within the *Plasmodium*.

15 Unique fragments, however, exclude fragments completely composed of the
nucleotide sequences that are contained within SEQ ID NO:38-45, 54-59 and that are
known as of the filing date of this application.

Unique fragments can be used as probes in Southern blot, Northern blot, and
Gene Chip/microarray assays to identify such nucleic acid molecules, or can be used
20 in amplification assays such as those employing PCR. As known to those skilled in
the art, large probes such as 200 nucleotides or more are preferred for certain uses
such as Southern blots, while smaller fragments will be preferred for uses such as in
PCR and Gene Chip/microarray assays. Unique fragments also can be used to
produce fusion proteins for generating antibodies or determining binding of the
25 polypeptide fragments, or for generating immunoassay components. Likewise,
unique fragments can be employed to produce nonfused fragments of the *Plasmodium*
polypeptides that are useful, for example, in the preparation of antibodies in
immunoassays. Unique fragments further can be used as antisense molecules to
inhibit the expression of *Plasmodium* nucleic acids and polypeptides, particularly for
30 therapeutic purposes as described in greater detail below.

As will be recognized by those skilled in the art, the size of the unique
fragment will depend upon its conservancy in the genetic code. Thus, some regions

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of SEQ ID NOs:38-44, 54-59 and their complements will require longer segments to be unique while others will require only short segments, typically between 12 and 32 nucleotides or more in length (e.g. 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, and 32 or more), up to the entire length of the disclosed
5 sequence. Many segments of the polynucleotide coding region or complements thereof that are 18 or more nucleotides in length will be unique. Those skilled in the art are well versed in methods for selecting such sequences, typically on the basis of the ability of the unique fragment to selectively distinguish the sequence of interest from non-*Plasmodium* nucleic acid molecules. A comparison of the sequence of the
10 fragment to those on known data bases typically is all that is necessary, although *in vitro* confirmatory hybridization and sequencing analysis may be performed.

A unique fragment can be a functional fragment. A functional fragment of a nucleic acid molecule of the invention is a fragment which retains some functional property of the larger nucleic acid molecule, such as coding for a functional
15 polypeptide, binding to proteins, regulating transcription of operably linked nucleic acid molecules, and the like. One of ordinary skill in the art can readily determine using the assays described herein and those well known in the art to determine whether a fragment is a functional fragment of a nucleic acid molecule using no more than routine experimentation.

20 As used herein, the term "antisense oligonucleotide" or "antisense" describes an oligonucleotide that is an oligoribonucleotide, oligodeoxyribonucleotide, modified oligoribonucleotide, or modified oligodeoxyribonucleotide which hybridizes under physiological conditions to DNA comprising a particular gene or to a transcript of that gene and, thereby, inhibits the transcription of that gene and/or the translation of that
25 mRNA. The antisense molecules are designed so as to interfere with transcription or translation of a target gene upon hybridization with the target gene or transcript. Those skilled in the art will recognize that the exact length of the antisense oligonucleotide and its degree of complementarity with its target will depend upon the specific target selected, including the sequence of the target and the particular bases
30 which comprise that sequence. It is preferred that the antisense oligonucleotide be constructed and arranged so as to bind selectively with the target under physiological conditions, i.e., to hybridize substantially more to the target sequence than to any

other sequence in the target cell under physiological conditions. Based upon SEQ ID NOs:38-45, 54-59, or upon allelic or homologous genomic and/or cDNA sequences, one of skill in the art can easily choose and synthesize any of a number of appropriate antisense molecules for use in accordance with the present invention. In order to be
5 sufficiently selective and potent for inhibition, such antisense oligonucleotides should comprise at least 10 and, more preferably, at least 15 consecutive bases which are complementary to the target, although in certain cases modified oligonucleotides as short as 7 bases in length have been used successfully as antisense oligonucleotides (Wagner et al., *Nature Biotechnology* 14: 840-844, 1996).

10 Most preferably, the antisense oligonucleotides comprise a complementary sequence of 20-30 bases. Although oligonucleotides may be chosen which are antisense to any region of the gene or its transcripts, in preferred embodiments the antisense oligonucleotides correspond to N-terminal or 5' upstream sites such as translation initiation, transcription initiation, or promoter sites. In addition, 3'-
15 untranslated regions may be targeted. Targeting to mRNA splicing sites also has been used in the art but may be less preferred because alternative mRNA splicing of the *Plasmodium* transcript occurs. In addition, the antisense is targeted, preferably, to sites in which mRNA secondary structure is not expected (see, e.g., Sainio et al., *Cell Mol. Neurobiol.* 14(5):439-457, 1994) and at which proteins are not expected to bind.
20 The present invention also provides for antisense oligonucleotides which are complementary to genomic DNA and/or cDNA corresponding to SEQ ID NO:38, SEQ ID NO:39, SEQ ID NO:40, SEQ ID NO:41, SEQ ID NO:42, SEQ ID NO:43, SEQ ID NO:44, SEQ ID NO:45, SEQ ID NO:54, SEQ ID NO:55, SEQ ID NO:56, SEQ ID NO:57, SEQ ID NO:58, and/or SEQ ID NO:59. Antisense to allelic or
25 homologous cDNAs and genomic DNAs are enabled without undue experimentation.

In summary, the various aspects of the invention include one or more of the following utilities:

1. Treatment and prevention of malaria disease in human and animals:
30 (a) Development of a peptide, peptidomimetic, and/or protein antimalarial drugs partially or entirely based on the human Band 3 protein residues 720-761 and/or 807-826.

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- (b) Development of an antimalarial gene therapy using DNA plasmids encoding Band 3 protein sequence derived partially or entirely from the human Band 3 protein residues 720-761 and/or 807-826.
- (c) Development of a peptide, peptidomimetic, and/or protein antimalarial drug
5 using three dimensional structure information of human Band 3 protein containing a partial or the entire amino acid sequence of residues 720-761 and/or 807-826.
- (d) Development of a non-peptide, non-protein, and/or non-peptidomimetic antimalarial drug derived from three dimensional structure information of human Band 3 protein containing a partial or the entire amino acid sequence of residues 720-
10 761 and 807-826.
- (e) Development of an idiotype protein vaccine that produces anti-idiotypic antibodies mimicking the entire or partial structure of residues 720-761 and/or 807-826 of the human Band 3 protein, for use in malaria disease.
- (f) Development of an idiotype DNA vaccine in which DNA encodes the
15 idiotypic determinants to induce the production of such anti-idiotypic antibodies as defined above.
- (g) Use of a non-human erythrocyte Band 3 gene and/or protein sequence corresponding to the human Band 3 protein residues 720-761 and/or 807-826 for the purpose of developing drug or vaccine for human and/or animal malaria disease.
- 20 (h) Use of a gene and/or protein sequence corresponding to the MSP-1, BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, ABRA molecules of the invention for the purpose of developing drugs or vaccines for human and/or animal malaria disease and for malaria diagnostic purposes.
2. Screening of the malaria parasite ligand(s) binding to the host Band 3 receptor:
- 25 (a) Use of a peptide and/or protein containing partial or entire sequence of human Band 3 protein residues 720-761 and/or 807-826 in efforts to identify and/or develop drug or vaccine for human and/or animal malaria disease. These include experiments including, but not limited to: protein or peptide binding experiments carried out *in vitro* and *in vivo*; three-dimensional structure-based approaches; computer modeling;
30 combinatorial chemistry screening; other high throughput screening approaches. The malaria parasite ligand(s) identified and/or characterized by utilizing the inventions

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disclosed herein as new targets for the development of a highly efficient malaria vaccine and/or drug.

(b) Use of nucleotide sequence encoding partial or entire amino acid sequence of human Band 3 protein residues 720-761 and/or 807-826 to identify and/or functionally characterize the malaria parasite ligand(s) binding to the erythrocyte Band 3 protein.

(c) Use of a non-human erythrocyte Band 3 gene and/or protein sequence corresponding to the human Band 3 protein residues 720-761 and/or 807-826 for the purpose of carrying out the screening of malaria parasite ligand as described in (a) and (b) above.

3. Screening Assays to select agents which inhibit MSP1 binding to Band 3 protein, and screening assays to select agents that inhibit BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA binding to Band 3 protein.

(a) Use of a peptide and/or protein containing partial or entire sequence of MSP1, BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and/or ABRA polypeptide in combination with a peptide and/or protein containing human Band 3 protein (e.g., Band 3 residues 720-761 and/or 807-826) to identify and/or develop drug or vaccine for human and/or animal malaria disease. These include experiments including, but not limited to: protein or peptide binding experiments carried out *in vitro* and/or *in vivo*; three-dimensional structure-based approaches; computer modeling; combinatorial chemistry screening; other high throughput screening approaches. These screening assays can be used to detect lead molecules in mixtures (e.g., libraries) of synthetic or naturally-occurring molecules.

Examples

Introduction to the Examples:

To extend our observation that the erythrocyte Band 3 (-/-) mice are completely resistant to invasion by murine malaria parasite *Plasmodium yoelii* 17 XL, we have conducted a series of experiments to identify the key host receptor that mediates malaria parasite invasion into human red blood cells. Using a peptide scanning technique, we have identified two specific regions of the erythroid Band 3 protein (also known as Anion Exchanger 1 or AE1) that serve as the receptor for

malaria parasite invasion into the human erythrocytes. These two regions are located within ectoplasmic domains 5 and 6 of the Band 3 protein. In our study, the ectoplasmic domains 5 and 6 are defined as below based on the published human Band 3 amino acid sequence

5 Ectoplasmic domain 5 (amino acid residues 720-761): (SEQ ID NO:22)
GMPWLSATTVRSVTHANALTVMGKASTPGAAAQIQEVKEQRI

Ectoplasmic domain 6 (amino acid residues 807-857): (SEQ ID NO:23)
10 DRILLLFKPPKYHPDVPYVKRVKTWRMHLFTGIQIICLAVLWVVKSTPASL

Four peptides (B3_{5A}, B3_{5B}, B3_{5C}, B3_{6A}) derived from ectoplasmic domains 5 and 6 of the Band 3 protein inhibited the invasion of the most lethal *Plasmodium falciparum* malaria parasite into human erythrocytes *in vitro*, while other peptides used in the scanning experiment had no significant effect as compared to the control
15 sample. The peptides B3_{5C} and B3_{6A} showed highest inhibition amongst all peptides tested. The amino acid sequences of the four peptides are:

B3_{5A} (720-739): GMPWLSATTVRSVTHANALT (SEQ ID NO:1)
B3_{5B} (731-750): SVTHANALTVMGKASTPGAA (SEQ ID NO:2)
B3_{5C} (742-761): GKASTPGAAAQIQEVKEQRI (SEQ ID NO:3)
20 B3_{6A} (807-826): DRILLLFKPPKYHPDVPYVK (SEQ ID NO:4)

The blot overlay technique using the peptides B3_{5C} and B3_{6A} revealed that these peptides specifically bind to a set of human *P. falciparum* malaria proteins expressed at the merozoite stage. One of the proteins identified by the blot overlay
25 assay corresponds to the molecular mass of Merozoite Surface Protein-1 (MSP-1), a well known malaria parasite protein.

We have further studied the interaction between the Band 3 peptides and MSP1 and have identified sequences within MSP1 which selectively bind to the Band 3 receptor. These studies and sequences are described in detail below. We have also
30 examined the interaction between the Band 3 polypeptides and BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and ABRA polypeptides. These studies are described in more detail below. The amino acid sequences of the BBP-1, BBP-2, BBP-3, BBP-4, BBP-5, BBP-6, RhopH3, and ABRA polypeptides are SEQ ID
35 NOs:46-53. These polypeptides of the invention are encoded by the nucleic acid molecules comprising nucleotide sequences selected from the group consisting of SEQ ID NOs:38-45.

Based on our findings, we propose an parasite invasion pathway model illustrated in Figure 6.

EXAMPLE 1. Anti-Band 3 Antibody Preparation

5 Using the method of G. Kohler and C. Milstein, Nature 256, 495-496, (1975), a BALB/c mouse is immunized with washed human red cells expressing Band 3 or isolated Band 3 by weekly intraperitoneal administration of approximately 10^7 erythrocytes each or an equivalent amount of the isolated Band 3 protein. The spleen of the mouse is then removed and a cell suspension prepared in tissue culture medium
10 (RPMI-1640 with additional glutamine, 5mM). The spleen cell suspension is mixed with a suspension of the mouse myeloma cell line P3/NSO-Ag4-1 (NS-0) (obtained from the ATCC) which, being deficient in the enzyme hypoxanthine-guanine phosphoribosyltransferase (HGPRT), will not grow in tissue culture media containing hypoxanthine, aminopterin and thymidine (HAT media).

15 The mixture contains four spleen cells to one myeloma cell. Fusion is promoted by the addition of polyethylene glycol (PEG) of an average molecular weight of 2000. After fusion, the cells are washed free of PEG, resuspended in HAT medium and allowed to grow to a density of 10^6 live cells per ml and aliquots of 0.1 ml added to the wells of a 24-well tray containing feeder cells (from BALB/c
20 thymus).

 Partial changes of culture medium are performed at approximately 3, 5 and 7 days and the supernatants are removed approximately 14 days postfusion and tested for the presence of antibodies that bind to Band 3. Since the process is conducted in the presence of HAT, essentially non-fused myeloma cells are dead at this time,
25 which prevents them from possibly overgrowing the fused (hybrid) ones.

 The unfused spleen cells are also dead because of their very limited capacity to grow ex vivo in this tissue culture medium. The hybrid cells grow and multiply because the normal spleen cells contribute the enzyme HGPRT and the myelomatous cells for the capacity for indefinite proliferation. The supernatants from the wells
30 containing colonies of hybrid cells are assayed on a panel including isolated Band 3 (e.g., immobilized in microtiter plate wells). The cellular contents of the positive

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well(s) that contain antibodies to Band 3 are then recovered and suspended to a concentration of 3 cells per ml.

Aliquots of 0.1 ml are then added to fresh wells, so that on the average only one of every three wells receives a cell and, thus, the colonies that result from the growth of this very diluted suspension are likely to be true "clones", i.e., descendants from a single progenitor. When the colonies attained a size of 10^2 cells their supernatants are again screened for the presence of Band 3 antibodies and the most strongly positive ones are allowed to expand to a number of 10^5 to 10^6 . Dilution of these cell suspensions to a concentration of 3 cells/ml and plating volumes of 0.1 ml as before results in the growth of doubly-cloned hybrid, antibody-producing cell lines.

The concentration of monoclonal antibody at the time of maturity of a culture flask is $20 \pm 5 \mu\text{g/ml}$. The cloned hybridoma also grows *in vivo* in mice of the BALB/c strain or of its first generation (F1) hybrids. This growth is in the form of malignant myelomatous tumors. When live hybridoma cells are injected into susceptible animals, they secrete high concentrations of antibody into the peritoneal spaces. "Priming" the animals with irritants such as incomplete Freund's adjuvant or Pristane injected into the peritoneum, before grafting the hybridomas, results in the formation of large volumes of ascites containing antibody in concentrations higher than 5 mg/ml.

20

EXAMPLE 2. Anti-Band 3 Antibody Blocks Penetration of *P. falciparum* Merozoites into Erythrocytes.

The methods disclosed herein are based on those disclosed in U.S. patent no. 5,101,017, which reportedly are adapted from those described in L. H. Miller, S. J. Mason, J. A. Dvorak, T. Shiroishi and M. H. McGinnis, "Erythrocyte Receptors for Malarial Merozoites and the Duffy Blood Group System", Human Blood Groups, 5th International Convocation on Immunology, Buffalo, NY, 1976, Basel Karger, pp. 394-400, 1977.

Standard numbers of merozoites are incubated with standard numbers of erythrocytes. In parallel wells, red cells of primates of different species are exposed to *P. falciparum* in the presence of an Anti-Band 3 antibody or non-specific antibody as a control, e.g., anti-Rh29 and anti-K14 antibodies (where are reactive with

30

essentially all human and most primate red blood cells, but their antigenic epitopes are unrelated to Band 3). A third well for each erythrocyte donor contains only tissue culture medium instead of monoclonal antibodies. The two latter, control wells allow the determination of the proportion of red cells that are "normally" penetrated by the parasite under these conditions. Thus, comparison between this proportion and that in the well containing the anti-Band 3 monoclonal antibody permits estimation of its inhibitory effect

EXAMPLE 3. Specificity Determination.

The anti-Band 3 monoclonal antibody is tested for its capacity to immunoprecipitate or otherwise selectively bind to the Band 3 protein and/or Band 3 peptides of the invention (e.g., immobilized, labeled with a detectable agent). For example, red blood cell membrane proteins (including Band 3) or isolated Band 3 protein or peptides of the invention, separated by SDS-PAGE and blotted onto nitrocellulose filters, are exposed to labeled monoclonal antibody molecules and a single Band of the appropriate molecular size (of the Band 3 protein or peptides of the invention) and overall chemical characteristics is obtained.

EXAMPLE 4. Preparation of Monoclonal Anti-idiotypic Antibodies.

All antibody molecules are, at the same time, antigens since their ability to function as antibodies, i.e., to bind to antigen, depends on a special stereochemical configuration which is specific for each antibody and is called an "idiotypic". A monoclonal antibody immunoglobulin is constituted of exactly identical molecules, each having the same specific combining site, which, being complementary to the respective antigen, becomes antigenic for the antibody-producing host and to other animals of the same strain. In other words, the idiotypic of an antibody leads to the production of anti-idiotypic antibodies. This antigenic property can thus be used to elicit such anti-idiotypic antibodies by injecting naive hosts with purified monoclonal antibodies produced in animals of the same inbred strain.

This procedure is based on that described in U.S. patent no. 5,101,017. The preparation of anti-idiotypic antibodies is accomplished by first purifying the original monoclonal antibody (designated Abl) by affinity chromatography, emulsifying it in

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complete Freund's adjuvant and injecting this emulsion into the peritoneum and under the skin in multiple sites of BALB/c mice. A second, identical injection is given approximately two weeks later. Subsequent injections require the use of incomplete Freund's adjuvant. The schedule of these injections and the quantities of

5 immunoglobulin injected are empirical and different procedures have been reported to be successful (e.g., two further injections in complete Freund's can be given two weeks apart and following the initial two injections in complete Freund's adjuvant by two weeks). The recipient mice are rested for two months followed by two bi-weekly injections of the Anti-Band 3 antibody (Ab1). One week later, the spleens are

10 removed and fused with NSO myeloma cells. Hybridomas are grown as described elsewhere in this application and screening is performed by competitive inhibition of Ab1 binding to human red cells which express Band 3 or screening by competitive inhibition of Ab1 binding to isolated Band 3 or a Band 3 peptide of the invention. This inhibition test consists of adding the supernatants of hybridomas putatively

15 producing anti-idiotypic antibodies (Ab2) to a dilution of Anti-Band 3 antibody (Ab1) and allowing the mixture to react with Band 3 (isolated or expressed by an erythrocyte or other cell). The presence of Ab2 inhibits that reaction. Confirmation of the specificity of presumptive anti-idiotypic antibody produced by the hybridomas is conducted by measuring its binding to red blood cells (there should be none) and the

20 inhibition of monoclonal antibodies of the unrelated specificities: Rh29, K2, K14, M, N, B and Wr.sup.b (again there should be none). All these control tests being negative, the cells making the anti-anti-Band 3 antibody are cloned by limiting dilution.

These Ab2-producing clones (anti-anti-Band 3 ab) are then expanded and used

25 to produce large amounts of supernatant and ascitic fluids. Ab2 binding to, and inhibition of the red cell penetration by, *P. falciparum* merozoites in subsequent experiments demonstrates that the epitope recognized by the anti-anti-Band 3 monoclonal antibody is indeed the site used by *P. falciparum* since the parasite shares the binding structure of the monoclonal antibody.

30 Thus, using standard immunological, combinatorial chemistry, and three dimensional structural approaches, novel compounds are identified that specifically inhibit the invasion of the malaria parasite into host erythrocytes. The development of

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these compounds is based on the structure of ectoplasmic domains 5 and 6 of the Band 3 protein, the receptor region as defined by the amino acid sequence of the peptides B3_{5A}, B3_{5B}, B3_{5C}, and B3_{6A}, and their corresponding MSP-1 ligand(s) on the merozoite surface.

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EXAMPLE 5 Band 3 Is The Host Receptor Binding Merozoite Surface Protein-1 in the Malaria Parasite *Plasmodium Falciparum* Invasion of Red Blood Cells

10 **Methods**

Synthetic Band 3 Peptides

Human Band 3 peptides were synthesized with an N-terminal biotin tag and purified to homogeneity by HPLC (Peptide 1, amino acid 424-435; 2, 477-491; 3A, 538-557; 3B, 551-570; 4A, 623-642; 4B, 634-653; 4C, 644-663; 5A, 720-739; 5B, 731-750; 5C, 742-761; 6A, 807-826; 6B, 823-842; 6C, 838-857). Peptides were initially solubilized with minimal DMSO and serially diluted with either PBS or pertinent buffer to give $\leq 1\%$ DMSO final concentration in all subsequent assays. Peptide 6C was not soluble under these conditions and could not be used in the study.

20 ***Parasite Culture and Infection Determination***

Plasmodium parasite cultures were maintained at 37°C as described by Klotz, F.W., et. al., *J Exp Med* 165:1713-1718, 1987. To determine infection in RBCs, rings were counted from RBCs in Geimsa-stained thin smears.

25 ***Recombinant MSP₁₃₈, MSP₁₄₂ and MSP₁₁₉***

MSP₁₃₈ (SEQ ID NO:33), MSP₁₄₂ (SEQ ID NO:34), and MSP₁₁₉ (SEQ ID NO:35) genes were amplified from a *P. falciparum* (FCB1 strain) cDNA library (J. B. Dame) by PCR using the following primers: 5'-CTCGAGCTCAGGATAAACCC (SEQ ID NO:14, MSP₁₃₈, sense, 3121-3133, *Xho*I), 5'-GCGGCCGCACTTGTTAGT (SEQ ID NO:15, MSP₁₃₈, antisense, 4200-4193, *Not*I), 5'-CTCGAGCTGGAGAAGCAGTAACT (SEQ ID NO:16, MSP₁₄₂, sense, 4201-4218, *Xho*I), 5'-GCGGCCGCACTAAATGAAACTGTATA (SEQ ID NO:17, MSP₁₄₂, antisense, 5334-5321 *Not*I), 5'-CCGGGATCCAACATTTCACAACACCAA: (SEQ ID NO:18, MSP₁₁₉, sense, 4993-5009 *Bam*HI), 5'-

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CCGGAATTCAATGAACTGTATAATA (SEQ ID NO:19, MSP1₁₉, antisense, 5334-5318, *EcoRI*). Similar cDNA libraries as well as cDNA pools are available to the public from MR4, ATCC (www.malaria.atcc.org).

Primer sequences were based on the Wellcome strain of *P. falciparum* MSP1 (Holder, A.A., et al., *Nature* 317:270-273, 1985; Miller, L.H., et al., *Mol Biochem Parasitol* 59:1-14, 1993). MSP1₃₈ and MSP1₄₂ were cloned into pGEX6P-2 (Amersham Pharmacia Biotech, Piscataway, NJ), and MSP1₁₉ was cloned into pGEX-2TK (Amersham Pharmacia Biotech) using restriction sites indicated above. All three MSP1 domains were expressed as GST-fusion proteins in *E. coli* DH5 α , affinity purified using GSH beads. ³²P-labeled MSP1₁₉ was obtained by radiolabeling GST-MSP1₁₉ with ³²P- γ -ATP and removing the GST domain with thrombin (as described in Manufacturer's instructions).

Recombinant Band 3 Peptides 5ABC, 5BC, and 6AB

The 5ABC (amino acid 720-761), 5BC (731-761), and 6AB (807-842) genes were amplified from a human reticulocyte cDNA library by PCR and cloned into pGEX-2TK (Pharmacia) as GST fusion proteins. Primers: 5'-CCGGGATCCGGGATGCCCTGGCTCAGTGCCA (SEQ ID NO:20, 5ABC, sense, 2272-2293, *Bam*HI), 5'-CCGGAATTCCTTAGATCCGCTGCTCTTTGACCTC (SEQ ID NO:21, 5ABC and 5BC, antisense, 2397-2377, *Eco*RI), 5'-CCGGGATCCTCCGTCACCCATGCCAACGCC (SEQ ID NO:24, 5BC, sense, 2305-2325, *Bam*HI), 5'-CCGGGATCCGACCGCATCTTGCTTCTGTTCA (SEQ ID NO:25, 6AB, sense, 2533-2554, *Bam*HI), and 5'-CCGGAATTCCTTAGATCTGGATGCCCGTGAA (SEQ ID NO:26, 6AB, antisense, 2640-2620, *Eco*RI). GST-5ABC and GST-5BC were expressed, affinity purified, and radiolabeled with ³²P as above to obtain ³²P-labeled 5ABC and 5BC. GST-6AB expressed under same conditions was not soluble and could not be used in the study.

Blot Overlay Assay

Naturally released *P. falciparum* (3D7 strain) merozoites isolated as described (Mrema, J.E., et al., *Exp Parasitol* 54:285-295, 1982) and human RBC ghosts prepared as described (Dodge, J.T., et al., *Arch Biochem Biophys* 100:119-130, 1963)

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were subjected to SDS-PAGE. Proteins transferred onto nitrocellulose membrane were blocked overnight with 10% milk, 2% BSA, TBST (0.05%) at 4°C. After washing (4X) in TBST, the blot was incubated with biotinylated Band 3 peptides (each 400 µM) in TBST (0.025%), 10 mM phosphate buffer (pH 8.0), 60 mM KCl for 4 h at RT. After extensive washing, the blot was incubated with neutravidin-linked horseradish peroxidase (1:21,000, Pierce Chemical Co., Rockford, IL) in TBST (0.05%) with 2% BSA for 5-6 h at RT. After washing the blot with TBST (5X) and TBS (2X), bound peptides were visualized by the ECL method (Pierce Chemical Co.).

10 *Native MSP1 Binding to mAb 5.2 and 5ABC*

Radiolabeled parasite protein extract (180 µl) was incubated with either mAb 5.2 (20 µl) or GST-5ABC beads (40 µl, 50% slurry) for 22-24 h at 4°C. Protein G agarose beads (50 µl, 50% slurry) were added to the former mixture and further incubated for 3h. In both samples, beads were washed with PBS (2X) and proteins associated with beads were analyzed by SDS-PAGE followed by Coomassie staining and autoradiography. GST beads were used as control.

Yeast Two-Hybrid Assay

The 5ABC, 5BC, and 6AB genes were amplified by PCR as above and cloned into pGBKT7 (CLONTECH Laboratories, Inc., Palo Alto, CA). Primers were the same as the above except *EcoRI* (sense) and *BamHI* (antisense) restriction sites were used. MSP1_{38a}, MSP1_{38b}, MSP1₄₂, and MSP1₁₉ constructs were prepared by amplifying the MSP1 gene by PCR using the same template as above and cloned into pGADT7 (Clontech). Primers used were: 5'-

25 GGCCATATGGATGATACATCACATT (SEQ ID NO:27, MSP1_{38a}, sense, 3148-3163, *NdeI*), 5'-GGCCTCGAGGTTTCTAAACTGGCAT (SEQ ID NO:28, MSP1_{38a}, antisense, 3780-3764, *XhoI*), 5'-GGCCATATGTTTAAAGTATTAAGTA (SEQ ID NO:29, MSP1_{38b}, sense, 3781-3796, *NdeI*), 5'-GGCCTCGAGTTCTCCTGTTACTACTTG (SEQ ID NO:30, MSP1_{38b}, antisense, 4206-4189, *XhoI*), 5'-GCCGAATTCGCAGTAACTCCTTCCG (SEQ ID NO:31, MSP1₄₂, sense, 4207-4222, *EcoRI*) 5'-GCCGGATCCAATGAAACTGTATAATA (SEQ ID NO:32, MSP1₄₂, antisense, 5334-5318, *BamHI*). PCR primers for MSP1₁₉

were the same as above except *EcoRI* (sense) and *BamHI* (antisense) restriction sites were used. Yeast two hybrid assays using these recombinant plasmids were carried out using MATCHMAKER yeast-two hybrid system 3 (CLONTECH Laboratories, Inc.), see Table 2 for summary of results.

5

Indirect Immunofluorescence Assay

RBCs freshly collected into citrate phosphate dextrose buffer were washed (3X) and resuspended in RPMI (20% hematocrit). Thin smears of RBCs on glass slides were allowed to air-dry and fixed in methanol (20-30 sec). The slides were
10 washed with PBS (1X), blocked with 10% fetal bovine serum (FBS) in PBS (pH 7.4) for 1.5 h at 37 °C, and again washed with PBS (5X, 10 min each). Fixed cells were incubated with GST-MSP1₄₂ (1 µM), GST-MSP1₃₈ (8 µM), or GST (9µM) for 3.5 h at 37°C in 10% FBS. Slide samples were washed with PBS (5X), incubated with goat anti-GST antibody (1:1,000, Amersham Pharmacia Biotech), washed again (5X), and
15 incubated with rabbit anti-goat FITC-conjugated antibody (1:60, Sigma-Aldrich, St. Louis, MO). For visualizing spectrin, fixed cells treated with 10% FBS were incubated with a rabbit antibody raised against human spectrin (Sigma-Aldrich) followed by goat anti-rabbit FITC-conjugated antibody. All dilutions of proteins and antibodies were made in 10% FBS except FITC-conjugated antibodies.

20

Binding MSP1₃₈ and MSP1₁₉ to RBCs in Suspension

RBCs (500 µl of 50% suspension) were treated with Nm (3 mU, *Clostridium perfringens*, Roche) in 1 ml of RPMI (37 °C, 1 h) or α-ChT (0.5 mg/ml, Sigma-Aldrich) in 1 ml RPMI (RT, 10 or 40 min) followed by PMSF (2 mM) for 30 min. To
25 determine MSP1₃₈ binding, affinity purified GST-MSP1₃₈ was concentrated in PBS (pH 7.4) using Amicon spin column (10 kDa) and incubated with either Nm-treated or untreated human RBCs (7 µl packed volume) in PBS (pH 7.4, final 200 µl) at RT for 2 h. The mixture was passed through a bed of silicon oil (300 µl) by centrifugation. The RBC pellet was washed (1 ml) and resuspended (50-60 µl) in PBS and subjected
30 to SDS-PAGE followed by Western blot using anti-GST antibody (Amersham Pharmacia Biotech). GST was used as control. MSP1₁₉ binding: Enzyme-treated or untreated human and mouse RBCs (7 µl packed volume for human and 10 µl for

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mouse RBCs) were incubated with ^{32}P -labeled MSP1₁₉ in PBS as above. The incubation mixture was passed through silicon oil and RBCs were washed once as above. Radioactivity associated with resulting RBCs was measured using β -scintillation counter. Negative controls included samples with no RBCs and with
5 only RBCs (no ^{32}P -labeled MSP1₁₉).

Solution Binding of MSP1₃₈ and MSP1₄₂

Binding assays were performed as described (Oh, S.S., et al., *Mol Biochem Parasitol* 108:237-247, 2000) using the following conditions: 20 mM phosphate
10 buffer (pH 7.4), 120 mM NaCl, 1 mM DTT, 1.0 mg/ml BSA, 25 °C, 3.5 h, 280 μl final vol. ^{32}P -labeled 5ABC (10, 20, 40, 80 μM) and 5BC (21, 42, 84, 168 μM) respectively bound to GST-MSP1₄₂ (Fig. 4B top panel) and GST-MSP1₃₈ (Fig. 4B middle panel) on beads in concentration-dependent manner. ^{32}P -labeled MSP1₁₉ bound specifically to the 5ABC domain (Fig. 4B bottom panel) as statistically
15 analyzed by Student's *t* test. Binding to GST at comparable concentrations was not significant in all cases.

Metabolic Radiolabeling and Extraction of Parasite Proteins

Trophozoite-infected RBCs (60-100 μl packed volume) were purified on 70%
20 Percoll gradient from a synchronized *P. falciparum* (3D7) culture and returned to culture in RPMI (without methionine, cysteine, and leucine) containing 15% human serum without adding fresh RBCs. ^{35}S -methionine and cysteine (3:1 mixture, 1.6 mCi, specific activity 1,175 Ci/mmol, NEN (PerkinElmer Life Sciences, Boston, MA) and ^3H -leucine (250 μCi , specific activity 166 Ci/mmol, Amersham Pharmacia Biotech)
25 was then added and the culture was kept for 12 hr. The radioactive medium was replaced with the cold RPMI with 15% serum, and incubation continued on shaker (60-80 rpm) until segmenters and released merozoites appeared in the culture (about 8 h). Pellets were collected initially from the culture (500 rpm, 7 min) and then from the resulting supernatant (3,500 rpm, 15 min). Combined pellets were stored at –
30 80°C or used immediately in the next step. Parasite pellets were treated with extraction buffer containing 50 mM Tris-HCl (pH 8.0), 5 mM EDTA, 5 mM EGTA, 150 mM NaCl, 0.5% Triton X-100, 0.5% BSA, 2 $\mu\text{g/ml}$ aprotinin, 1 $\mu\text{g/ml}$ of

leupeptin, pepstatin A, bestatin, 10 mM PMSF and protease inhibitor cocktail (1X, Roche Molecular Biochemicals, Indianapolis, IN). The mixture was kept on ice for 1hr and spun at 15,000 rpm for 15min at 4°C. The supernatant was aliquoted and either used immediately or stored at -80 °C in binding assays.

5

Results

Band 3-Null RBCs Are Completely Resistant to *P. falciparum* Infection

To investigate the role of Band 3 in the host RBC membrane during the *P. falciparum* invasion process, we tested the susceptibility of Band 3 null mouse RBCs to *P. falciparum* (3D7 strain) invasion *in vitro*. When infected with synchronized *P. falciparum* trophozoites, wild type and Band 3 (+/-) mouse RBCs showed a typical invasion profile after 24 hours (Figure 1A) consistent with a previously study (Klotz, F.W., et al., *J Exp Med* 165:1713-1718, 1987). However, the Band 3 null RBCs did not show any new infection (rings) by *P. falciparum*. During the course of the experiment, Band 3 (-/-) mouse RBCs remained essentially intact in the culture as judged by the smears (Figure 1B) suggesting that the increased fragility of these RBCs (Perkins, L.L., et al., *Cell* 86:917-927, 1996) could not have played a significant role in the outcome of this experiment.

We found protein 4.2 (-/-) mice (Peters, L.L., et al., *J Clin Invest* 103:1527-1537, 1999) developed parasitemia at a rate comparable to the wild type when challenged with *P. yoelii* 17XL (unpublished data). In view of these findings, the lack of Band 3 in the RBC membrane appears to be the primary cause for complete resistance to *P. falciparum* infection in our Band 3-null RBC model. We hypothesize that Band 3 is functioning as a host receptor independently or in conjunction with GPA during *P. falciparum* invasion into RBCs.

Band 3 Peptides Block *P. falciparum* Invasion into RBCs

To investigate the possible role of Band 3 as a host receptor in *P. falciparum* invasion into RBCs, we employed a peptide scanning strategy based on recent topology models of human erythroid Band 3 (Figure 2A). Synthetic peptides were derived from the putative ectodomains of human RBC Band 3, and their ability to inhibit the *P. falciparum* (3D7 strain) invasion of human RBCs in culture was tested

30

using both visual counting (Schlichtherle, M., et al., *Methods in Malaria Research* 2000) and ^3H -hypoxanthine incorporation (Chulay, J.D., et al., *Exp Parasitol* 55:138-146, 1983) methods. In the visual counting assay (Figure 2B), Band 3 peptides 5A ($p = 0.016$), 5B ($p = 0.013$), 5C ($p = 0.006$), and 6A ($p = 0.006$) showed significant
5 levels of inhibition of invasion at 500 μM concentration as compared to other Band 3 peptides and the control (no peptide). Inhibition by these four Band 3 peptides was concentration dependent (50, 200, 500, and 1000 μM peptides, data not shown). In the ^3H -hypoxanthine uptake assay, peptide 6A ($p = 0.003$) showed strong inhibition whereas peptides 1 ($p = 0.148$), 5B ($p = 0.031$) and 5C ($p = 0.023$) showed moderate
10 but significant levels of inhibition of invasion at 400 μM concentration as compared to other Band 3 peptides and control (no peptides and unrelated peptide) samples (Figure 2C). Growth inhibition study using a similar ^3H -hypoxanthine uptake method showed that peptides 5B ($p = 0.033$), 5C ($p = 0.032$) and 6A ($p = 0.478$) added to the culture at 400 μM concentration did not affect the intraerythrocytic growth of the
15 parasite (Figure 2D). However, peptide 1 ($p = 0.320$), 2 ($p = 0.140$), and 5A ($p = 0.448$) samples showed mild reduction in ^3H -hypoxanthine uptake as compared to control (no peptide) suggesting these peptides might be inhibiting parasite growth in culture. Thus, our study showed peptides 5B, 5C and 6A were the most effective inhibitors targeting *P. falciparum* invasion among all Band 3 peptides tested. Further, invasion
20 blocking effects of peptides 5B, 5C, and 6A did not correlate with the net charge or pI of the peptides as these properties for peptides 4B, 1, and 3A, respectively were closely similar (Figure 2E).

The IC_{50} values (50% inhibition in the parasite invasion of RBCs) determined for peptide 5C and peptide 6A using the ^3H -hypoxanthine incorporation method were
25 $591 \pm 131 \mu\text{M}$ and $316 \pm 45 \mu\text{M}$, respectively (mean of three experiments \pm standard error). The IC_{50} values for peptides 5A and 5B could not be estimated because a saturable peptide concentration could not be reached with a low DMSO ($\leq 1\%$) concentration. It is noteworthy that our peptides 5A, 5B, 5C, and 6A were designed by randomly dividing the two ectodomains of Band 3, and thus these peptides may
30 not necessarily represent Band 3 amino acid compositions having the best inhibitory property. Our results, however, clearly demonstrate that peptides derived from two specific regions (amino acids 720-761 and 807-826) of human Band 3 inhibit the *P.*

falciparum invasion of human RBCs at a significant level and in a concentration dependent manner. These findings are consistent with the view that Band 3 functions as an important receptor in the parasite invasion of RBCs.

5 Band 3 Peptides Interact with *P. falciparum* Merozoite Proteins

A binding study was carried out between the synthetic Band 3 peptides and merozoite proteins to understand the mechanism by which Band 3 peptides 5B, 5C, and 6A inhibit the parasite invasion of RBCs. *P. falciparum* merozoites essentially free of contaminating RBC membrane components were isolated (Figure 3A), and
10 total merozoite proteins were separated by SDS-PAGE (Figure 3B). The purified merozoite protein mixture did not contain human RBC ghost proteins as judged by Coomassie blue staining. This was confirmed by Western blot using anti-spectrin and anti-Band 3 antibodies (not shown). A 1:1 mixture of peptides 5C and 6A showed specific binding to a number of merozoite proteins in the binding assay using a blot
15 overlay method (Figure 3C). Approximate molecular masses of these merozoite proteins are 175, 150, 125, 52, 48, 42, and 35 kDa (shown by arrowheads). In control samples, peptides 3A, 4A, and 2 did not show significant binding to any of these merozoite proteins, although a couple of weak signals were observed in the peptide 3A+4A sample. The peptide 5C+6A mixture did not show specific binding to RBC
20 ghost proteins. Our blot overlay results provide evidence that Band 3 functions as a receptor in the *P. falciparum* invasion of RBCs, and suggest that the underlying mechanism for the observed inhibition of invasion involves a specific binding of the Band 3 peptides to one or more merozoite ligands, thus competitively blocking its interaction with the RBC Band 3 receptor.

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Native MSP1 Binds to Recombinant Band 3

Among the merozoite proteins that specifically interacted with Band 3 peptides 5C and 6A the blot overlay binding assay (Figure 3C) were polypeptides migrating at approximately 42 kDa and 35 kDa mass. To investigate whether at least
30 one of the two polypeptides could be the 42 kDa or 38 kDa proteolytic fragment of MSP1 (MSP1₄₂ or MSP1₃₈), recombinant human Band 3 peptide 5ABC was prepared as a GST-fusion protein and affinity purified on GSH beads (Figure 3D). *P.*

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falciparum proteins metabolically labeled with ^{35}S -methionine and ^3H -leucine at the trophozoite stage were extracted from the mixture of late schizonts (segmenters) and naturally released merozoites using 0.5% Triton X-100. From this radiolabeled parasite protein extract, full-length MSP1 and MSP1₄₂ were immunoprecipitated with mAb 5.2 (MRA-94), a MSP1₁₉ (the 19 kDa C-terminal domain of MSP1)-specific monoclonal antibody raised against native *P. falciparum* MSP1 (Figure 3E, lane 3). In parallel, full-length MSP1 and MSP1₄₂ in the total protein mixture also bound specifically to the 5ABC domain in the binding assay using GST-5ABC (lane 1) and GST (lane 2, control) beads. Similar to the blot overlay assay results, a few radioactive protein Bands were specifically associated with GST-5ABC beads (not shown).

Characterization of MSP1-Band 3 Interaction

During its maturation, *P. falciparum* MSP1 (full-length) is processed to give proteolytic fragments referred to as MSP1₈₃, MSP1₃₀, MSP1₃₈, and MSP1₄₂ which together form a non-covalent complex on the merozoite surface as merozoites are released into circulation from infected RBCs (Holder, A.A., et al., *Mem Inst Oswaldo Cruz* 3:37-42, 1992). MSP1₄₂ then undergoes secondary processing producing MSP1₁₉ (C-terminal domain of MSP1₄₂) that is retained on the merozoite surface and carried into the newly invaded RBC while all other MSP1 fragments are shed off by an unclear mechanism. To characterize the interaction of the Band 3 receptor with proteolytic fragments of MSP1, two independent binding studies were carried out in solution *in vitro* and in yeast two-hybrid system *in vivo*. For the *in vitro* binding study, *P. falciparum* MSP1₃₈, MSP1₄₂ and MSP1₁₉ (Figure 4A) as well as human Band 3 peptides 5BC and 5ABC (Figure 3D) were expressed in *E. coli* and purified as GST-fusion proteins. GST-MSP1₃₈ was purified as a mixture of three C-terminal truncated polypeptides. GST-MSP1₃₈, GST-MSP1₄₂ and GST-MSP1₁₉ reacted with *P. falciparum* MSP1 T9/94 rabbit antiserum (MRA-75) generated against full-length MSP1 in Western blot (not shown). Similarly, GST-MSP1₄₂ and GST-MSP1₁₉ reacted with mAb 5.2 (Western blot shown for GST-MSP1₁₉ in Figure 4A, lane 9). Further, GST-MSP1₁₉, GST-5BC, and GST-5ABC were labeled with ^{32}P and treated

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with thrombin to afford pure ^{32}P -labeled MSP1₁₉ (Figure 4A, lane 10), 5BC (lane 11), and 5ABC (lane 12).

In solution-binding assay, ^{32}P -labeled 5ABC and 5BC bound to GST-MSP1₄₂ ($K_d = 36\ \mu\text{M}$) and GST-MSP1₃₈ ($K_d = 67\ \mu\text{M}$), respectively, when neither showed
5 significant binding to GST alone (Figures 4B, 4C, and 4D). Further, ^{32}P -labeled MSP1₁₉ bound to GST-5ABC at a significant level ($p = 0.027$) as compared to the GST control sample. These results demonstrate specific binding interactions between the MSP1 domain and the Band 3 peptide domain. The three-dimensional structure for the 5ABC or 5BC region of Band 3 is not known. A difference of 11 amino acids
10 in the primary structure of 5ABC and 5BC may be important for showing different binding property towards MSP1 in our *in vitro* experiments. What is clearly implicated from these results is that an ectoplasmic region of human Band 3 represented by amino acids 720-761 constitutes the binding site for MSP1₃₈, MSP1₄₂, and MSP1₁₉ presumably involving a number of specific binding interactions
15 contained within this region.

In the second approach using a GAL4-based yeast two-hybrid assay, Band 3 peptides (5ABC, 5BC, 6AB) and MSP1 domains (MSP1_{38a}, MSP1_{38b}, MSP1₄₂, MSP1₁₉) were expressed as a fusion to the GAL4 DNA-binding (DNA-BD) domain and GAL4 activation domain (AD), respectively. MSP1₃₈ was divided into MSP1_{38a}
20 and MSP1_{38b} in view of C-terminal truncated MSP1₃₈ used in the solution-binding assay. (See Table 2). Peptide 5ABC interacted with MSP1₄₂, MSP1₁₉, and MSP1_{38b}, peptide 5BC interacted with only MSP1_{38a}, and peptide 6AB interacted with MSP1₄₂ and MSP1₁₉. For the yeast two-hybrid assay, the co-transformation method was used to analyze the protein interaction in yeast AH109 cells using SD/-Leu/-Trp selection
25 plates. Activation of the *MEL1* reporter gene upon specific binding of a MSP1 domain to a Band 3 peptide gave positive blue colonies using α -galactosidase assay. Plasmids expressing only the inserted gene of a Band 3 peptide or MSP1 domain did not undergo autonomous transcriptional activation of the reporter gene. No interaction was observed in other two-hybrid samples of Band 3 peptide and MSP1
30 domain. All positive and negative controls gave anticipated results. Specific interactions demonstrated with peptides 5ABC and 5BC in the yeast two-hybrid assay were consistent with the solution-binding assay results (Figures 4B, 4C, and 4D). The

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- expression of 6AB as a soluble form in yeast cells (data not shown) provides support that the observed 6AB-MSP1₄₂ and 6AB-MSP1₁₉ interactions are specific. Binding results from two independent *in vitro* and *in vivo* methods summarized in Figure 4E were remarkably similar indicating that a key function of MSP1₃₈, MSP1₄₂, and
- 5 MSP1₁₉ is to interact with Band 3 in the RBC membrane during merozoite invasion of RBCs.

Table 2. Summary of Yeast Two-Hybrid Assay

Fusion to DNA- BD (pGBKT7 vector)	Fusion to AD (PgaDT7 vector)	Binding Property
None	None	—
p53	None	—
None	SV40 large T-antigen	—
p53	SV40 large T-antigen	+++
Lamin C	SV40 large T-antigen	—
5ABC	SV40 large T-antigen	—
5BC	SV40 large T-antigen	—
6AB	SV40 large T-antigen	—
Lamin C	MSP1 _{38a}	—
Lamin C	MSP1 _{38b}	—
Lamin C	MSP1 ₄₂	—
p53	MSP1 ₄₂	—
p53	MSP1 ₁₉	—
5ABC	MSP1 _{38a}	—
5ABC	MSP1 _{38b}	+
5ABC	MSP1 ₄₂	+++
5ABC	MSP1 ₁₉	+++
5BC	MSP1 _{38a}	++
5BC	MSP1 _{38b}	—
5BC	MSP1 ₄₂	—

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5BC	MSP1 ₁₉	—
6AB	MSP1 _{38a}	—
6AB	MSP1 _{38b}	—
6AB	MSP1 ₄₂	+++
6AB	MSP1 ₁₉	+++

Proteolytic Fragments of MSP1 Bind to RBCs in Sialic Acid-Independent Manner

Earlier studies showed that native *P. falciparum* MSP1 (full-length) bound to RBCs in sialic acid-dependent manner (Perkins, M.E., et al., *J Immunol* 141:3190-3196, 1988; Su, S., et al., *J Immunol* 151:2309-2317, 1993). More recently, however, it has been shown that a number of peptides derived from MSP1₈₃, MSP1₃₈, and MSP1₄₂ bound to sialic acid-depleted RBCs with relatively high affinity (Urquiza, M., et al., *Parasite Immunol* 18:515-526, 1996). A recombinant segment (115 amino acids) of MSP1₃₈ referred to as p115MSP-1 also bound to wild-type human RBCs as well as En(a-) human RBCs lacking GPA (Nikodem, D., et al., *Mol Biochem Parasitol* 108:79-91, 2000).

To further examine the RBC binding property of MSP1 proteolytic fragments, we first carried out a binding study using GST-MSP1₃₈ and intact human RBCs either untreated or pretreated with neuraminidase (Nm). Removal of sialic acids on the surface of Nm-treated RBCs was confirmed by periodic acid-Schiff (PAS) staining using RBC ghosts (Figure 5A, lane 9). Both types of RBCs were incubated with purified GST-MSP1₃₈, sedimented through a bed of silicon oil as described (Nikodem, D., et al., *Mol Biochem Parasitol* 108:79-91, 2000) and subjected to SDS-PAGE followed by Western blotting using anti-GST antibody. Two truncated forms of GST-MSP1₃₈ (45 and 30 kDa Bands shown above in Figure 4A, lane 5) bound to both treated (Figure 5B, lanes 1) and untreated (lane 3) intact RBCs in suspension. GST alone did not bind to either type of RBCs (lanes 2, 4). These results demonstrate that MSP1₃₈ specifically interacted with the extracellular component of human RBCs in sialic acid-independent manner. This is consistent with our finding that peptide 5BC representing a non-glycosylated ectodomain of Band 3 bound to MSP1₃₈ in solution and MSP1_{38a} in the yeast two-hybrid system.

In demonstrating the sialic acid-independent binding of MSP1₄₂ to intact RBCs, we used its C-terminal secondary processing fragment MSP1₁₉ known to be carried into newly invaded RBCs (Blackman, M.J., et al., *J Exp Med* 172:379-382, 1990). To carry out this study, we treated intact human and mouse RBCs with either chymotrypsin (ChT) or Nm (Figure 5A). In ChT-treated human (Figure 5A, lane 3) and mouse (lane 6) RBCs, full-length Band 3 (arrowheads) was digested into 55 kDa N-terminal and 38 kDa C-terminal fragments (arrows) as reported (Steck, T.L., et al., *Biochemistry* 17:1216-1222, 1978). The 38 kDa fragment known to be less stable than the 55 kDa fragment often appeared as a diffused faint Band in Coomassie gel (lane 6). As expected, Band 3 (arrowhead) was intact in Nm-treated human RBCs (lane 2) and mouse RBCs (lane 5). Western blotting of the human RBC ghost samples using anti-Band 3 antibody specific for the N-terminal cytoplasmic domain of human Band 3 confirmed these results (lanes 10-12). The extracellular region of GPA was also digested by ChT at an appreciable rate as evident in the PAS-stained gel (lane 8) and anti-GPA Western blot (lane 14) of human RBC samples. A significant level of sialic acid residues attached to ChT-digested GPA fragments was retained on the RBC surface as judged by PAS staining (lane 8). The ChT-digested GPA fragments at approximately 62 kDa and 47 kDa molecular mass (shown with asterisks) in the human RBC sample (lane 14) were consistent with the previous report (Roggwiller, E., et al., *Mol Biochem Parasitol* 82:13-24, 1996).

These various types of RBCs were reacted with ³²P-labeled MSP1₁₉ (Figure 4A, lane 10) in suspension, and the radioactivity associated with RBCs was analyzed. Human RBCs treated with ChT for 10 min and 40 min showed 30% ($p=0.010$) and 37% ($p=0.040$) reduction and with Nm (40 min) showed 52% increase ($p=0.054$) in their ability to bind MSP1₁₉ as compared to the untreated RBCs (Figure 5C). In mouse RBC samples, ChT-treated RBCs (40 min) showed 41% reduction ($p=0.005$) and Nm-treated RBCs (40 min) showed 51% increase ($p=0.086$) as compared to the untreated RBCs. These results show for the first time that *P. falciparum* MSP1₁₉ binds to intact human and mouse RBCs in sialic acid-independent manner. Previously, a synthetic peptide referred to as peptide 5501 (20 amino acids) derived

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from the N-terminus of MSP1₁₉ was reported to bind human RBCs (Urquiza, M., et al., *Parasite Immunol* 18:515-526, 1996). Further, the reduced binding of MSP1₁₉ in ChT-treated RBC samples in our assays is directly correlated with alterations in Band 3 and GPA peptide backbones on the RBC surface. Nm treatment provided both human and mouse RBCs with increased ability to bind MSP1₁₉ demonstrating that the binding interaction does not involve sialic acids. Presumably, the increase in binding results from relatively unhindered access to the protein receptor upon removal of sialic acid residues from the RBC surface.

10 Band 3 is important for *P. falciparum* MSP1 Binding to RBCs

Two approaches were considered to obtain evidence that Band 3 mediates sialic acid-independent interaction between MSP1 and RBCs. First, the above-described RBC binding assay was performed using ³²P-labeled MSP1₁₉ and intact Band 3 (–/–) mouse RBCs in suspension. As compared to untreated wild-type mouse RBCs, there was a 72% reduction ($p = 0.277 \times 10^{-6}$) of radioactivity associated with Band 3 (–/–) mouse RBCs (Figure 5C). However, since background radioactivity from the negative control sample containing no RBCs was about 18% of the positive control (untreated wild-type RBCs), the actual radioactivity associated with Band 3 (–/–) mouse RBCs was mere 10% above the background. Thus, intact Band 3 (–/–) mouse RBCs lacking both Band 3 and GPA from the plasma membrane showed a relatively insignificant level of binding to MSP1₁₉. In the second method, we carried out an indirect immunofluorescence assay (IFA) using wild-type human and mouse RBCs and Band 3 (–/–) mouse RBCs fixed in methanol. GST-MSP1₄₂ and GST-MSP1₃₈ (truncated forms) bound to human as well as mouse wild-type RBCs while GST alone did not, demonstrating that the observed binding was specific to the MSP1₄₂ and MSP1₃₈ domain, respectively. However, neither GST-MSP1₄₂, GST-MSP1₃₈, nor GST alone bound to Band 3 (–/–) mouse RBCs. In the indirect immunofluorescence assay of *P. falciparum* MSP1 binding to human and mouse RBCs, Anti-spectrin antibody staining confirmed all RBCs were morphologically normal.

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Our truncated MSP1₃₈ (the 45 kDa GST-fusion protein) specifically bound to sialic acid-depleted RBCs (Figure 5B), and the p115MSP-1 construct substantially overlapping (at least about 60 amino acids) our MSP1₃₈ was shown to bind to intact human RBCs lacking the sialoglycoprotein GPA (Nikodem, D., et al., *Mol Biochem Parasitol* 108:79-91, 2000). In view of these findings, our IFA results demonstrate that the peptide backbone of Band 3 in the RBC membrane is important for binding MSP1₃₈ to the RBC surface. In this context, peptide 5ABC representing a putative ectoplasmic region (amino acids 720-761) of Band 3 specifically bound to MSP1₃₈ and MSP1_{38a} in our binding assays (Figure 4E). This region of Band 3 shares 98% sequence identity between mouse and human. Our results thus indicate that *P. falciparum* MSP1₃₈ interacts with the Band 3 receptor on both human and mouse RBC surface during the parasite invasion of RBCs.

The MSP1₄₂ binding results in our IFA are consistent with the binding of its C-terminal fragment MSP1₁₉ to RBCs in suspension (Figure 5C). Further, the level of MSP1₁₉ binding to RBCs decreased considerably with the limited ChT digestion of Band 3 and GPA peptide backbones on the RBC surface. Previously, ChT-treated human RBCs were shown to have marked reduction in invasion by *P. falciparum* as compared to untreated RBCs (Perkins, M., *J Cell Biol* 90:563-567, 1981). Since recombinant *P. falciparum* MSP1₄₂ and MSP1₁₉ bound to two distinct non-glycosylated ectoplasmic regions (5ABC and 6AB) of erythroid Band 3 (Figure 4E) and native MSP1₄₂ specifically bound to 5ABC (Figure 3E), our results taken together show that Band 3 functions as the receptor also for MSP1₄₂ and MSP1₁₉. A possibility that the peptide backbone of GPA might also play a secondary role in binding MSP1₄₂ and/or MSP1₁₉ to RBCs cannot be completely ruled out. Results from our RBC binding studies support the idea that Band 3 (-/-) RBCs are completely refractory to *P. falciparum* invasion due to the lack of an important interaction involving host Band 3 and proteolytic fragments of merozoite MSP1 such as MSP1₃₈, MSP1₄₂ and MSP1₁₉.

Example 6 Identification of *Plasmodium* Polypeptides that Interact with Band 3**Methods***The "Bait" construct*

The bait construct containing the 5ABC domain (amino acids 720-761) of human Band 3 was prepared by a PCR amplification of the corresponding gene from a human reticulocyte cDNA library and cloning into a yeast two-hybrid vector pGBKT7 (Clontech) as a fusion to GAL4 DNA-binding (DNA-BD). Primers used were 5'-CCGGAATTCGGGATGCCCTGGCTCAGTGCCA-3' (SEQ ID NO:36, sense, 2272-2293, EcoRI), 5'-CCGGGATCCTTAGATCCGCTGCTCTTTGACCTC-3' (SEQ ID NO:37, antisense, 2397-2377, BamHI). The bait construct pGBKT7-5ABC was transformed into yeast AH109. The expression of 5ABC domain as a soluble fusion protein in yeast was confirmed by Western blotting of the cell lysate supernatant as described in the Clontech manual. The absence of autonomous transcriptional activation of the reporter gene by the bait domain 5ABC was confirmed on agar plates made with a minimal synthetic dropout medium (SD) lacking tryptophan (SD/-Trp), tryptophan and histidine (SD/-Trp-His), and tryptophan and adenine (SD/-Trp-Ade) using the X- α -Gal assay according to the Clontech manual.

Screening of the cDNA library in yeast two-hybrid system

The screening of *P. falciparum* (3D7 strain) cDNA library transformed into yeast PJ69-2A (Clontech) was performed by the standard yeast mating method using the bait construct pGBKT7-5ABC transformed into yeast Y187 (Clontech) as described in the Clontech manual (MATCHMAKER Two-Hybrid System 3). The mating mixture was spread onto 50 large (150 mm) plates of SD/-His-Leu-Trp (TDO). The cells were grown at 30°C for 10 days. His⁺ colonies selected by this procedure were streaked onto SD/-His-Ade-Leu-Trp/X-a-Gal (QDO) plates and grown for 1 week at 30°C. Both positive (pGBKT7-53 + pGADT7-T) and negative (pGBKT7 + pGADT7) controls (Clontech) were included at each round of selection. Ade⁺, His⁺, Mel⁺ yeast colonies were selected from these plates for further analysis. Following overnight growth in the SD/-Leu-Trp-His broth at 30 °C, cells from each clone were harvested. Plasmid DNAs isolated upon cell lysis were transformed into

E. coli DH5 α , and transformants were selected on LB/ampicillin plates. A selected colony from each transformation plate was subcultured overnight in the LB/ampicillin broth and the plasmid DNA was isolated by alkaline lysis. The nucleotide sequence of *P. falciparum* cDNA inserts was determined to identify these positive clones.

5

Results

Analysis of positive clones

From the cDNA library containing 9.8×10^6 independent transformants, we selected 116 His⁺ colonies on SD/-His-Leu-Trp plates. These 116 His⁺ colonies were subjected to a second round of selection by streaking onto the SD/-Ade-His-Trp-Leu plate. Because the induction of the more tightly controlled GAL2-ADE2 reporter gene is required under these higher stringent selection conditions (Ade⁺), only 20 out of 116 colonies grew. Approximately 500 base pair sequence of *P. falciparum* cDNA inserts was determined from the 5' end of the gene. Subsequent Blast analyses (at the NCBI and PlasmoDB website) of these insert cDNA sequences revealed that two of them were *P. falciparum* ABRA and RhopH3 (Table 3). All other cDNA insert sequences (total= 18) were found only in PlasmoDB. Upon further analysis of the insert sequences, we found that only six of the eighteen insert cDNAs were non-redundant and in correct reading frame. These six were novel *P. falciparum* genes with no known functions associated with their gene products (Table 3). We have designated names to these six gene products as Band 3 Binding Protein (BBP)-1, 2, 3, 4, 5, and 6. The binding interaction between the Band 3 peptide 5ABC and each of the eight *P. falciparum* gene products was independently confirmed using the cotransformation and/or mating method (Clontech manual) in subsequent yeast two-hybrid assays under various stringency conditions as summarized in Table 3.

25

Table 3. Summary of cDNA library screening in yeast two-hybrid system

Clone Number	BD-5ABC	-T-	-T-L-	-T-L-	-T-L-A-	Gene Name used by		Designated
	+ AD-X ^a	L	H	A	H	PlasmID ^b	Name ^c	
8		++	++	+	+	chr4_1.gen_205	BBP-1	
12		++	++	+	+	chr9_1.gen_311 ^d	RhopH3	
14		++	++	+	+	chr12_1.gen_395 ^e	ABRA	
48		++	++	+	+	chrBLOB_004238.gen_2	BBP-2	
59		++	++	+	+	chr5_1.gen_122	BBP-3	
74		++	++	+	+	chr14_1.gen_490	BBP-4	
94		++	++	+	+	chr7_000072.gen_1	BBP-5	
101		++	++	+	+	chr5_1.gen_79	BBP-6	

^a X denotes the clone number used in our cDNA library screening assay.^b See attachment, Supplemental Material for Dataset 2: Description of genes by PlasmIDB.^c BBP (Band 3 Binding Protein)-1, 2, 3, 4, 5, and 6 are names we have designated to respective gene products.^d *P. falciparum* RhopH3, GenBank Accession No: M65059.^e The 101 KDa *P. falciparum* malaria antigen (p101) termed Acidic Basic Repeat Antigen (ABRA), GenBank Accession No: J03902.

Table 4 depicts the Blast Sequence Homology results obtained for the Band 3 peptides having SEQ ID NOs:1, 2, 3, and 4.

TABLE 4. BAND 3 BLAST HOMOLOGY SEQUENCES

SEQ ID NO. 1

P23562,NP_036783.1,CAA31128.1,NP_000333.1,CAA27555.1,NP_035533.1,AAA37278.1,AAD43354.1,AAD43593.1,A30816 P15575,
P32847,AAF19584.2,AAG23156.1,AAG23157.1,AAG23155.1,AAF00977.1,NP_003031.1,AAF23240.1,XP_004678.1,NP_033233.1,AAG23158.1,AAC50964.1,NP_058744.1,AAF19583.2,AAB66833.1,AAC59881.1,AAG23154.1,P48746,O18917,NP_033234.1,AAG25582.1,AAB05850.1,AAG25583.1,XP_002605.1,NP_058745.1,AAD14330.1,NP_005061.1,S31828,CAA60670.1,Q9Z0S8,AAF50207.1,BAA34459.1,NP_067505.1,NP_004849.1,NP_071341.1,BAB17922.1,AAF50207.1,AAF52496.1,AAF52497.1,KADBID 004606,
AAA54840.1,AAA54837.1,AAA54839.1,NM_012651.1,X77738.1,NM_000342.1,M27819.1,L35930.1,X12609.1,J02756.1,X03917.1,M29379.1,NM_011403.1,X02677.1,J04793.1,XM_008364.1,AC003043.1,AF163826.1,AF163828.1,AF163827.1,M19496.1,M23404.1,X61699.1,Z50848.1,U62531.1,U76669.2,AF012895.1,AF255774.1,NM_009207.1,J04036.1,XM_004678.1,NM_003040.1,S45791.1,X62137.1,NM_017048.1,U48889.1,J05166.1,X03918.1,U20523.1,AF120099.1,M87060.1,AC009955.4,NM_009208.1,M28383.1,AF031650.1,NM_017049.1,AF294651.1,J05167.1,S80168.1,L27213.1,XM_002605.1,NM_005070.1,U05596.1,X87211.1,X70797.1,AF121253.1,AE003550.2,AE003550.2,BF760317.1,BF724738.1,BF726058.1,BF726058.1,BF726058.1,AW239627.1,BE255812.1,BE667859.1,BE683882.1,BE259443.1,BF760317.1,BF724738.1,BE683941.1,AA822979.1,BE512723.1,BF726058.1,AL121219.1,BE231685.1,AA362927.1,BF526005.1,F06947.1,AI592399.1,AI121401.1,BE683881.1,BF688963.1,BF688491.1,AW372960.1,AA755536.1,N58147.1,AW358179.1,BE387636.1,AA979500.1,T86708.1,AC025326.3,AC010973.4,AC016330.5,AC016170.2,AC010044.5,AC014376.1,AL291529.1,I08446.1,E15207.1,I08447.1,AX001285.1,AX001281.1,AX001279.1,NM_000342.1,XM_008364.1,NM_003040.1,XM_004678.1,XM_002605.1,NM_005070.1,NP_000333.1,NP_003031.1,XP_004678.1,XP_002605.1,NP_005061.1,NT_010755.1,

Seq ID NO:2

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EQUIVALENTS

- Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the following claims.

All references disclosed herein are incorporated by reference in their entirety.

We claim:

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Claims

1. An isolated Band 3 polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO. 1, 2, 3, and 4 as shown herein:
 - 5 SEQ ID NO:1: GMPWLSATTVRSVTHANALT (also referred to herein as sequence B3_{5A});
 - SEQ ID NO:2: SVTHANALTVMGKASTPGAA (also referred to herein as sequence B3_{5B});
 - SEQ ID NO:3: GKASTPGAAAQIQEVKEQRI (also referred to herein as sequence
 - 10 B3_{5C});
 - SEQ ID NO:4: DRILLLFKPPKYHPDVPYVK (also referred to herein as sequence B3_{6A}); and unique fragments thereof, wherein the unique fragments
 - (1) bind to an MSP-1 polypeptide and
 - (2) exclude the sequences set forth in Table 4:Band 3 Blast Homology
 - 15 Sequences.
2. An isolated nucleic acid molecule that encodes the isolated polypeptide of claim 1.
- 20 3. An expression vector comprising the isolated nucleic acid of claim 2 operably linked to a promoter.
4. A host cell transfected or transformed with an expression vector of claim 3.
- 25 5. An immunogenic composition comprising:
 - one or more isolated polypeptides of claim 1; and
 - a pharmaceutically acceptable carrier;
 - wherein the polypeptides are present in an effective amount to induce an immune system response.
- 30 6. The composition of claim 5, further comprising an adjuvant.

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7. A method of making a medicament, comprising:
placing one or more isolated polypeptides of claim 1 in a pharmaceutically acceptable carrier.
- 5 8. A method for identifying a candidate mimetic of an isolated polypeptide of claim 1, comprising
providing an MSP-1 polypeptide which binds the isolated polypeptide of claim 1,
contacting the MSP-1 polypeptide with a test molecule, and
10 determining the binding of the test molecule to the MSP-1 polypeptide,
wherein a test molecule which binds to the MSP-1 polypeptide and inhibits binding of the MSP-1 polypeptide to the polypeptide of claim 1 is a candidate mimetic of the isolated polypeptide of claim 1.
- 15 9. A protein microarray comprising at least one isolated Band 3 polypeptide selected from the group consisting of SEQ ID NOS. 1, 2, 3, and 4.
10. An anti-Band 3 antibody or fragment thereof that selectively binds to a polypeptide of claim 1, wherein the antibody inhibits infection of cells by *P. falciparum* merozoite malaria parasite.
20
11. An anti-idiotypic antibody which selectively binds to the idiotype of the antibody of claim 10.
- 25 12. A method for making an anti-idiotypic antibody comprising:
immunizing an animal with an antibody of claim 10 under conditions to elicit an immune system response to an idiotype of said antibody of claim 10.
- 30 13. A method for treating a malaria infection, comprising:
administering to a subject in need of such treatment, an effective amount of an anti-Band 3 antibody of claim 10 to treat the malaria infection.

14. A method for inducing an immune system response to treat a malaria infection, comprising:

administering to a subject in need of such treatment, an effective amount of an anti-Band 3 antibody of claim 10 under conditions to induce an anti-idiotypic immune response to the anti-Band 3 antibody idiotype.

15. A method for identifying a candidate mimetic of a MSP-1 polypeptide, comprising

providing an isolated Band 3 polypeptide which binds a MSP-1 polypeptide,
contacting the Band 3 polypeptide with a test molecule, and
determining the binding of the test molecule to the Band 3 polypeptide,
wherein a test molecule which binds to the isolated Band 3 polypeptide and inhibits binding of the Band 3 polypeptide to the MSP-1 polypeptide is a candidate mimetic of the MSP-1 polypeptide.

15

16. The method of claim 15, wherein the MSP-1 polypeptide has a sequence selected from the group consisting of SEQ ID NO:11, SEQ ID NO:12, SEQ ID NO:13, SEQ ID NO:33, SEQ ID NO. 34, and SEQ ID NO:35.

17. The method of claim 15, wherein the test molecule is an antibody.

18. An isolated polypeptide, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NOs:11, 12, 13, 33, 34, and 35, or fragments thereof.

25

19. A pharmaceutical composition comprising:
one or more isolated polypeptides of claim 18 and
a pharmaceutically acceptable carrier;
wherein the polypeptides are present in an effective amount to induce an immune system response.

30

20. The pharmaceutical composition of claim 19, further comprising an adjuvant.

21. A method of making a medicament, comprising:
placing one or more isolated polypeptides of claim 19 in a pharmaceutically acceptable carrier.
- 5 22. A method of preventing or treating a malaria infection, comprising administering a pharmaceutical composition of claim 19 to a subject in need of such treatment in an amount effective to prevent or treat the malaria infection.
- 10 23. A malaria polypeptide binding polypeptide that selectively binds to a isolated malaria polypeptide of claim 18, wherein the binding polypeptide is an antibody or antigen-binding fragment of an antibody.
24. A pharmaceutical composition comprising the malaria polypeptide binding
15 polypeptide of claim 23, in a pharmaceutically acceptable carrier.
25. A method of preventing or treating a malaria infection, comprising administering the pharmaceutical composition of claim 24 to a subject in need of such treatment in an amount effective to prevent or treat the malaria infection.
- 20 26. An isolated nucleic acid, wherein the nucleic acid molecule comprises a nucleotide sequence selected from the group consisting of SEQ ID NOs:54-59, or fragments thereof.
- 25 27. An isolated Band 3 polypeptide, comprising an amino acid sequence selected from the group consisting of SEQ ID NO. 1, 2, 3, and 4 as shown herein:
- SEQ ID NO:1: GMPWLSATTVRSVTHANALT (also referred to herein as sequence B3_{5A});
- 30 SEQ ID NO:2: SVTHANALTVMGKASTPGAA (also referred to herein as sequence B3_{5B});

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SEQ ID NO:3: GKASTPGAAAQIQEVKEQRI (also referred to herein as sequence B3_{sc});

SEQ ID NO:4: DRILLFKPPKYHPDVPYVK (also referred to herein as sequence B3_{6A}), and unique fragments thereof, wherein the unique fragments

- 5 (1) bind to an isolated polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:46-53, or fragment thereof, and
- (2) exclude the sequences set forth in Table 4:Band 3 Blast Homology Sequences.
- 10 28. An isolated nucleic acid molecule that encodes the isolated Band 3 polypeptide of claim 27.
29. An expression vector comprising the isolated nucleic acid of claim 29 operably linked to a promoter.
- 15 30. A host cell transfected or transformed with an expression vector of claim 29.
31. An immunogenic composition comprising:
 one or more isolated Band 3 polypeptides of claim 27 and
20 a pharmaceutically acceptable carrier;
 wherein the Band 3 polypeptides are present in an effective amount to induce an immune system response.
32. The composition of claim 31, further comprising an adjuvant.
- 25 33. A method of making a medicament, comprising:
 placing one or more isolated Band 3 polypeptides of claim 27 in a pharmaceutically acceptable carrier.
- 30 34. A method for identifying a candidate mimetic of an isolated Band 3 polypeptide of claim 27, comprising

providing a malaria polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:46-53, or fragment thereof that binds the isolated Band 3 polypeptide or fragment thereof of claim 27,

contacting the malaria polypeptide or fragment thereof, with a test molecule,
5 and

determining the binding of the test molecule to the malaria polypeptide or fragment thereof, wherein a test molecule which binds to the polypeptide or fragment thereof and inhibits binding of the isolated Band 3 polypeptide to the malaria polypeptide, is a candidate mimetic of the isolated Band 3 polypeptide of claim 27.

10

35. A method for identifying a candidate mimetic of an isolated malaria polypeptide, comprising

providing a Band 3 molecule which binds a malaria polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOs:46-53,

15

contacting the Band 3 molecule with a test molecule, and

determining the binding of the test molecule to the Band 3 molecule, wherein a test molecule which binds to the Band 3 molecule and inhibits binding of the malaria polypeptide with the Band 3 polypeptide is a candidate mimetic of the malaria polypeptide.

20

36. The method of claim 35, wherein the test molecule is an antibody.

37. An isolated polypeptide molecule comprising an amino acid sequence selected from the group consisting SEQ ID NOs:46-52.

25

38. A pharmaceutical composition comprising:

one or more isolated polypeptides of claim 37 and

a pharmaceutically acceptable carrier;

wherein the polypeptides are present in an effective amount to induce an

30 immune system response.

39. The pharmaceutical composition of claim 38, further comprising an adjuvant.

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40. A method of making a medicament, comprising:
placing one or more isolated polypeptides of claim 38 in a pharmaceutically acceptable carrier.

5

41. A method of preventing or treating a malaria infection, comprising administering a pharmaceutical composition of claim 38 to a subject in need of such treatment in an amount effective to prevent or treat the malaria infection.

10 42. A malaria polypeptide binding polypeptide that selectively binds to a isolated malaria polypeptide of claim 37, wherein the binding polypeptide is an antibody or antigen-binding fragment of an antibody.

43. A pharmaceutical composition comprising the malaria polypeptide binding
15 polypeptide of claim 42, in a pharmaceutically acceptable carrier.

44. A method of preventing or treating a malaria infection, comprising administering the pharmaceutical composition of claim 43 to a subject in need of such treatment in an amount effective to prevent or treat the malaria infection.

20

45. An isolated nucleic acid molecule selected from the group consisting of:
(a) nucleic acid molecules which hybridize under stringent conditions to a nucleic acid molecule comprising a nucleotide sequence selected from the group consisting of SEQ ID NO:38-44 and which codes for a *Plasmodium* polypeptide,

25 (b) deletions, additions and substitutions of the nucleic acid molecules of (a), which code for a *Plasmodium* polypeptide,

(c) nucleic acid molecules that differ from the nucleic acid molecules of (a) or (b) in codon sequence due to the degeneracy of the genetic code, and
(d) complements of (a), (b) or (c).

30

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46. The isolated nucleic acid molecule of claim 45, wherein the isolated nucleic acid molecule comprises a nucleotide sequence selected from the group consisting of SEQ ID NO:38-44.
- 5 47. An isolated nucleic acid molecule selected from the group consisting of:
(a) a unique fragment of the nucleotide sequence selected from the group consisting of:
nucleotides 1-1287 of SEQ ID NO:38 between 12 and 1286 nucleotides in length,
nucleotides 1-3576 of SEQ ID NO:39 between 12 and 3557 nucleotides in length,
10 nucleotides 1-903 of SEQ ID NO:40 between 12 and 902 nucleotides in length,
nucleotides 1-1203 of SEQ ID NO:41 between 12 and 1202 nucleotides in length,
nucleotides 1-3996 of SEQ ID NO:42 between 12 and 3995 nucleotides in length, and
nucleotides 1-876 of SEQ ID NO:43 between 12 and 875 nucleotides in length, and
nucleotides 1-2712 of SEQ ID NO:44 between 12 and 2711 nucleotides in length, and
15 (b) complements of (a),
wherein the unique fragments exclude nucleic acids having nucleotide sequences that are contained within SEQ ID NO:38-44, and that are known as of the filing date of this application.
- 20 48. An expression vector comprising the isolated nucleic acid molecule of claim 46 operably linked to a promoter.
49. An isolated polypeptide molecule comprising a unique fragment of amino acid sequence SEQ ID NO:53 that binds to a Band 3 polypeptide.

25

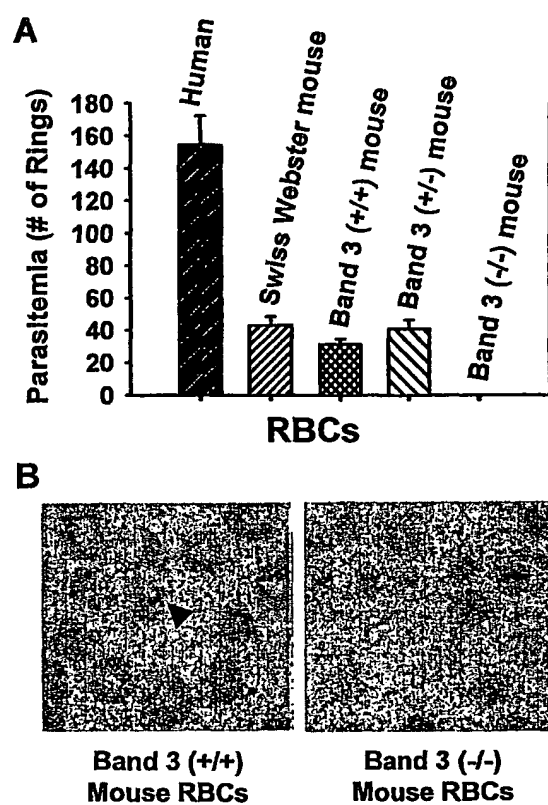


Figure 1

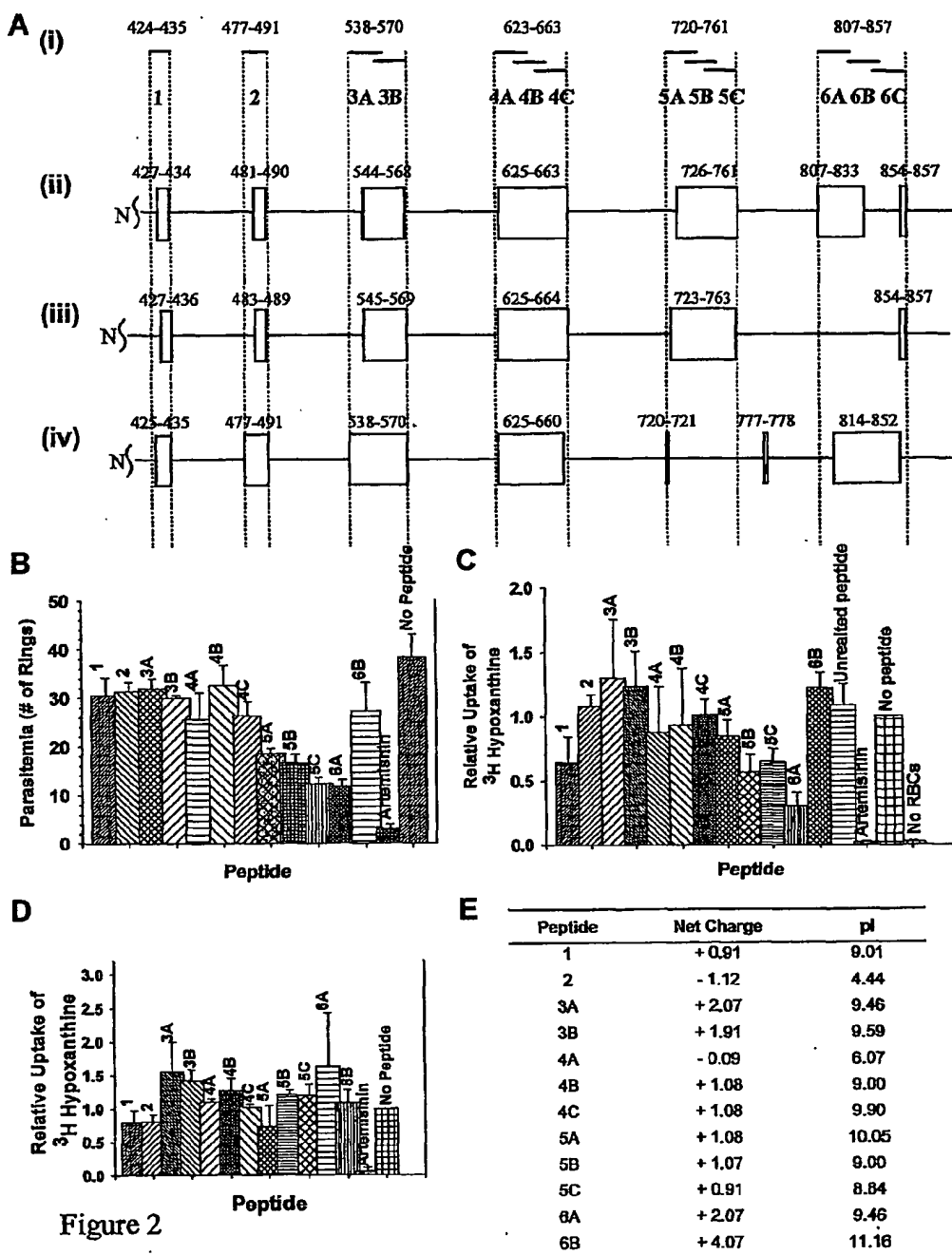


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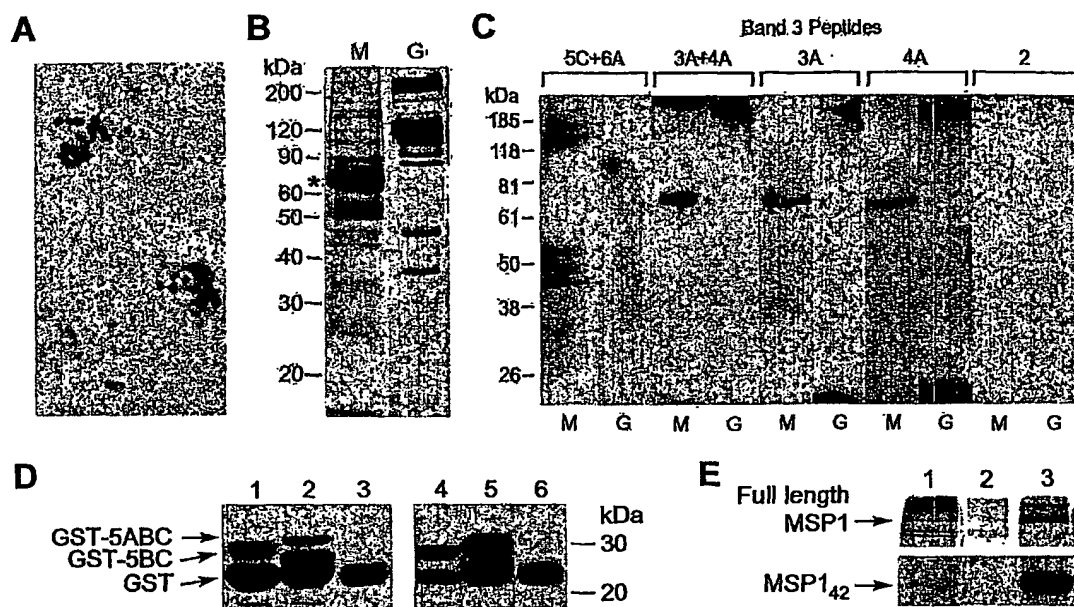
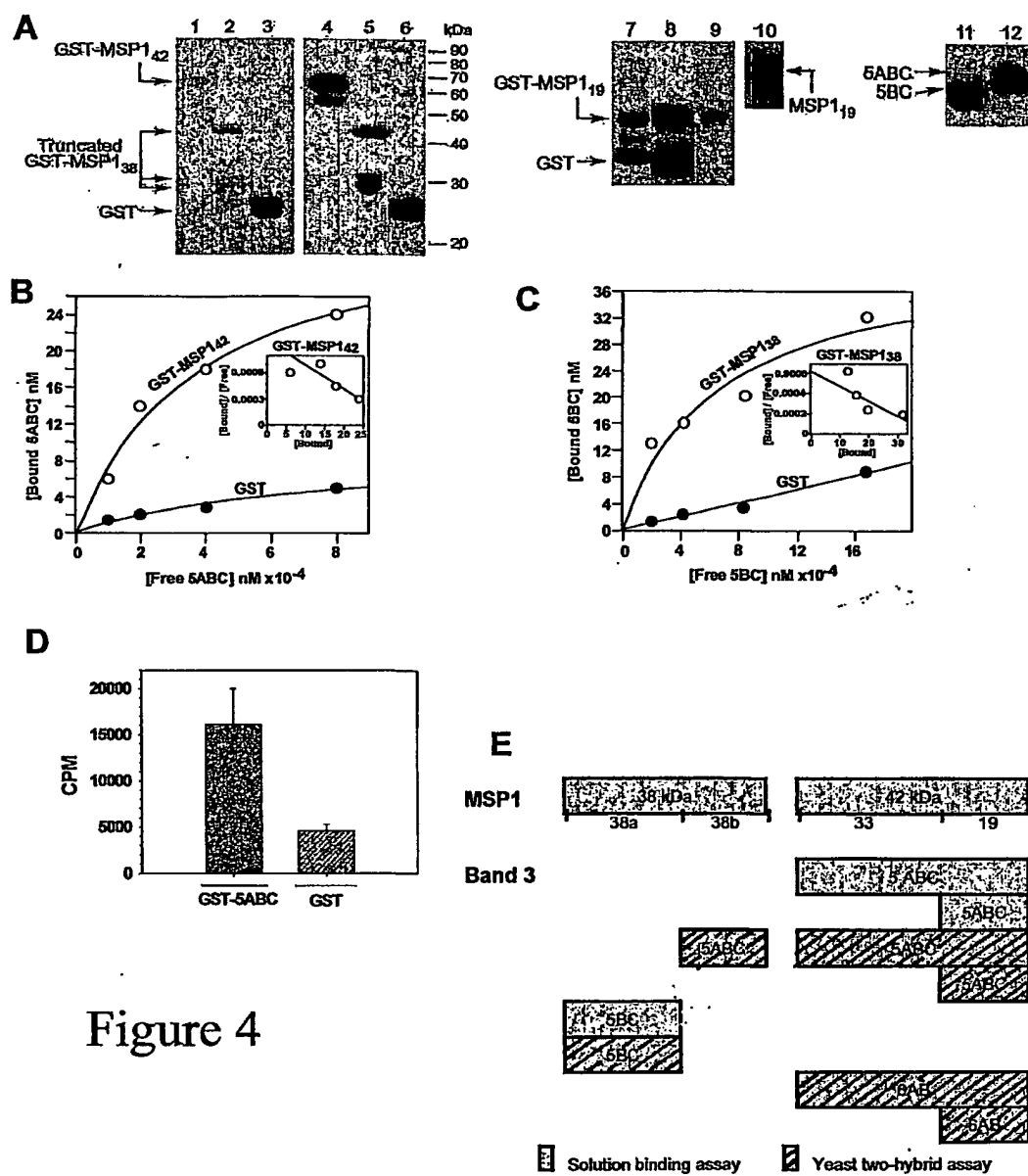


Figure 3



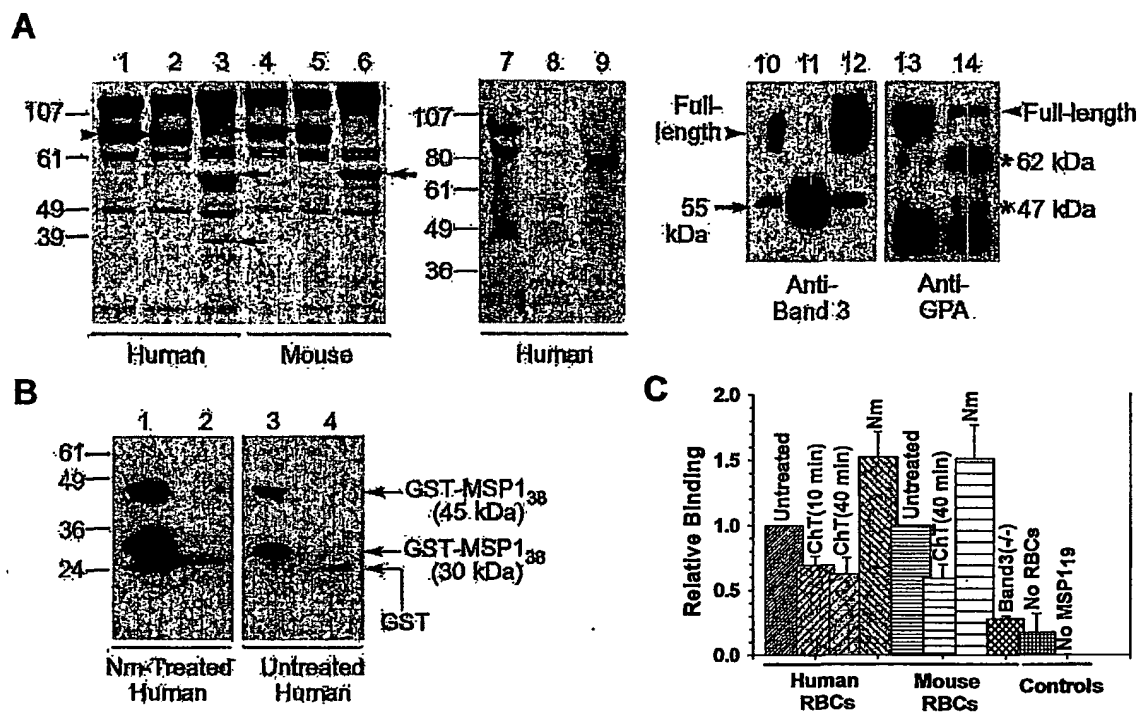


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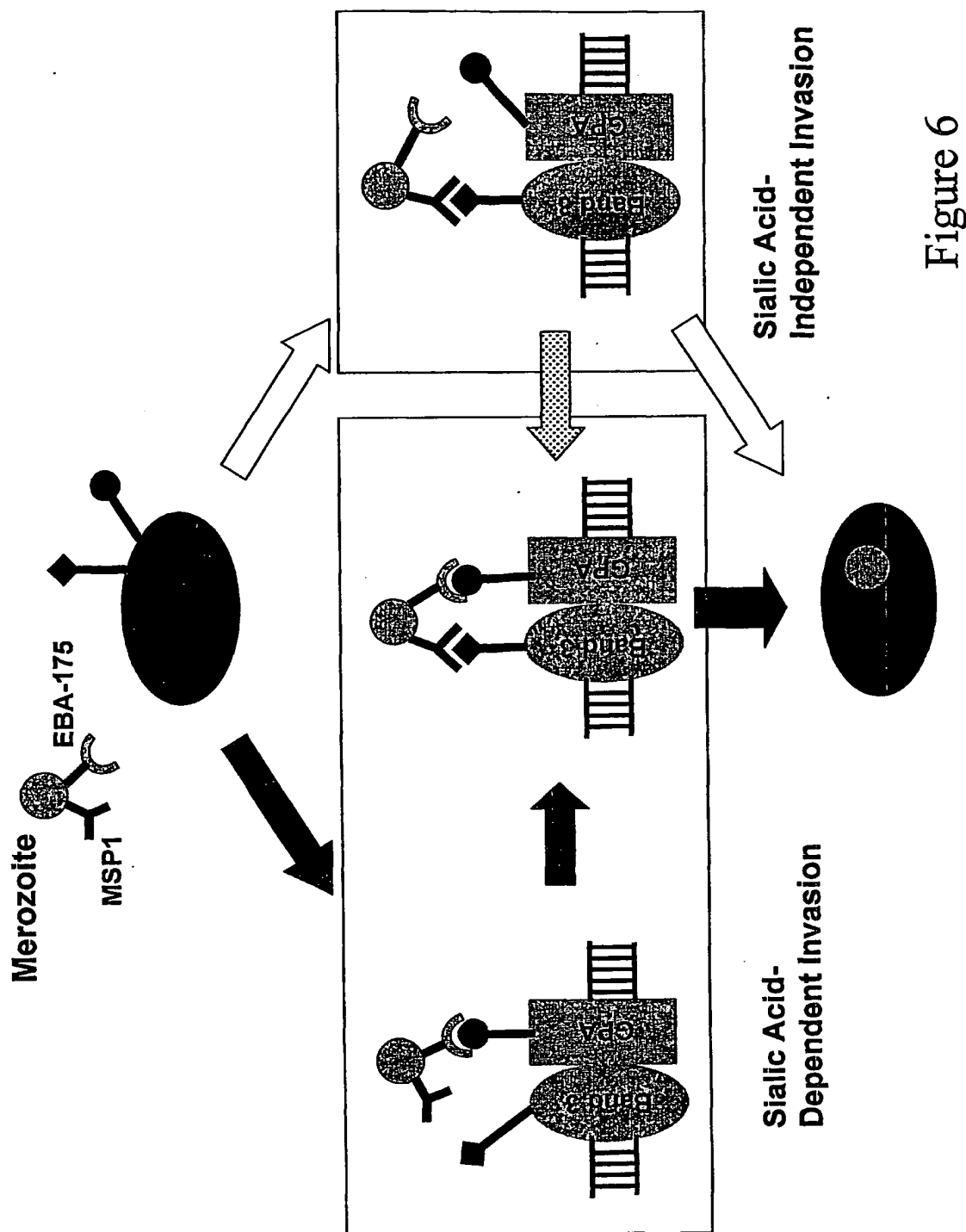


Figure 6

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Pro His Leu Ser His Leu Thr Phe Trp Ser Leu Leu Glu Leu Arg Arg
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Val Phe Thr Lys Gly Thr Val Leu Leu Asp Leu Gln Glu Thr Ser Leu
115         120         125
Ala Gly Val Ala Asn Gln Leu Leu Asp Arg Phe Ile Phe Glu Asp Gln
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Ile Arg Pro Gln Asp Arg Glu Glu Leu Leu Arg Ala Leu Leu Leu Lys
145         150         155         160
His Ser His Ala Gly Glu Leu Glu Ala Leu Gly Gly Val Lys Pro Ala
165         170         175
Val Leu Thr Arg Ser Gly Asp Pro Ser Gln Pro Leu Leu Pro Gln His
180         185         190
Ser Ser Leu Glu Thr Gln Leu Phe Cys Glu Gln Gly Asp Gly Gly Thr
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Glu Gly His Ser Pro Ser Gly Ile Leu Glu Lys Ile Pro Pro Asp Ser
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Glu Ala Thr Leu Val Leu Val Gly Arg Ala Asp Phe Leu Glu Gln Pro

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Ala Pro His Ile	Asp Tyr Thr Gln Leu Gly	Arg Ala Ala Ala	Thr Leu			
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Met Ser Glu Arg	Val Phe Arg Ile Asp	Ala Tyr Met Ala	Gln Ser Arg			
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Gly Glu Leu Leu	His Ser Leu Glu Gly	Phe Leu Asp Cys	Ser Leu Val			
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Pro Val Gln Arg	Glu Leu Leu Arg Arg	Arg Tyr Gln Ser	Ser Pro Ala			
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Lys Pro Asp Ser	Ser Phe Tyr Lys Gly	Leu Asp Leu Asn	Gly Gly Pro			
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Asp Ile Arg Arg	Arg Tyr Pro Tyr Tyr	Leu Ser Asp Ile	Thr Asp Ala			
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Phe Ser Pro Gln	Val Leu Ala Ala Val	Ile Phe Ile Tyr	Phe Ala Ala			
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Leu Ser Pro Ala	Ile Thr Phe Gly Gly	Leu Leu Gly Glu	Lys Thr Arg			
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Ile Leu Phe Ala	Leu Leu Gly Ala Gln	Pro Leu Leu Val	Val Gly Phe			
	450	455	460			
Ser Gly Pro Leu	Leu Val Phe Glu Glu	Ala Phe Phe Ser	Phe Cys Glu			
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Thr Asn Gly Leu	Glu Tyr Ile Val Gly	Arg Val Trp Ile	Gly Phe Trp			
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Leu Ile Leu Leu	Val Val Leu Val Val	Ala Phe Glu Gly	Ser Phe Leu			
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Val Arg Phe Ile	Ser Arg Tyr Thr Gln	Glu Ile Phe Ser	Phe Leu Ile			
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Ser Leu Ile Phe	Ile Tyr Glu Thr Phe	Ser Lys Leu Ile	Lys Ile Phe			
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Gln Asp His Pro	Leu Gln Lys Thr Tyr	Asn Tyr Asn Val	Leu Met Val			

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Leu Met Ala Gly Thr Phe Phe Phe Ala Met Met Leu Arg Lys Phe Lys						
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Asn Ser Ser Tyr Phe Pro Gly Lys Leu Arg Arg Val Ile Gly Asp Phe						
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Gln Asp Thr Tyr Thr Gln Lys Leu Ser Val Pro Asp Gly Phe Lys Val						
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Ser Asn Ser Ser Ala Arg Gly Trp Val Ile His Pro Leu Gly Leu Arg						
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Ser Glu Phe Pro Ile Trp Met Met Phe Ala Ser Ala Leu Pro Ala Leu						
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Leu Val Phe Ile Leu Ile Phe Leu Glu Ser Gln Ile Thr Thr Leu Ile						
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Val Ser Lys Pro Glu Arg Lys Met Val Lys Gly Ser Gly Phe His Leu						
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Met Pro Trp Leu Ser Ala Thr Thr Val Arg Ser Val Thr His Ala Asn						
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Ala Leu Thr Val Met Gly Lys Ala Ser Thr Pro Gly Ala Ala Ala Gln						
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Ile Gln Glu Val Lys Glu Gln Arg Ile Ser Gly Leu Leu Val Ala Val						
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Lys Tyr His Pro Asp Val Pro Tyr Val Lys Arg Val Lys Thr Trp Arg						
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Met His Leu Phe Thr Gly Ile Gln Ile Ile Cys Leu Ala Val Leu Trp						
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Val Val Lys Ser Thr Pro Ala Ser Leu Ala Leu Pro Phe Val Leu Ile						
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Leu Thr Val Pro Leu Arg Arg Val Leu Leu Pro Leu Ile Phe Arg Asn						

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 Trp Val Gln Leu Glu Glu Asn Leu Gly Glu Asn Gly Ala Trp Gly Arg
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Glu	Ala	Thr	Leu	Val	Leu	Val	Gly	Arg	Ala	Asp	Phe	Leu	Glu	Gln	Pro	225	230	235	240
Val	Leu	Gly	Phe	Val	Arg	Leu	Gln	Glu	Ala	Ala	Glu	Leu	Glu	Ala	Val	245	250	255	
Glu	Leu	Pro	Val	Pro	Ile	Arg	Phe	Leu	Phe	Val	Leu	Leu	Gly	Pro	Glu	260	265	270	
Ala	Pro	His	Ile	Asp	Tyr	Thr	Gln	Leu	Gly	Arg	Ala	Ala	Ala	Thr	Leu	275	280	285	
Met	Ser	Glu	Arg	Val	Phe	Arg	Ile	Asp	Ala	Tyr	Met	Ala	Gln	Ser	Arg	290	295	300	
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ccattagact	tattgtttta	tatacaaaat	aacatacctg	taatgtattc	tatgtttgat	2940
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aatttatata	aacttaagga	taatgacaaa	attaaaaatt	tattagagga	agcgaaaaaa	3060
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caggataaac	ccgaagtaag	tgcaaatgat	gatacatcac	attctacaaa	tttgaataat	3180
agttttaa	tatttgaaaa	catattgagt	cttggaaaaa	acaaaaatat	ataccaagaa	3240
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aatgatgaat	caaaaaggaa	gaaattagaa	gaagatatta	ataaattaaa	aaaaacttta	3420
cagttatcat	ttgatttata	taataaatat	aaattaaaaat	tagaaagatt	atttgataaa	3480
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ttagaatcaa	aattgaattc	acttaataac	caaagcatg	tattacaaaa	cttttctgtt	3600
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aaaatattat	tgaaacatta	taaaggactt	gttaaatatt	ataatggtga	atcatctcca	3720
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ggagaagcag	taactccttc	cgtaattgat	aacatacttt	ctaaaattga	aatgaatat	4260
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cgtgaaaatt	tcaaaaatgt	tttagaatca	gatttaattc	catataaaga	tttaacatca	4440
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gttcttggat	attataaaat	attatccgaa	aaatataaat	cagatttaga	ttcaattaaa	4620
aaatatatca	acgacaaaca	aggtgaaaat	gagaaatacc	ttcccttttt	aaacaatatt	4680
gagaccttat	ataaaacagt	taatgataaa	attgatattt	ttgtaattca	tttagaagca	4740
aaagttctaa	attatacata	tgagaaatca	aacgtagaag	ttaaaataaa	agaacttaat	4800

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tacttaaaaa caattcaaga caaattggca gattttaaaa aaaataacaa tttcgttgga 4860
 attgctgatt tatcaacaga ttataacccat aataacttat tgacaaagtt ccttagtaca 4920
 ggtatgggtt ttgaaaatct tgctaaaacc gttttatcta atttacttga tggaaacttg 4980
 caaggatgt taaacatttc acaacaccaa tgcgtaaaaa aacaatgtcc acaaaattct 5040
 ggatgtttca gacatttaga tgaaagagaa gaatgtaat gtttattaaa ttacaaacaa 5100
 gaaggatgata aatgtgttga aaatccaaat cctacttgta acgaaaataa tgggtggatgt 5160
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 tatatatata tatatatata taaaatatta cataatatat atatatatat ttagttatta 5460
 caggaatagt gatattttag tcatgttcaa aatatattaa aaaattataa atattataat 5520
 aaaaaaaaaa aaaaaaaaaa attatatact tataaattta tacatttata catatatata 5580
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 ataatatata tatttttttt tacgcataca taaaaagcat tttttttttt tataaacatt 5820
 ccaacaatta taaaataact ttaataataa cattaaattt ttattttttt ttttaaaaaa 5880
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<210> 10

<211> 1639

<212> PRT

<213> Plasmodium falciparum

<400> 10

Met Lys Ile Ile Phe Phe Leu Cys Ser Phe Leu Phe Phe Ile Ile Asn
 1 5 10 15

Thr Gln Cys Val Thr His Glu Ser Tyr Gln Glu Leu Val Lys Lys Leu
 20 25 30

Glu Ala Leu Glu Asp Ala Val Leu Thr Gly Tyr Ser Leu Phe Gln Lys
 35 40 45

Glu Lys Met Val Leu Asn Glu Gly Thr Ser Gly Thr Ala Val Thr Thr
 50 55 60

Ser Thr Pro Gly Ser Lys Gly Ser Val Ala Ser Gly Gly Ser Gly Gly

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65		70		75		80
Ser Val Ala Ser Gly Gly Ser Val Ala Ser Gly Gly Ser Val Ala Ser	85		90		95	
Gly Gly Ser Val Ala Ser Gly Gly Ser Gly Asn Ser Arg Arg Thr Asn	100		105		110	
Pro Ser Asp Asn Ser Ser Asp Ser Asp Ala Lys Ser Tyr Ala Asp Leu	115		120		125	
Lys His Arg Val Arg Asn Tyr Leu Leu Thr Ile Lys Glu Leu Lys Tyr	130		135		140	
Pro Gln Leu Phe Asp Leu Thr Asn His Met Leu Thr Leu Cys Asp Asn	145		150		155	160
Ile His Gly Phe Lys Tyr Leu Ile Asp Gly Tyr Glu Glu Ile Asn Glu		165		170		175
Leu Leu Tyr Lys Leu Asn Phe Tyr Phe Asp Leu Leu Arg Ala Lys Leu	180		185		190	
Asn Asp Val Cys Ala Asn Asp Tyr Cys Gln Ile Pro Phe Asn Leu Lys	195		200		205	
Ile Arg Ala Asn Glu Leu Asp Val Leu Lys Lys Leu Val Phe Gly Tyr	210		215		220	
Arg Lys Pro Leu Asp Asn Ile Lys Asp Asn Val Gly Lys Met Glu Asp	225		230		235	240
Tyr Ile Lys Lys Asn Lys Lys Thr Ile Glu Asn Ile Asn Glu Leu Ile		245		250		255
Glu Glu Ser Lys Lys Thr Ile Asp Lys Asn Lys Asn Ala Thr Lys Glu	260		265		270	
Glu Glu Lys Lys Lys Leu Tyr Gln Ala Gln Tyr Asp Leu Ser Ile Tyr	275		280		285	
Asn Lys Gln Leu Glu Glu Ala His Asn Leu Ile Ser Val Leu Glu Lys	290		295		300	
Arg Ile Asp Thr Leu Lys Lys Asn Glu Asn Ile Lys Glu Leu Leu Asp	305		310		315	320
Lys Ile Asn Glu Ile Lys Asn Pro Pro Pro Ala Asn Ser Gly Asn Thr		325		330		335
Pro Asn Thr Leu Leu Asp Lys Asn Lys Lys Ile Glu Glu His Glu Lys	340		345		350	
Glu Ile Lys Glu Ile Ala Lys Thr Ile Lys Phe Asn Ile Asp Ser Leu	355		360		365	
Phe Thr Asp Pro Leu Glu Leu Glu Tyr Tyr Leu Arg Glu Lys Asn Lys	370		375		380	
Asn Ile Asp Ile Ser Ala Lys Val Glu Thr Lys Glu Ser Thr Glu Pro						

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385		390		395		400
Asn Glu Tyr Pro	Asn Gly Val Thr Tyr	Pro Leu Ser Tyr	Asn Asp Ile			
	405	410	415			
Asn Asn Ala Leu	Asn Glu Leu Asn Ser	Phe Gly Asp Leu	Ile Asn Pro			
	420	425	430			
Phe Asp Tyr Thr	Lys Glu Pro Ser Lys	Asn Ile Tyr Thr	Asp Asn Glu			
	435	440	445			
Arg Lys Lys Phe	Ile Asn Glu Ile Lys	Glu Lys Ile Lys	Ile Glu Lys			
	450	455	460			
Lys Lys Ile Glu	Ser Asp Lys Lys Ser	Tyr Glu Asp Arg	Ser Lys Ser			
	465	470	475	480		
Leu Asn Asp Ile	Thr Lys Glu Tyr Glu	Lys Leu Leu Asn	Glu Ile Tyr			
	485	490	495			
Asp Ser Lys Phe	Asn Asn Asn Ile	Asp Leu Thr Asn	Phe Glu Lys Met			
	500	505	510			
Met Gly Lys Arg	Tyr Ser Tyr Lys	Val Glu Lys Leu	Thr His His Asn			
	515	520	525			
Thr Phe Ala Ser	Tyr Glu Asn Ser	Lys His Asn Leu	Glu Lys Leu Thr			
	530	535	540			
Lys Ala Leu Lys	Tyr Met Glu Asp	Tyr Ser Leu Arg	Asn Ile Val Val			
	545	550	555	560		
Glu Lys Glu Leu	Lys Tyr Tyr Lys	Asn Leu Ile Ser	Lys Ile Glu Asn			
	565	570	575			
Glu Ile Glu Thr	Leu Val Glu Asn	Ile Lys Lys Asp	Glu Glu Gln Leu			
	580	585	590			
Phe Glu Lys Lys	Ile Thr Lys Asp	Glu Asn Lys Pro	Asp Glu Lys Ile			
	595	600	605			
Leu Glu Val Ser	Asp Ile Val Lys	Val Gln Val Gln	Lys Val Leu Leu			
	610	615	620			
Met Asn Lys Ile	Asp Glu Leu Lys	Lys Thr Gln Leu	Ile Leu Lys Asn			
	625	630	635	640		
Val Glu Leu Lys	His Asn Ile His	Val Pro Asn Ser	Tyr Lys Gln Glu			
	645	650	655			
Asn Lys Gln Glu	Pro Tyr Tyr Leu	Ile Val Leu Lys	Lys Glu Ile Asp			
	660	665	670			
Lys Leu Lys Val	Phe Met Pro Lys	Val Glu Ser Leu	Ile Asn Glu Glu			
	675	680	685			
Lys Lys Asn Ile	Lys Thr Glu Gly	Gln Ser Asp Asn	Ser Glu Pro Ser			
	690	695	700			
Thr Glu Gly Glu	Ile Thr Gly Gln	Ala Thr Thr Lys	Pro Gly Gln Gln			

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705		710		715		720
Ala Gly Ser Ala	Leu Glu Gly Asp Ser Val Gln Ala Gln Ala Gln Glu					
	725			730		735
Gln Lys Gln Ala	Gln Pro Pro Val Pro Val Pro Val Pro Glu Ala Lys					
	740		745			750
Ala Gln Val Pro Thr Pro Pro Ala Pro Val Asn Asn Lys Thr Glu Asn						
	755		760			765
Val Ser Lys Leu Asp Tyr Leu Glu Lys Leu Tyr Glu Phe Leu Asn Thr						
	770		775			780
Ser Tyr Ile Cys His Lys Tyr Ile Leu Val Ser His Ser Thr Met Asn						
	785		790			800
Glu Lys Ile Leu Lys Gln Tyr Lys Ile Thr Lys Glu Glu Glu Ser Lys						
	805		810			815
Leu Ser Ser Cys Asp Pro Leu Asp Leu Leu Phe Asn Ile Gln Asn Asn						
	820		825			830
Ile Pro Val Met Tyr Ser Met Phe Asp Ser Leu Asn Asn Ser Leu Ser						
	835		840			845
Gln Leu Phe Met Glu Ile Tyr Glu Lys Glu Met Val Cys Asn Leu Tyr						
	850		855			860
Lys Leu Lys Asp Asn Asp Lys Ile Lys Asn Leu Leu Glu Glu Ala Lys						
	865		870			875
Lys Val Ser Thr Ser Val Lys Thr Leu Ser Ser Ser Ser Met Gln Pro						
	885		890			895
Leu Ser Leu Thr Pro Gln Asp Lys Pro Glu Val Ser Ala Asn Asp Asp						
	900		905			910
Thr Ser His Ser Thr Asn Leu Asn Asn Ser Leu Lys Leu Phe Glu Asn						
	915		920			925
Ile Leu Ser Leu Gly Lys Asn Lys Asn Ile Tyr Gln Glu Leu Ile Gly						
	930		935			940
Gln Lys Ser Ser Glu Asn Phe Tyr Glu Lys Ile Leu Lys Asp Ser Asp						
	945		950			955
Thr Phe Tyr Asn Glu Ser Phe Thr Asn Phe Val Lys Ser Lys Ala Asp						
	965		970			975
Asp Ile Asn Ser Leu Asn Asp Glu Ser Lys Arg Lys Lys Leu Glu Glu						
	980		985			990
Asp Ile Asn Lys Leu Lys Lys Thr Leu Gln Leu Ser Phe Asp Leu Tyr						
	995		1000			1005
Asn Lys Tyr Lys Leu Lys Leu Glu Arg Leu Phe Asp Lys Lys Lys						
	1010		1015			1020
Thr Val Gly Lys Tyr Lys Met Gln Ile Lys Lys Leu Thr Leu Leu						

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1025		1030		1035
Lys Glu Gln Leu Glu Ser	Lys	Leu Asn Ser Leu Asn	Asn Pro Lys	
1040	1045	1050		
His Val Leu Gln Asn Phe	Ser	Val Phe Phe Asn Lys	Lys Lys Glu	
1055	1060	1065		
Ala Glu Ile Ala Glu Thr	Glu	Asn Thr Leu Glu Asn	Thr Lys Ile	
1070	1075	1080		
Leu Leu Lys His Tyr Lys	Gly	Leu Val Lys Tyr Tyr	Asn Gly Glu	
1085	1090	1095		
Ser Ser Pro Leu Lys Thr	Leu	Ser Glu Glu Ser Ile	Gln Thr Glu	
1100	1105	1110		
Asp Asn Tyr Ala Ser Leu	Glu	Asn Phe Lys Val Leu	Ser Lys Leu	
1115	1120	1125		
Glu Gly Lys Leu Lys Asp	Asn	Leu Asn Leu Glu Lys	Lys Lys Leu	
1130	1135	1140		
Ser Tyr Leu Ser Ser Gly	Leu	His His Leu Ile Ala	Glu Leu Lys	
1145	1150	1155		
Glu Val Ile Lys Asn Lys	Asn	Tyr Thr Gly Asn Ser	Pro Ser Glu	
1160	1165	1170		
Asn Asn Thr Asp Val Asn	Asn	Ala Leu Glu Ser Tyr	Lys Lys Phe	
1175	1180	1185		
Leu Pro Glu Gly Thr Asp	Val	Ala Thr Val Val Ser	Glu Ser Gly	
1190	1195	1200		
Ser Asp Thr Leu Glu Gln	Ser	Gln Pro Lys Lys Pro	Ala Ser Thr	
1205	1210	1215		
His Val Gly Ala Glu Ser	Asn	Thr Ile Thr Thr Ser	Gln Asn Val	
1220	1225	1230		
Asp Asp Glu Val Asp Asp	Val	Ile Ile Val Pro Ile	Phe Gly Glu	
1235	1240	1245		
Ser Glu Glu Asp Tyr Asp	Asp	Leu Gly Gln Val Val	Thr Gly Glu	
1250	1255	1260		
Ala Val Thr Pro Ser Val	Ile	Asp Asn Ile Leu Ser	Lys Ile Glu	
1265	1270	1275		
Asn Glu Tyr Glu Val Leu	Tyr	Leu Lys Pro Leu Ala	Gly Val Tyr	
1280	1285	1290		
Arg Ser Leu Lys Lys Gln	Leu	Glu Asn Asn Val Met	Thr Phe Asn	
1295	1300	1305		
Val Asn Val Lys Asp Ile	Leu	Asn Ser Arg Phe Asn	Lys Arg Glu	
1310	1315	1320		
Asn Phe Lys Asn Val Leu	Glu	Ser Asp Leu Ile Pro	Tyr Lys Asp	

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1325	1330	1335
Leu Thr Ser Ser Asn Tyr Val	Val Lys Asp Pro Tyr Lys Phe Leu	
1340	1345	1350
Asn Lys Glu Lys Arg Asp Lys	Phe Leu Ser Ser Tyr Asn Tyr Ile	
1355	1360	1365
Lys Asp Ser Ile Asp Thr Asp	Ile Asn Phe Ala Asn Asp Val Leu	
1370	1375	1380
Gly Tyr Tyr Lys Ile Leu Ser	Glu Lys Tyr Lys Ser Asp Leu Asp	
1385	1390	1395
Ser Ile Lys Lys Tyr Ile Asn	Asp Lys Gln Gly Glu Asn Glu Lys	
1400	1405	1410
Tyr Leu Pro Phe Leu Asn Asn	Ile Glu Thr Leu Tyr Lys Thr Val	
1415	1420	1425
Asn Asp Lys Ile Asp Leu Phe	Val Ile His Leu Glu Ala Lys Val	
1430	1435	1440
Leu Asn Tyr Thr Tyr Glu Lys	Ser Asn Val Glu Val Lys Ile Lys	
1445	1450	1455
Glu Leu Asn Tyr Leu Lys Thr	Ile Gln Asp Lys Leu Ala Asp Phe	
1460	1465	1470
Lys Lys Asn Asn Asn Phe Val	Gly Ile Ala Asp Leu Ser Thr Asp	
1475	1480	1485
Tyr Asn His Asn Asn Leu Leu	Thr Lys Phe Leu Ser Thr Gly Met	
1490	1495	1500
Val Phe Glu Asn Leu Ala Lys	Thr Val Leu Ser Asn Leu Leu Asp	
1505	1510	1515
Gly Asn Leu Gln Gly Met Leu	Asn Ile Ser Gln His Gln Cys Val	
1520	1525	1530
Lys Lys Gln Cys Pro Gln Asn	Ser Gly Cys Phe Arg His Leu Asp	
1535	1540	1545
Glu Arg Glu Glu Cys Lys Cys	Leu Leu Asn Tyr Lys Gln Glu Gly	
1550	1555	1560
Asp Lys Cys Val Glu Asn Pro	Asn Pro Thr Cys Asn Glu Asn Asn	
1565	1570	1575
Gly Gly Cys Asp Ala Asp Ala	Lys Cys Thr Glu Glu Asp Ser Gly	
1580	1585	1590
Ser Asn Gly Lys Lys Ile Thr	Cys Glu Cys Thr Lys Pro Asp Ser	
1595	1600	1605
Tyr Pro Leu Phe Asp Gly Ile	Phe Cys Ser Ser Ser Asn Phe Leu	
1610	1615	1620
Gly Ile Ser Phe Leu Leu Ile	Leu Met Leu Ile Leu Tyr Ser Phe	

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1625 1630 1635

Ile

<210> 11
 <211> 378
 <212> PRT
 <213> Plasmodium falciparum

<400> 11

Gly Glu Ala Val Thr Pro Ser Val Ile Asp Asn Ile Leu Ser Lys Ile
 1 5 10 15

Glu Asn Glu Tyr Glu Val Leu Tyr Leu Lys Pro Leu Ala Gly Val Tyr
 20 25 30

Arg Ser Leu Lys Lys Gln Leu Glu Asn Asn Val Met Thr Phe Asn Val
 35 40 45

Asn Val Lys Asp Ile Leu Asn Ser Arg Phe Asn Lys Arg Glu Asn Phe
 50 55 60

Lys Asn Val Leu Glu Ser Asp Leu Ile Pro Tyr Lys Asp Leu Thr Ser
 65 70 75 80

Ser Asn Tyr Val Val Lys Asp Pro Tyr Lys Phe Leu Asn Lys Glu Lys
 85 90 95

Arg Asp Lys Phe Leu Ser Ser Tyr Asn Tyr Ile Lys Asp Ser Ile Asp
 100 105 110

Thr Asp Ile Asn Phe Ala Asn Asp Val Leu Gly Tyr Tyr Lys Ile Leu
 115 120 125

Ser Glu Lys Tyr Lys Ser Asp Leu Asp Ser Ile Lys Lys Tyr Ile Asn
 130 135 140

Asp Lys Gln Gly Glu Asn Glu Lys Tyr Leu Pro Phe Leu Asn Asn Ile
 145 150 155 160

Glu Thr Leu Tyr Lys Thr Val Asn Asp Lys Ile Asp Leu Phe Val Ile
 165 170 175

His Leu Glu Ala Lys Val Leu Asn Tyr Thr Tyr Glu Lys Ser Asn Val
 180 185 190

Glu Val Lys Ile Lys Glu Leu Asn Tyr Leu Lys Thr Ile Gln Asp Lys
 195 200 205

Leu Ala Asp Phe Lys Lys Asn Asn Asn Phe Val Gly Ile Ala Asp Leu
 210 215 220

Ser Thr Asp Tyr Asn His Asn Asn Leu Leu Thr Lys Phe Leu Ser Thr
 225 230 235 240

Gly Met Val Phe Glu Asn Leu Ala Lys Thr Val Leu Ser Asn Leu Leu
 245 250 255

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Asp Gly Asn Leu Gln Gly Met Leu Asn Ile Ser Gln His Gln Cys Val
 260 265 270
 Lys Lys Gln Cys Pro Gln Asn Ser Gly Cys Phe Arg His Leu Asp Glu
 275 280 285
 Arg Glu Glu Cys Lys Cys Leu Leu Asn Tyr Lys Gln Glu Gly Asp Lys
 290 295 300
 Cys Val Glu Asn Pro Asn Pro Thr Cys Asn Glu Asn Asn Gly Gly Cys
 305 310 315 320
 Asp Ala Asp Ala Lys Cys Thr Glu Glu Asp Ser Gly Ser Asn Gly Lys
 325 330 335
 Lys Ile Thr Cys Glu Cys Thr Lys Pro Asp Ser Tyr Pro Leu Phe Asp
 340 345 350
 Gly Ile Phe Cys Ser Ser Ser Asn Phe Leu Gly Ile Ser Phe Leu Leu
 355 360 365
 Ile Leu Met Leu Ile Leu Tyr Ser Phe Ile
 370 375

<210> 12
 <211> 360
 <212> PRT
 <213> Plasmodium falciparum

<400> 12

Gln Asp Lys Pro Glu Val Ser Ala Asn Asp Asp Thr Ser His Ser Thr
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 Asn Leu Asn Asn Ser Leu Lys Leu Phe Glu Asn Ile Leu Ser Leu Gly
 20 25 30
 Lys Asn Lys Asn Ile Tyr Gln Glu Leu Ile Gly Gln Lys Ser Ser Glu
 35 40 45
 Asn Phe Tyr Glu Lys Ile Leu Lys Asp Ser Asp Thr Phe Tyr Asn Glu
 50 55 60
 Ser Phe Thr Asn Phe Val Lys Ser Lys Ala Asp Asp Ile Asn Ser Leu
 65 70 75 80
 Asn Asp Glu Ser Lys Arg Lys Lys Leu Glu Glu Asp Ile Asn Lys Leu
 85 90 95
 Lys Lys Thr Leu Gln Leu Ser Phe Asp Leu Tyr Asn Lys Tyr Lys Leu
 100 105 110
 Lys Leu Glu Arg Leu Phe Asp Lys Lys Lys Thr Val Gly Lys Tyr Lys
 115 120 125
 Met Gln Ile Lys Lys Leu Thr Leu Leu Lys Glu Gln Leu Glu Ser Lys
 130 135 140
 Leu Asn Ser Leu Asn Asn Pro Lys His Val Leu Gln Asn Phe Ser Val
 145 150 155 160

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Phe Phe Asn Lys Lys Lys Glu Ala Glu Ile Ala Glu Thr Glu Asn Thr
 165 170 175
 Leu Glu Asn Thr Lys Ile Leu Leu Lys His Tyr Lys Gly Leu Val Lys
 180 185 190
 Tyr Tyr Asn Gly Glu Ser Ser Pro Leu Lys Thr Leu Ser Glu Glu Ser
 195 200 205
 Ile Gln Thr Glu Asp Asn Tyr Ala Ser Leu Glu Asn Phe Lys Val Leu
 210 215 220
 Ser Lys Leu Glu Gly Lys Leu Lys Asp Asn Leu Asn Leu Glu Lys Lys
 225 230 235 240
 Lys Leu Ser Tyr Leu Ser Ser Gly Leu His His Leu Ile Ala Glu Leu
 245 250 255
 Lys Glu Val Ile Lys Asn Lys Asn Tyr Thr Gly Asn Ser Pro Ser Glu
 260 265 270
 Asn Asn Thr Asp Val Asn Asn Ala Leu Glu Ser Tyr Lys Lys Phe Leu
 275 280 285
 Pro Glu Gly Thr Asp Val Ala Thr Val Val Ser Glu Ser Gly Ser Asp
 290 295 300
 Thr Leu Glu Gln Ser Gln Pro Lys Lys Pro Ala Ser Thr His Val Gly
 305 310 315 320
 Ala Glu Ser Asn Thr Ile Thr Thr Ser Gln Asn Val Asp Asp Glu Val
 325 330 335
 Asp Asp Val Ile Ile Val Pro Ile Phe Gly Glu Ser Glu Glu Asp Tyr
 340 345 350
 Asp Asp Leu Gly Gln Val Val Thr
 355 360

<210> 13

<211> 220

<212> PRT

<213> Plasmodium falciparum

<400> 13

Gln Asp Lys Pro Glu Val Ser Ala Asn Asp Asp Thr Ser His Ser Thr
 1 5 10 15
 Asn Leu Asn Asn Ser Leu Lys Leu Phe Glu Asn Ile Leu Ser Leu Gly
 20 25 30
 Lys Asn Lys Asn Ile Tyr Gln Glu Leu Ile Gly Gln Lys Ser Ser Glu
 35 40 45
 Asn Phe Tyr Glu Lys Ile Leu Lys Asp Ser Asp Thr Phe Tyr Asn Glu
 50 55 60
 Ser Phe Thr Asn Phe Val Lys Ser Lys Ala Asp Asp Ile Asn Ser Leu

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65		70		75		80									
Asn	Asp	Glu	Ser	Lys	Arg	Lys	Lys	Leu	Glu	Glu	Asp	Ile	Asn	Lys	Leu
				85					90					95	
Lys	Lys	Thr	Leu	Gln	Leu	Ser	Phe	Asp	Leu	Tyr	Asn	Lys	Tyr	Lys	Leu
			100					105					110		
Lys	Leu	Glu	Arg	Leu	Phe	Asp	Lys	Lys	Lys	Thr	Val	Gly	Lys	Tyr	Lys
		115					120					125			
Met	Gln	Ile	Lys	Lys	Leu	Thr	Leu	Leu	Lys	Glu	Gln	Leu	Glu	Ser	Lys
		130				135					140				
Leu	Asn	Ser	Leu	Asn	Asn	Pro	Lys	His	Val	Leu	Gln	Asn	Phe	Ser	Val
145					150					155					160
Phe	Phe	Asn	Lys	Lys	Lys	Glu	Ala	Glu	Ile	Ala	Glu	Thr	Glu	Asn	Thr
			165						170					175	
Leu	Glu	Asn	Thr	Lys	Ile	Leu	Leu	Lys	His	Tyr	Lys	Gly	Leu	Val	Lys
			180					185					190		
Tyr	Tyr	Asn	Gly	Glu	Ser	Ser	Pro	Leu	Lys	Thr	Leu	Ser	Glu	Glu	Ser
		195					200					205			
Ile	Gln	Thr	Glu	Asp	Asn	Tyr	Ala	Ser	Leu	Glu	Asn				
		210				215					220				

<210> 14

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

<221> misc_feature

<223> Synthetic Oligonucleotide

<400> 14

ctcgagctca ggataaaccc

20

<210> 15

<211> 18

<212> DNA

<213> Artificial Sequence

<220>

<221> misc_feature

<223> Synthetic Oligonucleotide

<400> 15

gcggccgcac ttgtagt

18

<210> 16

<211> 23

<212> DNA

-25-

<213> Artificial Sequence

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<221> misc_feature

<223> Synthetic Oligonucleotide

<400> 16

ctcgagctgg agaagcagta act

23

<210> 17

<211> 26

<212> DNA

<213> Artificial Sequence

<220>

<221> misc_feature

<223> Synthetic Oligonucleotide

<400> 17

gcggccgcac taaatgaaac tgtata

26

<210> 18

<211> 27

<212> DNA

<213> Artificial Sequence

<220>

<221> misc_feature

<223> Synthetic Oligonucleotide

<400> 18

ccgggatcca acatttcaca acaccaa

27

<210> 19

<211> 26

<212> DNA

<213> Artificial Sequence

<220>

<221> misc_feature

<223> Synthetic Oligonucleotide

<400> 19

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26

<210> 20

<211> 31

<212> DNA

<213> Artificial Sequence

<220>

<221> misc_feature

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<223> Synthetic Oligonucleotide

<400> 20

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31

<210> 21

<211> 33

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<223> Synthetic Oligonucleotide

<400> 21

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33

<210> 22

<211> 42

<212> PRT

<213> Homo sapien

<400> 22

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Asn	Ala	Leu	Thr	Val	Met	Gly	Lys	Ala	Ser	Thr	Pro	Gly	Ala	Ala	Ala
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Gln	Ile	Gln	Glu	Val	Lys	Glu	Gln	Arg	Ile
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<210> 23

<211> 51

<212> PRT

<213> Homo sapien

<400> 23

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Pro	Tyr	Val	Lys	Arg	Val	Lys	Thr	Trp	Arg	Met	His	Leu	Phe	Thr	Gly
		20					25						30		

Ile	Gln	Ile	Ile	Cys	Leu	Ala	Val	Leu	Trp	Val	Val	Lys	Ser	Thr	Pro
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Ala	Ser	Leu
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<211> 30

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<220>
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<210> 25
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<400> 26
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<400> 27
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<210> 28
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-28-

<400> 28
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<210> 29
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<400> 29
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<210> 30
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<400> 31
gccgaattcg cagtaactcc ttccg 25

<210> 32
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<223> Synthetic Oligonucleotide

<400> 32
gccggatcca atgaaactgt ataata 26

-29-

<210> 33
 <211> 334
 <212> PRT
 <213> Plasmodium falciparum
 <400> 33

Gln Asp Lys Pro Glu Val Ser Ala Asn Asp Asp Thr Ser His Ser Thr
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 20 25 30
 Lys Asn Lys Asn Ile Tyr Gln Glu Leu Ile Gly Gln Lys Ser Ser Glu
 35 40 45
 Asn Phe Tyr Glu Lys Ile Leu Lys Asp Ser Asp Thr Phe Tyr Asn Glu
 50 55 60
 Ser Phe Thr Asn Phe Val Lys Ser Lys Ala Asp Asp Ile Asn Ser Leu
 65 70 75 80
 Asn Asp Glu Ser Lys Arg Lys Lys Leu Glu Glu Asp Ile Asn Lys Leu
 85 90 95
 Lys Lys Thr Leu Gln Leu Ser Phe Asp Leu Tyr Asn Lys Tyr Lys Leu
 100 105 110
 Lys Leu Glu Arg Leu Phe Asp Lys Lys Lys Thr Val Gly Lys Tyr Lys
 115 120 125
 Met Gln Ile Lys Lys Leu Thr Leu Leu Lys Glu Gln Leu Glu Ser Lys
 130 135 140
 Leu Asn Ser Leu Asn Asn Pro Lys His Val Leu Gln Asn Phe Ser Val
 145 150 155 160
 Phe Phe Asn Lys Lys Lys Glu Ala Glu Ile Ala Glu Thr Glu Asn Thr
 165 170 175
 Leu Glu Asn Thr Lys Ile Leu Leu Lys His Tyr Lys Gly Leu Val Lys
 180 185 190
 Tyr Tyr Asn Gly Glu Ser Ser Pro Leu Lys Thr Leu Ser Glu Glu Ser
 195 200 205
 Ile Gln Thr Glu Asp Asn Tyr Ala Ser Leu Glu Asn Phe Lys Val Leu
 210 215 220
 Ser Lys Leu Glu Gly Lys Leu Lys Asp Asn Leu Asn Leu Glu Lys Lys
 225 230 235 240
 Lys Leu Ser Tyr Leu Ser Ser Gly Leu His His Leu Ile Ala Glu Leu
 245 250 255
 Lys Glu Val Ile Lys Asn Lys Asn Tyr Thr Gly Asn Ser Pro Ser Glu
 260 265 270

-30-

Asn Asn Thr Asp Val Asn Asn Ala Leu Glu Ser Tyr Lys Lys Phe Leu
 275 280 285

Pro Glu Gly Thr Asp Val Ala Thr Val Val Ser Glu Ser Gly Ser Asp
 290 295 300

Thr Leu Glu Gln Ser Gln Pro Lys Lys Pro Ala Ser Thr His Val Gly
 305 310 315 320

Ala Glu Ser Asn Thr Ile Thr Thr Ser Gln Asn Val Asp Asp
 325 330

<210> 34

<211> 376

<212> PRT

<213> Plasmodium falciparum

<400> 34

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 20 25 30

Leu Lys Lys Gln Leu Glu Asn Asn Val Met Thr Phe Asn Val Asn Val
 35 40 45

Lys Asp Ile Leu Asn Ser Arg Phe Asn Lys Arg Glu Asn Phe Lys Asn
 50 55 60

Val Leu Glu Ser Asp Leu Ile Pro Tyr Lys Asp Leu Thr Ser Ser Asn
 65 70 75 80

Tyr Val Val Lys Asp Pro Tyr Lys Phe Leu Asn Lys Glu Lys Arg Asp
 85 90 95

Lys Phe Leu Ser Ser Tyr Asn Tyr Ile Lys Asp Ser Ile Asp Thr Asp
 100 105 110

Ile Asn Phe Ala Asn Asp Val Leu Gly Tyr Tyr Lys Ile Leu Ser Glu
 115 120 125

Lys Tyr Lys Ser Asp Leu Asp Ser Ile Lys Lys Tyr Ile Asn Asp Lys
 130 135 140

Gln Gly Glu Asn Glu Lys Tyr Leu Pro Phe Leu Asn Asn Ile Glu Thr
 145 150 155 160

Leu Tyr Lys Thr Val Asn Asp Lys Ile Asp Leu Phe Val Ile His Leu
 165 170 175

Glu Ala Lys Val Leu Asn Tyr Thr Tyr Glu Lys Ser Asn Val Glu Val
 180 185 190

Lys Ile Lys Glu Leu Asn Tyr Leu Lys Thr Ile Gln Asp Lys Leu Ala
 195 200 205

Asp Phe Lys Lys Asn Asn Asn Phe Val Gly Ile Ala Asp Leu Ser Thr
 210 215 220

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Asp Tyr Asn His Asn Asn Leu Leu Thr Lys Phe Leu Ser Thr Gly Met
225 230 235 240

Val Phe Glu Asn Leu Ala Lys Thr Val Leu Ser Asn Leu Leu Asp Gly
245 250 255

Asn Leu Gln Gly Met Leu Asn Ile Ser Gln His Gln Cys Val Lys Lys
260 265 270

Gln Cys Pro Gln Asn Ser Gly Cys Phe Arg His Leu Asp Glu Arg Glu
275 280 285

Glu Cys Lys Cys Leu Leu Asn Tyr Lys Gln Glu Gly Asp Lys Cys Val
290 295 300

Glu Asn Pro Asn Pro Thr Cys Asn Glu Asn Asn Gly Gly Cys Asp Ala
305 310 315 320

Asp Ala Lys Cys Thr Glu Glu Asp Ser Gly Ser Asn Gly Lys Lys Ile
325 330 335

Thr Cys Glu Cys Thr Lys Pro Asp Ser Tyr Pro Leu Phe Asp Gly Ile
340 345 350

Phe Cys Ser Ser Ser Asn Phe Leu Gly Ile Ser Phe Leu Leu Ile Leu
355 360 365

Met Leu Ile Leu Tyr Ser Phe Ile
370 375

<210> 35

<211> 114

<212> PRT

<213> Plasmodium falciparum

<400> 35

Asn Ile Ser Gln His Gln Cys Val Lys Lys Gln Cys Pro Gln Asn Ser
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Gly Cys Phe Arg His Leu Asp Glu Arg Glu Glu Cys Lys Cys Leu Leu
20 25 30

Asn Tyr Lys Gln Glu Gly Asp Lys Cys Val Glu Asn Pro Asn Pro Thr
35 40 45

Cys Asn Glu Asn Asn Gly Gly Cys Asp Ala Asp Ala Lys Cys Thr Glu
50 55 60

Glu Asp Ser Gly Ser Asn Gly Lys Lys Ile Thr Cys Glu Cys Thr Lys
65 70 75 80

Pro Asp Ser Tyr Pro Leu Phe Asp Gly Ile Phe Cys Ser Ser Ser Asn
85 90 95

Phe Leu Gly Ile Ser Phe Leu Leu Ile Leu Met Leu Ile Leu Tyr Ser
100 105 110

Phe Ile

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<210> 36
 <211> 31
 <212> DNA
 <213> Artificial Sequence

<220>
 <221> misc_feature
 <223> Synthetic Oligonucleotide

<400> 36
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<210> 37
 <211> 33
 <212> DNA
 <213> Artificial Sequence

<220>
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 <223> Synthetic Oligonucleotide

<400> 37
 ccgggatcct tagatccgct gctctttgac etc 33

<210> 38
 <211> 1287
 <212> DNA
 <213> Plasmodium falciparum

<400> 38
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 gttataggaa tgttatgtat aaaattaagg gataaatatg aaggatatgc tgcttcaggt 180
 atacaaaaca ataattgtata tttaagaaat ttatcagagt tacaaaaggg aaatcaacct 240
 tgcttgagac atacaaacag aacggataat tcaaagatga acaaagtcaa aaataataat 300
 cagacagaaa ataattgacaa caaaaaaaag ctaggtaata aggaagataa ccagggaaaa 360
 aataaaaata ataataataa agaaaaacaa aatgacatta ataaaagagg aacacaaaat 420
 accgaaacta aaaaaagtaa taaaaaatta agtcaggact ataattgatgt aaataagaaa 480
 ttacaaaag aacaaatgaa aaatttagtt aattcattag atgaaattcc accccgaaac 540
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 aaaaaaaaaat taaaagaata tgaacaaaaa tatggaagat gctatgaaga gagaccaaat 660
 cgttttggat catatgaaca ggtgttaata agccagccac atgaatttaa tgaaagatta 720

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ggaactattt	atgataccag	taagaaagat	atgaagaaaa	ataaaaacaa	aaagcaaaat	960
ataaaacaag	gaggtaaaaa	ggaagaggta	aaacaagaag	gtaaaaagga	agaggtaaaa	1020
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ggtaaaaagg	aagaggtaaa	acaaggaggt	aaaaaggaag	aggtaaaaca	aggaggtaaa	1140
aaggaagagg	taaaacaagg	aggtaaaaag	gaagaggtaa	aacaaggagg	taaaaaggag	1200
gaggtaaaac	aaggaggtaa	aaaggaggag	gtaaaacaag	gaggtaaaaa	ggaagaggtg	1260
aaaaaagaat	taaaaaaaaa	caattaa				1287

<210> 39

<211> 3576

<212> DNA

<213> Plasmodium falciparum

<400> 39

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gaaatattaa	agaatgatta	taatacgcta	acagaaagtc	ataatataat	taatagaagg	180
tctagaaatt	taggagcgaa	tccagaatcc	attagtttag	gttatgaatt	aagtgaaaag	240
gatgaaggaa	ataaaaatga	tctaataaat	agtgctacag	atgtatcaac	agaattagag	300
aatttaaaag	aacgtttatt	tcctgaacta	gaattatata	caaacgatca	aaattcaaga	360
aataatactc	caaatttacg	taagggttct	ttgggatttg	atagttttaa	aaaattggaa	420
ttaggaacac	taaatcaatt	tgataaagat	aaaatgatta	atctgaaaga	tgaaaccaat	480
atgaatgaat	ttgaaggatt	tctaggaaga	aattcaatgg	ctagtaatgt	agttacatcc	540
gaattatttg	atgaaccagt	agatgatagt	agtagtacta	ctactagcac	aggtacaaaa	600
ttgcaaaacg	ttccatcgaa	tgataataac	ggtgaacttt	tgaaagatga	acctatagat	660
gattatataa	ataataattc	gaaagttgaa	tcggaagata	attattatgc	acaacagaat	720
atgcaaagtc	agtcgaaaga	taattatgct	tcagaacaaa	atgtagcaga	tcaatcgaca	780
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caagagtatt	ttgatagagg	tgaacaattg	aatgacgtaa	gtgcagataa	caatacaagt	900
aataaattga	aagacgaacc	tgtagataac	aatacaagta	ataaattgaa	agacgaacct	960
gtagataaca	atacaagtaa	taaattgaaa	gacgaacctg	tagatgacaa	tacaagtaat	1020

aaattgaaag acgaacctgt agataacaat acaattaata aattgaaaga cgaacctgta	1080
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ggatttaatc caatggaaca gagaagagaa gaagacaggg gacatatggg aggaagagg	2760
agtagatacc cagaagagga aagatataat tataacaata agagaagtaa tagtatacct	2820

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gaaagaataa gaagattaga ttatacagta tctgttaaag atatgtttat attatggaat 3240
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ttccatcaat tcttacgtaa aggttcttgt gaaaacgtg aatttttata ttttattaat 3480
tctaaaagaa aaggatgggc tgatcttacg gaaacaatga aaaatatatg gatggaaaga 3540
ttaacttata aaatgagaaa atatagtgga gcataa 3576

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<210> 40

<211> 903

<212> DNA

<213> Plasmodium falciparum

<400> 40

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gaacccataa ttcaaaataa aatagttgaa atacccaaag aagtatatct tgaaaagatt 180
gtagaagttc ctcaaataaa aactgtagaa agaatagtgg aacagataag gcccgttatt 240
aagtacaaaa atgtgtataa acccaaaatt gtatatgttg aaaaagttaa aaatgtagat 300
aaaattatat accaagagaa aattgttgaa gttccacaaa taaaactgt tgaaaaaatt 360
gtagaagtcc cagtatatgt taacagagaa agaattatta ctgttccaag atatatggtt 420
gtagaaaaag taataccctg attaaaaaca tccaaaagag aaagtataat ggaagttcca 480
gaagttaatt gcccacacat tgatataagt aaagaagtag aagataaaga agaaatacca 540
attaacgaat taaaagagaa ccaaacata agtcttgctg atgaaaaaga aatccaaata 600
ttaaatgact taactagcca aaaggtagat tctaatagcaa ccattaatat ggaaggtgaa 660
caagatacaa ctgtagatac tattacacaa gaaaacttct gtggaacagt tagttgtaat 720
ttcttaccaa attatccaaa cttctccaaa attggaaacc cattatgcaa aggaggtcca 780

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 taa 903

<210> 41
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 <213> Plasmodium falciparum

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 aaattaccaa attttatata taaagaacct attgtattca atactaataa aggagaaata 480
 gtttgcttaa tactcagctt tgctatagga ttgcgttggga tattttataa agacttcatt 540
 acacataacg ttttggcagt ttctttttgt tttcaagcca tatctttggg aattcttagc 600
 aactttttta taggattctt attattatct ggtttgtttg tatatgatat tttctggggt 660
 tttggaaacg atgttatggg tacagtagct aagctttttg aagctccagt aaaattgtta 720
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 ccaggaatat tgatgtcttt atgtttacgt tttgattatt atttatttaa gaataacata 840
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 gccatattag cttgttccat atgcaaaaga gaatttaa ataatgataa atatcaagaa 1080
 attacagaca aatccaatac tgtagatgat gcaagtaaga ataaaaaaaa agataaggaa 1140
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<210> 42
 <211> 3996
 <212> DNA
 <213> Plasmodium falciparum

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<400> 42

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ttaaaatgct attcaaattt gaatatcaac ttgaataagg ttccttatga tttggtctat	240
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aacatttttag gaaagattgg aaaagataca tatattaaaa gatgtagtag taattataac	1740

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aataaggaca	ataataataa	tgatggaaat	ggtagtagta	ataataataa	taatgatgat	2040
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<212> DNA
<213> Plasmodium falciparum

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<211> 2712
<212> DNA
<213> Plasmodium falciparum

<400> 44

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ttaaatagat	catgggtatc	tgaattttta	aaagaatatg	aagagccatt	tgtaaactct	300
gttatgaaat	ttctagataa	agagcaaaga	ttatTTTTt	catataactt	tggagatgta	360
gaaccacaag	gtaaatatac	atatttccca	gttaaggaat	ttcacaaata	ttgtatacta	420
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gaaaatgaaa	gtacaaatga	tatatctgtt	caatgcccaa	cttataacat	aaaattacat	660
tatactaaag	aatgtgctaa	tagtaataat	atattaaaat	gtattgatga	atttcttaga	720
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<210> 45

<211> 2232

<212> DNA

<213> Plasmodium falciparum

<400> 45

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aaaatgaata tgttaaaaga aaatgttgac tatattcaaa aaaatcaaaa cttattttaa 540
gaattaatga atcaaaaagc tacctactct tttgttaata ccaaaaaaaa aattatttct 600
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<210> 46

<211> 428

<212> PRT

<213> Plasmodium falciparum

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<400> 46

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Gly	Asp	Arg	Ser	Thr	Lys	Met	Lys	Gly	Lys	Ile	Cys	Ser	Ser	Tyr	Val	20	25	30	
Lys	Tyr	Ile	Cys	Leu	Thr	Ile	Cys	Val	Ile	Gly	Met	Leu	Cys	Ile	Lys	35	40	45	
Leu	Arg	Asp	Lys	Tyr	Glu	Gly	Tyr	Ala	Ala	Ser	Gly	Ile	Gln	Asn	Asn	50	55	60	
Asn	Val	Tyr	Leu	Arg	Asn	Leu	Ser	Glu	Leu	Gln	Lys	Gly	Asn	Gln	Pro	65	70	75	
Cys	Leu	Arg	His	Thr	Asn	Arg	Thr	Asp	Asn	Ser	Lys	Met	Asn	Lys	Val	85	90	95	
Lys	Asn	Asn	Asn	Gln	Thr	Glu	Asn	Asn	Asp	Asn	Lys	Lys	Lys	Leu	Gly	100	105	110	
Asn	Lys	Glu	Asp	Asn	Gln	Gly	Lys	Asn	Lys	Asn	Asn	Asn	Asn	Lys	Glu	115	120	125	
Lys	Gln	Asn	Asp	Ile	Asn	Lys	Arg	Gly	Thr	Gln	Asn	Thr	Glu	Thr	Lys	130	135	140	
Lys	Ser	Asn	Lys	Lys	Leu	Ser	Gln	Asp	Tyr	Asn	Asp	Val	Asn	Lys	Lys	145	150	155	
Phe	Thr	Lys	Glu	Gln	Met	Lys	Asn	Leu	Val	Asn	Ser	Leu	Asp	Glu	Ile	165	170	175	
Pro	Pro	Arg	Asn	Asp	Met	Glu	Lys	Ile	Trp	Asn	His	Ala	Val	Lys	Thr	180	185	190	
Ala	Asn	Ser	Gly	Thr	Ser	Arg	Ile	Lys	Lys	Lys	Leu	Lys	Glu	Tyr	Glu	195	200	205	
Gln	Lys	Tyr	Gly	Arg	Cys	Tyr	Glu	Glu	Arg	Pro	Asn	Arg	Phe	Gly	Ser	210	215	220	
Tyr	Glu	Gln	Val	Leu	Ile	Ser	Gln	Pro	His	Glu	Phe	Asn	Glu	Arg	Leu	225	230	235	
Lys	Val	His	Glu	Asn	Asp	Tyr	Thr	Val	Phe	Phe	Tyr	Glu	Leu	Leu	Asp	245	250	255	
Lys	Asp	Pro	Thr	Leu	Asp	Glu	Ile	Lys	Asn	Tyr	Ile	Thr	Ser	Phe	Leu	260	265	270	
Glu	Gly	Phe	Gln	Asn	Leu	Ile	Asp	Phe	Leu	Phe	Asn	Lys	Tyr	Lys	Ile	275	280	285	
Ile	Phe	Leu	Gln	Thr	Thr	Thr	Glu	Ile	Pro	Ile	Asp	Gly	Thr	Ile	Tyr	290	295	300	

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Asp Thr Ser Lys Lys Asp Met Lys Lys Asn Lys Asn Lys Lys Gln Asn
305 310 315 320

Ile Lys Gln Gly Gly Lys Lys Glu Glu Val Lys Gln Glu Gly Lys Lys
325 330 335

Glu Glu Val Lys Gln Glu Gly Lys Lys Glu Glu Val Lys Gln Glu Gly
340 345 350

Lys Lys Glu Glu Val Lys Gln Glu Gly Lys Lys Glu Glu Val Lys Gln
355 360 365

Gly Gly Lys Lys Glu Glu Val Lys Gln Gly Gly Lys Lys Glu Glu Val
370 375 380

Lys Gln Gly Gly Lys Lys Glu Glu Val Lys Gln Gly Gly Lys Lys Glu
385 390 395 400

Glu Val Lys Gln Gly Lys Lys Glu Glu Val Lys Gln Gly Gly Lys
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Lys Glu Glu Val Lys Lys Glu Leu Lys Lys Asn Asn
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<210> 47

<211> 1191

<212> PRT

<213> Plasmodium falciparum

<400> 47

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Asn Lys Arg Asn Lys Tyr Asn Val Glu Ile Leu Lys Asn Asp Tyr Asn
35 40 45

Thr Leu Thr Glu Ser His Asn Ile Ile Asn Arg Arg Ser Arg Asn Leu
50 55 60

Gly Ala Asn Pro Glu Ser Ile Ser Leu Gly Tyr Glu Leu Ser Glu Lys
65 70 75 80

Asp Glu Gly Asn Lys Asn Asp Leu Ile Asn Ser Ala Thr Asp Val Ser
85 90 95

Thr Glu Leu Glu Asn Leu Lys Glu Arg Leu Phe Pro Glu Leu Glu Leu
100 105 110

Tyr Thr Asn Asp Gln Asn Ser Arg Asn Asn Thr Pro Asn Leu Arg Lys
115 120 125

Gly Ser Leu Gly Phe Asp Ser Phe Lys Lys Leu Glu Leu Gly Thr Leu
130 135 140

Asn Gln Phe Asp Lys Asp Lys Met Ile Asn Leu Lys Asp Glu Thr Asn
145 150 155 160

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 Val Val Thr Ser Glu Leu Phe Asp Glu Pro Val Asp Asp Ser Ser Ser
 180 185 190
 Thr Thr Thr Ser Thr Gly Thr Lys Leu Gln Asn Val Pro Ser Asn Asp
 195 200 205
 Asn Asn Gly Glu Leu Leu Lys Asp Glu Pro Ile Asp Asp Tyr Ile Asn
 210 215 220
 Asn Asn Ser Lys Val Glu Ser Glu Asp Asn Tyr Tyr Ala Gln Gln Asn
 225 230 235 240
 Met Gln Ser Gln Ser Lys Asp Asn Tyr Ala Ser Glu Gln Asn Val Ala
 245 250 255
 Asp Gln Ser Thr Asp Asn Tyr Pro Thr Gln His Asp Val Pro Val Gln
 260 265 270
 Leu Arg Asp Asn Tyr Ala Ser Glu Gln Glu Tyr Phe Asp Arg Gly Glu
 275 280 285
 Gln Leu Asn Asp Val Ser Ala Asp Asn Asn Thr Ser Asn Lys Leu Lys
 290 295 300
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 305 310 315 320
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 340 345 350
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 355 360 365
 Lys Asp Glu Pro Val Asp Asp His Ala Gly Lys His Leu Lys Asp Glu
 370 375 380
 Pro Val Asp Asp His Ala Gly Lys His Met Lys Asp Glu Pro Val Asp
 385 390 395 400
 Ile Asp Arg Thr Asn Ile Lys Lys Gly Leu Asn Glu Gln His Val Asn
 405 410 415
 Pro Trp Thr Thr Thr Leu Ala Asp Leu Lys Asn Ile Asn Asn Ser Met
 420 425 430
 Lys Ile Glu Lys Asn Asn Lys Ser Asn Glu Gln Val Lys Asn Thr Ser
 435 440 445
 Val Ser Lys Ser Cys Asp Ile Ile Lys Pro Ser Lys Phe Asn Lys Lys
 450 455 460
 Asn Leu Phe Glu Gln Arg Leu Gln Ser Val Glu Gly Lys Asn Phe Phe
 465 470 475 480

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Glu Gly Arg Ser Gln Asn Leu Glu Gly Arg Ser Asn Phe Asp Glu Arg
 485 490 495
 Ser Gln Ile Val Glu Gln Arg Arg Asn Phe Asp Asp Arg Asp Gln Asn
 500 505 510
 Ile Met Asp Arg Lys Asn Phe Asp Glu Arg Asn Gln Gln Val Asn Asp
 515 520 525
 Arg Arg Asn Phe Asp Glu Arg Asn Gln Gln Val Asn Asp Arg Arg Asn
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 580 585 590
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 595 600 605
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 625 630 635 640
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 Asn Asp Arg Arg Asn Phe Asp Glu Arg Asn Gln His Val Asn Asp Arg
 660 665 670
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 740 745 750
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 755 760 765
 Asp Arg Arg Asn Phe Asp Glu Arg Asn Gln His Val Asn Glu Arg Arg
 770 775 780
 Asn Phe Asp Glu Arg Asn Gln His Val Asn Glu Arg Tyr Gln Asn Val
 785 790 795 800

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 820 825 830
 Asp Pro Arg Asn Pro Asn Ile Pro Tyr Val Arg Phe Pro His His Gln
 835 840 845
 Trp Gly Gln Gly Met Met Tyr Gly Arg Pro Tyr Tyr Pro Trp Val Pro
 850 855 860
 Phe Met Gly Asp Gly Arg Gly Tyr Asn Phe Tyr Asn Pro His Gln His
 865 870 875 880
 Met Val Tyr Gly Arg Pro Tyr Tyr Trp Val Pro Pro Pro Pro Ala Leu
 885 890 895
 Glu Tyr Thr Lys Gly Phe Asn Pro Met Glu Gln Arg Arg Glu Glu Asp
 900 905 910
 Arg Gly His Met Gly Gly Arg Gly Ser Arg Tyr Pro Glu Glu Glu Arg
 915 920 925
 Tyr Asn Tyr Asn Asn Lys Arg Ser Asn Ser Ile Pro Glu Gly Arg Asn
 930 935 940
 Tyr Glu Glu Asn Ala Tyr Glu Arg Gly Gly Gly Asn Asn Lys Trp Asp
 945 950 955 960
 Phe Arg Asn Met Tyr Asp Arg Leu Arg Asp Glu Asp Glu Asn Asp Tyr
 965 970 975
 Asp Gln Pro Pro Ser Thr Ser Ser Ser Asn Arg Gly Arg Gly Asn Glu
 980 985 990
 Arg Tyr Ser Gln Ser Arg Asp Arg Arg Glu Glu Arg Asn Asn Tyr Asn
 995 1000 1005
 Ser Asp Tyr Tyr Thr Arg Gly Asn Glu Arg Thr Tyr Asn Asn Ser
 1010 1015 1020
 Asn Val Thr Ser Ser Ser Asn Arg Glu Leu Ile Pro Tyr Lys Lys
 1025 1030 1035
 Glu Ile Leu Pro Phe Gly Val Ser Asn Ser Glu Leu Glu Asp Lys
 1040 1045 1050
 Leu Thr Glu Glu Glu Leu Asn Glu Arg Ile Arg Arg Leu Asp Tyr
 1055 1060 1065
 Thr Val Ser Val Lys Asp Met Phe Ile Leu Trp Asn His Ile Leu
 1070 1075 1080
 Ala His Glu Arg Lys Lys Tyr Thr Lys Met Gln Glu Tyr Leu Met
 1085 1090 1095
 Tyr Tyr Ser Gln Tyr Leu Glu Lys Thr Tyr Leu Val Pro Thr Ala
 1100 1105 1110

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Phe Arg Lys Lys Tyr Trp Trp Arg Val His Tyr Met Leu Thr Glu
1115 1120 1125

Glu Val Val Lys Arg Glu Arg Thr Asp Asn Leu Asp Phe His Gln
1130 1135 1140

Phe Leu Arg Lys Gly Ser Cys Glu Lys Arg Glu Phe Leu Tyr Phe
1145 1150 1155

Ile Asn Ser Lys Arg Lys Gly Trp Ala Asp Leu Thr Glu Thr Met
1160 1165 1170

Lys Asn Ile Trp Met Glu Arg Leu Thr Tyr Lys Met Arg Lys Tyr
1175 1180 1185

Ser Gly Ala
1190

<210> 48

<211> 300

<212> PRT

<213> Plasmodium falciparum

<400> 48

Met Cys Ser Thr Asn Lys Asn Leu Ala Cys Cys Lys Gly Asp Asn Val
1 5 10 15

Phe Asp Gly Gln Ile Asn Gly Asn Glu Ser Tyr Pro Gln Val Val Asn
20 25 30

Lys Gln Leu Pro Pro Lys Val Leu Glu Pro Ile Ile Gln Asn Lys Ile
35 40 45

Val Glu Ile Pro Lys Glu Val Tyr Leu Glu Lys Ile Val Glu Val Pro
50 55 60

Gln Ile Lys Thr Val Glu Arg Ile Val Glu Gln Ile Arg Pro Val Ile
65 70 75 80

Lys Tyr Lys Asn Val Tyr Lys Pro Lys Ile Val Tyr Val Glu Lys Val
85 90 95

Lys Asn Val Asp Lys Ile Ile Tyr Gln Glu Lys Ile Val Glu Val Pro
100 105 110

Gln Ile Lys Thr Val Glu Lys Ile Val Glu Val Pro Val Tyr Val Asn
115 120 125

Arg Glu Arg Ile Ile Thr Val Pro Arg Tyr Met Val Val Glu Lys Val
130 135 140

Ile Pro Val Leu Lys Thr Ser Lys Arg Glu Ser Ile Met Glu Val Pro
145 150 155 160

Glu Val Asn Cys Pro His Ile Asp Ile Ser Lys Glu Val Glu Asp Lys
165 170 175

Glu Glu Ile Pro Ile Asn Glu Leu Lys Glu Asn Gln Thr Ile Ser Leu

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180										185					190				
Ala	Asp	Glu	Lys	Glu	Ile	Gln	Ile	Leu	Asn	Asp	Leu	Thr	Ser	Gln	Lys				
		195						200					205						
Val	Asp	Ser	Asn	Ala	Thr	Ile	Asn	Met	Glu	Gly	Glu	Gln	Asp	Thr	Thr				
		210				215						220							
Val	Asp	Thr	Ile	Thr	Gln	Glu	Asn	Phe	Cys	Gly	Thr	Val	Ser	Cys	Asn				
		225			230					235					240				
Phe	Leu	Pro	Asn	Tyr	Pro	Asn	Phe	Ser	Lys	Ile	Gly	Asn	Pro	Leu	Cys				
				245					250						255				
Lys	Gly	Gly	Pro	Glu	Lys	Glu	Lys	Arg	Phe	Ser	Ser	Ile	Ser	Ile	Tyr				
			260					265						270					
Lys	Ser	Lys	Asp	Ser	Gly	Phe	Pro	Ser	Ile	Arg	Ile	Ala	Lys	Thr	Pro				
			275				280					285							
Gln	Met	Phe	Gln	Arg	Asn	Leu	Tyr	Cys	Ser	Tyr	Ala								
		290				295					300								

<210> 49

<211> 400

<212> PRT

<213> Plasmodium falciparum

<400> 49

Met	Lys	Asn	Glu	Asn	Met	Gly	Asn	Ser	Ile	Phe	Tyr	Tyr	Ser	Cys	Tyr			
1				5					10					15				
Val	Ile	Ile	Val	Leu	Thr	Ile	Ile	Leu	Ser	Lys	Phe	Val	Val	Ile	Pro			
			20					25					30					
Leu	Met	Ala	Gln	Met	Phe	Leu	Tyr	Thr	Phe	Ile	Thr	Ile	Tyr	Ile	Gly			
		35					40					45						
Ser	His	Asp	Ser	Leu	Lys	Gln	Leu	Glu	Ile	Asp	Asp	Lys	Thr	Lys	Lys			
	50					55					60							
Ser	Asp	Asn	Ile	Thr	Ala	Tyr	Asp	Ala	Met	Met	Phe	Pro	Val	Ile	Gly			
65					70					75					80			
Ser	Ala	Ala	Leu	Leu	Thr	Leu	Tyr	Phe	Ala	Tyr	Lys	Phe	Leu	Asp	Pro			
				85					90					95				
Phe	Tyr	Val	Asn	Leu	Leu	Leu	Thr	Leu	Tyr	Leu	Thr	Leu	Ala	Gly	Val			
			100					105					110					
Phe	Ser	Leu	Gln	Gly	Val	Phe	Thr	Thr	Ile	Leu	Glu	Pro	Val	Phe	Pro			
		115					120					125						
Asn	Phe	Phe	Lys	Lys	Asp	Glu	Tyr	Val	Lys	Thr	Phe	Lys	Leu	Pro	Asn			
		130				135					140							
Phe	Ile	Tyr	Lys	Glu	Pro	Ile	Val	Phe	Asn	Thr	Asn	Lys	Gly	Glu	Ile			
145					150					155					160			

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Val Cys Leu Ile Leu Ser Phe Ala Ile Gly Leu Arg Trp Ile Phe Tyr
165 170 175

Lys Asp Phe Ile Thr His Asn Val Leu Ala Val Ser Phe Cys Phe Gln
180 185 190

Ala Ile Ser Leu Val Ile Leu Ser Asn Phe Leu Ile Gly Phe Leu Leu
195 200 205

Leu Ser Gly Leu Phe Val Tyr Asp Ile Phe Trp Val Phe Gly Asn Asp
210 215 220

Val Met Val Thr Val Ala Lys Ser Phe Glu Ala Pro Val Lys Leu Leu
225 230 235 240

Phe Pro Val Ser Ser Asp Pro Val His Tyr Ser Met Leu Gly Leu Gly
245 250 255

Asp Ile Ile Ile Pro Gly Ile Leu Met Ser Leu Cys Leu Arg Phe Asp
260 265 270

Tyr Tyr Leu Phe Lys Asn Asn Ile His Lys Gly Asn Leu Lys Lys Met
275 280 285

Phe Asn Asp Ile Ser Ile His Glu Ser Phe Lys Lys Tyr Tyr Phe Tyr
290 295 300

Thr Ile Ile Ile Phe Tyr Glu Leu Gly Leu Val Val Thr Tyr Cys Met
305 310 315 320

Leu Phe Tyr Phe Glu His Pro Gln Pro Ala Leu Leu Tyr Leu Val Pro
325 330 335

Ala Cys Ile Leu Ala Ile Leu Ala Cys Ser Ile Cys Lys Arg Glu Phe
340 345 350

Lys Leu Met Ile Lys Tyr Gln Glu Ile Thr Asp Lys Ser Asn Thr Val
355 360 365

Asp Asp Ala Ser Lys Asn Lys Lys Lys Asp Lys Glu Glu Ile Pro Lys
370 375 380

Ile Gln Glu Thr Pro Val Ser Asn Ala Lys Lys Arg Ile Thr Asn Lys
385 390 395 400

<210> 50

<211> 1331

<212> PRT

<213> Plasmodium falciparum

<400> 50

Met Val Leu Val Val Glu Tyr His Asn Ile Asn Thr Pro Val Gly Lys
1 5 10 15

Tyr Ser Glu Leu Glu Asn Leu Lys Glu Glu Lys Glu Lys Arg Leu Tyr
20 25 30

Asn Asn Leu Glu Tyr Val Asn Leu Leu Asp Ile Arg Thr Leu Glu Asn
35 40 45

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Lys Ser Ile Tyr Val Ser Ser Asp Leu Leu Asn Phe Leu Lys Cys Tyr
 50 55 60
 Ser Asn Leu Asn Ile Asn Leu Asn Lys Val Pro Tyr Asp Leu Val Tyr
 65 70 75 80
 Ser Phe Leu Leu Asp Gly Glu Leu Tyr Leu Gly Tyr Asp Ile Ser Val
 85 90 95
 Phe Ile Leu Leu Val Lys Ala Glu His Phe Glu Tyr Cys Arg Arg Ile
 100 105 110
 Asp Asn Glu Asn Ser Asp Lys Lys Glu Ser Phe Arg Thr Lys Asn Lys
 115 120 125
 Ser Thr Ile Lys Arg Ser Ser Gln Ile Asp Asp Glu Asp Asn Leu Gln
 130 135 140
 Gly Leu Leu Ile Lys Glu Lys Glu Asp Tyr Leu Ser Phe Leu Asn Glu
 145 150 155 160
 Asn Asn Glu Ala Leu Lys Gln Tyr Met Glu Ser Glu Lys Arg Gly Asn
 165 170 175
 Pro Leu Trp His Leu Asp Glu Ser Lys Tyr Met Asp Lys Asp Trp Tyr
 180 185 190
 Asp Glu Glu Asp Ser Ser Phe Ile Phe Lys Pro Thr Phe Asn Tyr Leu
 195 200 205
 Gly Lys Asn Asn Asn Asn Asn Asn Asn His Asn Asn Asn Asn Ala Phe
 210 215 220
 Ser Asn Phe Val Met Gly Asn Leu Ser Ser Asp Asn Ile Ser Gly Cys
 225 230 235 240
 Phe Phe Val Glu Lys Leu Asn Ala Tyr Leu Phe Ala Met Leu Asp Lys
 245 250 255
 Cys Ser Asn Lys Thr Val Ile Ser Val Phe Pro Tyr Glu Lys Phe Gly
 260 265 270
 Arg His Glu Ser Arg Asn Leu Ala Ile Gln Phe Ser Gln Tyr Glu Asp
 275 280 285
 Tyr Met His Arg Ile Ile Glu Asp Arg Leu Tyr Ala Asn Ile Gln Asn
 290 295 300
 Asn Leu Pro Ser Val His Asn Met Lys Asn Met Ser Asn Met Asn Asn
 305 310 315 320
 Ile Asn Asn Asn Asn Lys Asp Ile Ile Ile Asn Arg Ser Gly Ile Ser
 325 330 335
 Asn Gly Asn Ser Gln Ser Val Pro Cys Phe Glu Asn Ile Leu Asp Tyr
 340 345 350
 Asp Lys Leu Lys Phe Val Glu Tyr Ile Asn Ser Phe Ser Asp Val Lys
 355 360 365

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Lys Ser Ser Ser Phe Asp Ile Ile Gly Ser Ser Lys Asn Ile Tyr Glu
 370 375 380
 Gln Gly Glu Asn Leu Lys Asn Tyr Cys Ile Tyr His Asn Asn Asn Phe
 385 390 395 400
 Glu Ser Gly Phe Glu Asn Tyr Ile Leu Glu Asn Lys Gln Pro Leu Glu
 405 410 415
 Leu Ile Glu Asn His Phe Asp Ile Met Glu Asn Ile Lys Gly Met Tyr
 420 425 430
 Asp Asn Thr Asn Gln Glu Glu Met Asn Phe Asn Asn Val Ser Gly Leu
 435 440 445
 Leu Arg Glu Asp Asn Ser Asn Met Asn Glu Ile Tyr Leu Thr Arg Asp
 450 455 460
 Asn His Asn Asn Asn Tyr His Glu Asn Glu Asn Ile Tyr Ser Ile
 465 470 475 480
 Asn Ile Lys Tyr Ile Asn Asn His Phe Asn Asn Lys Asp Asp Met Ile
 485 490 495
 Met Lys Cys Lys Asn Met Lys Gly Ser Ile Ser Met Asp Asn Asn Ser
 500 505 510
 Ser Asn Ser Asn Ser Asn Asn Thr His Phe Glu Lys Thr Leu Glu Ser
 515 520 525
 Ile Asn Pro Asp Asp His Asn Ile Phe Asn Ser Glu Met Asp Ser Met
 530 535 540
 Lys Asn Glu Asn Asn Asp Glu Glu Glu Gln Thr Ala Thr Ser Ile Tyr
 545 550 555 560
 Asn Ile Leu Gly Lys Ile Gly Lys Asp Thr Tyr Ile Lys Arg Cys Ser
 565 570 575
 Ser Asn Tyr Asn Tyr Asp Asn Asn Asn Gly Tyr Ser Asn Glu Ser Ser
 580 585 590
 Asp Asn Tyr Asn Asn Gly Tyr Asn Asp Ser Thr Asp Asn Asn Asn Gly
 595 600 605
 Tyr Asn Ser Asn Ser Ser Tyr Asn Ser Asn Asn Asn Glu Asp Asp Asn
 610 615 620
 Asn Asn Asn Asn Asn Asn Asp Glu Asn Cys Asp Asn Asn Asn Asn His
 625 630 635 640
 Asn Asn Asn Asn Tyr Asn Asn Asn Asn Asn Tyr Gly Asn Asn Asn Asn
 645 650 655
 Asn Asn Asn Asn Asn Lys Asp Asn Asn Asn Asn Asp Gly Asn Gly Ser
 660 665 670
 Ser Asn Asn Asn Asn Asn Asp Asp Asp Asp Glu Glu Glu Glu Asp Asp
 675 680 685

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Glu Asp Asp Asn Asn Asn Asn Asn Asp Asp Asp Asn Met Ser Asp Asn
 690 695 700
 Glu Glu Met Glu Asp Asn Asp Glu Asp Asn Asp Glu Tyr Asn Asn Ser
 705 710 715 720
 Asn Asp Ser Tyr Lys Tyr Glu Glu Lys Asp Ser Asn His Glu Lys Asp
 725 730 735
 Leu Lys Lys Asp Ile Ile Glu Gly Asp Met Ile Asn Ser Val Lys Tyr
 740 745 750
 Asp Lys Asn Ile Gly His His Thr Thr Asn Lys Ser Glu Ile Ser Thr
 755 760 765
 Asn Tyr Phe Glu Asn Ser Cys Asn Met Ser Val Asn Asn Ser Asn Asn
 770 775 780
 Glu Ala Tyr Asp Asp Asn Cys Asn Asn Gly Phe Met Asn His Asp Glu
 785 790 795 800
 Gly Leu Thr Leu Asn Asn Gly Asn Val Ser Asn Asn Lys Cys Asp Ile
 805 810 815
 Ile Ile Pro Glu Asp Gly Ser Val Met Tyr Glu Asn Met Ile Asn Arg
 820 825 830
 Gly Asn Gly Leu Thr Ser Asn Ile Asn Asn Asn Asn Val Ser Asn
 835 840 845
 Asn Asn Ser Ile Ser Cys Asn Ala Asp Asp Asn Val Tyr Asn Asn Ile
 850 855 860
 Asn Asn Tyr Ile Asn Thr Tyr Met Glu Thr Thr Asn Asn Lys Asn His
 865 870 875 880
 Ile Glu Asn Arg Cys Asn Gln Asp Ser Tyr Ser Thr Asn Glu Glu Pro
 885 890 895
 Leu Ser Asn His Ser Ile Asn Asp Pro Gly Lys Ile Lys Asp Gly Ile
 900 905 910
 Met Tyr Asp Gly Asn Asp Leu Asp Met Asn Gly Thr Gln Glu His Ser
 915 920 925
 Lys Glu Glu Gly Met Asp Val Phe Glu Pro Asn Phe Phe Glu Leu Lys
 930 935 940
 Arg Asn Ser Ser Asp Gly Gln Asn Lys His Leu Glu Pro Gly Val Gln
 945 950 955 960
 Lys Lys Ile Ser Lys Lys Arg Ser Lys Val Lys His Glu Arg Asn Ser
 965 970 975
 Lys Ile Leu Asp Asp Glu Lys Lys Glu Val Leu Asn Lys Val Ser Gln
 980 985 990
 Ile Thr Arg Val Gly Gly Val Cys Phe Asp Lys Asn Arg Gln Arg Trp
 995 1000 1005

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Ile Ala	His Trp Lys	Ile Asp	Gly Lys Tyr	His Lys	His Tyr Phe
1010		1015		1020	
Pro Ile	Ser Gln Tyr Gly	Phe	Glu Asn Ala Arg	Glu	Arg Ala Val
1025		1030		1035	
Ser Cys	Arg Lys Gln Ala	Glu	Lys Leu Phe Asn	Leu	Pro Glu Ile
1040		1045		1050	
Gln Pro	Arg Asn Arg Trp	Asn	Gln Ile Lys Val	Asn	Gly Thr Ser
1055		1060		1065	
His Ile	Lys Lys Ala Ala	Lys	Leu Pro Arg Cys	Glu	Gly Ile Gly
1070		1075		1080	
Tyr Asp	Glu Leu Ser Gln	Ser	Trp Val Ser Thr	Phe	Val Val His
1085		1090		1095	
Lys Lys	Phe Ser Ile Glu	Glu	Leu Gly Phe Tyr	Glu	Ala Arg Glu
1100		1105		1110	
Lys Ala	Ile Tyr Cys Arg	Lys	Thr Phe Glu Lys	Val	Asn Val His
1115		1120		1125	
Asp Asp	Tyr Glu Cys Leu	Leu	Asn Asp Arg Leu	Gly	Leu Arg Asn
1130		1135		1140	
Glu Glu	Lys Asp Glu Leu	Ser	Asp Leu Ile Asn	Ile	Asp Lys Asn
1145		1150		1155	
Ala Leu	Asp Asn Leu Glu	Leu	Glu Thr Ser Val	His	Asn Asn Asn
1160		1165		1170	
Lys Val	Lys His Asn Asn	Asn	Asn Asn Asn Asn	Asn	Asn Asn Asn
1175		1180		1185	
Asn Asn	Asn Asn Asn Asn	Asn	Asn Ser Glu Lys	Met	Arg Ile Lys
1190		1195		1200	
Asn Asn	Asp Phe Ser Val	Asp	Asn Asn Asn Glu	Asn	Val Gly Thr
1205		1210		1215	
Gly Glu	Ile Lys Ile Ser	Asn	Asp Lys Tyr Leu	Lys	Ile Thr Gln
1220		1225		1230	
Glu Ala	Ile Glu Met Ile	Leu	Ser Asn Ile Lys	His	Lys Ser Leu
1235		1240		1245	
Pro Glu	Ile Lys Met Lys	Leu	Ile Asp Lys Gln	Lys	Phe Glu Asn
1250		1255		1260	
Tyr Asn	Thr Leu Leu Asp	Lys	His Phe Lys Phe	Ile	Thr Ser Val
1265		1270		1275	
Lys Asn	Ile Ser Gln Leu	Arg	Arg Tyr Ile Ser	Leu	Phe His Lys
1280		1285		1290	
Phe Ile	Ile Tyr His Thr	Leu	Pro His Asn Ile	Ser	Leu Arg Lys
1295		1300		1305	

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Gln Leu Phe Ile Ile Glu Ala Leu Glu Trp Ser Ser Phe Phe Ser
 1310 1315 1320

Gly Ala Ala Ser Glu Lys Val Glu
 1325 1330

<210> 51
 <211> 291
 <212> PRT
 <213> Plasmodium falciparum

<400> 51

Met Glu Val Thr Ser Thr Leu Leu Glu Lys Gly Lys Asn Phe Ala Gln
 1 5 10 15

Asp Pro Ser Glu Val Phe Pro Glu Ser Lys Lys Phe Phe Phe Ser Ser
 20 25 30

Ile Val Cys Leu Lys Thr Asn Phe Asp Lys Arg Thr Gly Ala Leu Gly
 35 40 45

Tyr Leu Asn Leu Ser Tyr Gly Met Gly Ile Ile Phe Gly Ser Phe Leu
 50 55 60

Ala Gly Val Met Val Asn Phe Val Gly Ser Arg Gly Asn Leu Leu Ile
 65 70 75 80

Ala Leu Leu Ser Gln Leu Ile Ala Leu Cys Ile Ser Thr Thr Leu Glu
 85 90 95

Glu Asp Pro Lys Leu Leu Lys Ser Ser Asn Val Asp Lys Met Lys Met
 100 105 110

Ser Glu Ile Leu Leu Ser Ile Lys Asn Glu Tyr Ile Arg Val Leu Asn
 115 120 125

Leu Phe Lys Lys Thr Tyr Gly Ile Cys Leu Leu Ile Leu Phe Gly Leu
 130 135 140

Leu Pro Ile Leu Met Thr Lys Phe Ala Phe Ala Pro Val Val Val Asp
 145 150 155 160

Met Phe Lys Leu Thr Pro Ser His Thr Ser Tyr Leu Met Thr Tyr Ala
 165 170 175

Gly Ile Ile Thr Ile Ile Ala Glu Gly Ile Leu Ala Pro Tyr Leu Ser
 180 185 190

Ser Leu Leu Gly Asp Met Ile Cys Cys Lys Tyr Ser Ile Pro Leu Thr
 195 200 205

Leu Thr Gly Phe Leu Leu Leu Ser Leu Cys Gly Ala Asn Glu Ser Leu
 210 215 220

Val Leu Ile Phe Met Ser Ile Pro Leu Cys Gly Gly Ala Leu Leu Tyr
 225 230 235 240

Ile Cys Gly Thr Ser Gln Met Thr Lys Arg Val Glu Glu Ser Glu Leu

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	245		250		255
Gly Ser Ile	Ile Gly Leu Asn Thr Ser	Leu Phe Tyr Ala Val Thr Ile			
	260	265		270	
Ile Ala Pro	Tyr Ile Ala Phe Lys Ser Tyr Ile Ala Leu Gly Leu Gly				
	275	280		285	
Leu Tyr Trp					
	290				
<210>	52				
<211>	903				
<212>	PRT				
<213>	Plasmodium falciparum				
<400>	52				
Met Arg Ile Trp Gly Lys Asp Val Phe Ala Gly Phe Val Thr Lys Lys					
1	5		10		15
Leu Lys Thr Leu Leu Asp Cys Asn Phe Ala Leu Tyr Tyr Asn Phe Lys					
	20		25		30
Gly Asn Gly Pro Asp Ala Gly Ser Phe Leu Asp Phe Val Asp Glu Pro					
	35		40		45
Glu Gln Phe Tyr Trp Phe Val Glu His Phe Leu Ser Val Lys Phe Arg					
	50		55		60
Val Pro Lys His Leu Lys Asp Lys Asn Ile His Asn Phe Thr Pro Cys					
65		70		75	80
Leu Asn Arg Ser Trp Val Ser Glu Phe Leu Lys Glu Tyr Glu Glu Pro					
	85		90		95
Phe Val Asn Pro Val Met Lys Phe Leu Asp Lys Glu Gln Arg Leu Phe					
	100		105		110
Phe Thr Tyr Asn Phe Gly Asp Val Glu Pro Gln Gly Lys Tyr Thr Tyr					
	115		120		125
Phe Pro Val Lys Glu Phe His Lys Tyr Cys Ile Leu Pro Pro Leu Ile					
	130		135		140
Lys Thr Asn Ile Lys Asp Gly Glu Ser Gly Glu Phe Leu Lys Tyr Gln					
145		150		155	160
Leu Asn Lys Glu Glu Tyr Lys Val Phe Leu Ser Ser Val Gly Ser Gln					
	165		170		175
Met Thr Ala Ile Lys Asn Leu Tyr Ser Thr Val Glu Asp Glu Gln Arg					
	180		185		190
Lys Gln Leu Leu Lys Val Ile Ile Glu Asn Glu Ser Thr Asn Asp Ile					
	195		200		205
Ser Val Gln Cys Pro Thr Tyr Asn Ile Lys Leu His Tyr Thr Lys Glu					
	210		215		220

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Cys Ala Asn Ser Asn Asn Ile Leu Lys Cys Ile Asp Glu Phe Leu Arg
 225 230 235 240
 Lys Thr Cys Glu Lys Lys Thr Glu Ser Lys His Pro Ser Ala Asp Leu
 245 250 255
 Cys Glu His Leu Gln Phe Leu Phe Glu Ser Leu Lys Asn Pro Tyr Leu
 260 265 270
 Asp Asn Phe Lys Lys Phe Met Thr Asn Ser Asp Phe Thr Leu Ile Lys
 275 280 285
 Pro Gln Ser Val Trp Asn Val Pro Ile Phe Asp Ile Tyr Lys Pro Lys
 290 295 300
 Asn Tyr Leu Asp Ser Val Gln Asn Leu Asp Thr Glu Cys Phe Lys Lys
 305 310 315 320
 Leu Asn Ser Lys Asn Leu Ile Phe Leu Ser Phe His Asp Asp Ile Pro
 325 330 335
 Asn Asn Pro Tyr Tyr Asn Val Glu Leu Gln Glu Ile Val Lys Leu Ser
 340 345 350
 Thr Tyr Thr Tyr Ser Ile Phe Asp Lys Leu Tyr Asn Phe Phe Phe Val
 355 360 365
 Phe Lys Lys Ser Gly Ala Pro Ile Ser Pro Val Ser Val Lys Glu Leu
 370 375 380
 Ser His Asn Ile Thr Asp Phe Ser Phe Lys Glu Asp Asn Ser Glu Ile
 385 390 395 400
 Gln Cys Gln Asn Val Arg Lys Ser Leu Asp Leu Glu Val Asp Val Glu
 405 410 415
 Thr Met Lys Gly Ile Ala Ala Glu Lys Leu Cys Lys Ile Ile Glu Lys
 420 425 430
 Phe Ile Leu Thr Lys Asp Asp Ala Ser Lys Pro Glu Lys Ser Asp Ile
 435 440 445
 His Arg Gly Phe Arg Ile Leu Cys Ile Leu Ile Ser Thr His Val Glu
 450 455 460
 Ala Tyr Asn Ile Val Arg Gln Leu Leu Asn Met Glu Ser Met Ile Ser
 465 470 475 480
 Leu Thr Arg Tyr Thr Ser Leu Tyr Ile His Lys Phe Phe Lys Ser Val
 485 490 495
 Thr Leu Leu Lys Gly Asn Phe Leu Tyr Lys Asn Asn Lys Ala Ile Arg
 500 505 510
 Tyr Ser Arg Ala Cys Ser Lys Ala Ser Leu His Val Pro Ser Val Leu
 515 520 525
 Tyr Arg Arg Asn Ile Tyr Ile Pro Glu Thr Phe Leu Ser Leu Tyr Leu
 530 535 540

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Gly Leu Ser Asn Leu Val Ser Ser Asn Pro Ser Ser Pro Phe Phe Glu
 545 550 555 560
 Tyr Ala Ile Ile Glu Phe Leu Val Thr Tyr Tyr Asn Lys Gly Ser Glu
 565 570 575
 Lys Phe Val Leu Tyr Phe Ile Ser Ile Ile Ser Val Leu Tyr Ile Asn
 580 585 590
 Glu Tyr Tyr Tyr Glu Gln Leu Ser Cys Phe Tyr Pro Lys Glu Phe Glu
 595 600 605
 Leu Ile Lys Ser Arg Met Ile His Pro Asn Ile Val Asp Arg Ile Leu
 610 615 620
 Lys Gly Ile Asp Asn Leu Met Lys Ser Thr Arg Tyr Asp Lys Met Arg
 625 630 635 640
 Thr Met Tyr Leu Asp Phe Glu Ser Ser Asp Ile Phe Ser Arg Glu Lys
 645 650 655
 Val Phe Thr Ala Leu Tyr Asn Phe Asp Ser Phe Ile Lys Thr Asn Glu
 660 665 670
 Gln Leu Lys Lys Lys Asn Leu Glu Glu Ile Ser Glu Ile Pro Val Gln
 675 680 685
 Leu Glu Thr Ser Asn Asp Gly Ile Gly Tyr Arg Lys Gln Asp Val Leu
 690 695 700
 Tyr Glu Thr Asp Lys Pro Gln Thr Met Asp Glu Ala Ser Tyr Glu Glu
 705 710 715 720
 Thr Val Asp Glu Asp Ala His His Val Asn Glu Lys Gln His Ser Ala
 725 730 735
 His Phe Leu Asp Ala Ile Ala Glu Lys Asp Ile Leu Glu Glu Lys Thr
 740 745 750
 Lys Asp Gln Asp Leu Glu Ile Glu Leu Tyr Lys Tyr Met Gly Pro Leu
 755 760 765
 Lys Glu Gln Ser Lys Ser Thr Ser Ala Ala Ser Thr Ser Asp Glu Ile
 770 775 780
 Ser Gly Ser Glu Gly Pro Ser Thr Glu Ser Thr Ser Thr Gly Asn Gln
 785 790 795 800
 Gly Glu Asp Lys Thr Thr Asp Asn Thr Tyr Lys Glu Met Glu Glu Leu
 805 810 815
 Glu Glu Ala Glu Gly Thr Ser Asn Leu Lys Lys Gly Leu Glu Phe Tyr
 820 825 830
 Lys Ser Ser Leu Lys Leu Asp Gln Leu Asp Lys Glu Lys Pro Lys Lys
 835 840 845
 Lys Lys Ser Lys Arg Lys Lys Lys Arg Asp Ser Ser Ser Asp Arg Ile
 850 855 860

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Leu Leu Glu Glu Ser Lys Thr Phe Thr Ser Glu Asn Glu Leu Met Arg
865 870 875 880

Lys Lys Lys Lys Lys Lys Lys Lys Lys Asn Asn Asn Glu Ile Lys Asn
885 890 895

Ile Arg Ile Tyr Tyr Asn Leu
900

<210> 53

<211> 743

<212> PRT

<213> Plasmodium falciparum

<400> 53

Met Met Asn Met Lys Ile Val Leu Phe Ser Leu Leu Leu Phe Val Ile
1 5 10 15

Arg Trp Asn Ile Ile Ser Cys Asn Lys Asn Asp Lys Asn Gln Gly Val
20 25 30

Asp Met Asn Val Leu Asn Asn Tyr Glu Asn Leu Phe Lys Phe Val Lys
35 40 45

Cys Glu Tyr Cys Asn Glu His Thr Tyr Val Lys Gly Lys Lys Ala Pro
50 55 60

Ser Asp Pro Gln Cys Ala Asp Ile Lys Glu Glu Cys Lys Glu Leu Leu
65 70 75 80

Lys Glu Lys Gln Tyr Thr Asp Ser Val Thr Tyr Leu Met Asp Gly Phe
85 90 95

Lys Ser Ala Asn Asn Ser Ala Asn Asn Gly Lys Lys Asn Asn Ala Glu
100 105 110

Glu Met Lys Asn Leu Val Asn Phe Leu Gln Ser His Lys Lys Leu Ile
115 120 125

Lys Ala Leu Lys Lys Asn Ile Glu Ser Ile Gln Asn Lys Lys His Leu
130 135 140

Ile Tyr Lys Asn Lys Ser Tyr Asn Pro Leu Leu Leu Ser Cys Val Lys
145 150 155 160

Lys Met Asn Met Leu Lys Glu Asn Val Asp Tyr Ile Gln Lys Asn Gln
165 170 175

Asn Leu Phe Lys Glu Leu Met Asn Gln Lys Ala Thr Tyr Ser Phe Val
180 185 190

Asn Thr Lys Lys Lys Ile Ile Ser Leu Lys Ser Gln Gly His Lys Lys
195 200 205

Glu Thr Ser Gln Asn Gln Asn Glu Asn Asn Asp Asn Gln Lys Tyr Gln
210 215 220

Glu Val Asn Asp Glu Asp Asp Val Asn Asp Glu Glu Asp Thr Asn Asp
225 230 235 240

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Asp	Glu	Asp	Thr	Asn	Asp	Glu	Glu	Asp	Thr	Asn	Asp	Asp	Glu	Asp	Thr	245	250	255
Asn	Asp	Asp	Glu	Asp	Thr	Asn	Asp	Glu	Glu	Asp	Thr	Asn	Asp	Glu	Glu	260	265	270
Asp	His	Glu	Asn	Asn	Asn	Ala	Thr	Ala	Tyr	Glu	Leu	Gly	Ile	Val	Pro	275	280	285
Val	Asn	Asp	Val	Leu	Asn	Val	Asn	Met	Lys	Asn	Met	Ile	Thr	Gly	Asn	290	295	300
Asn	Phe	Met	Asp	Val	Val	Lys	Asn	Thr	Leu	Ala	Gln	Ser	Gly	Gly	Leu	305	310	315
Gly	Ser	Asn	Asp	Leu	Ile	Asn	Phe	Leu	Asn	Gln	Gly	Lys	Glu	Ile	Gly	325	330	335
Glu	Asn	Leu	Leu	Asn	Ile	Thr	Lys	Met	Asn	Leu	Gly	Asp	Lys	Asn	Asn	340	345	350
Leu	Glu	Ser	Phe	Pro	Leu	Asp	Glu	Leu	Asn	Met	Leu	Lys	Asp	Asn	Leu	355	360	365
Ile	Asn	Tyr	Glu	Phe	Ile	Leu	Asp	Asn	Leu	Lys	Thr	Ser	Val	Leu	Asn	370	375	380
Lys	Leu	Lys	Asp	Leu	Leu	Leu	Arg	Leu	Leu	Tyr	Lys	Ala	Tyr	Val	Ser	385	390	395
Tyr	Lys	Lys	Arg	Lys	Ala	Gln	Glu	Lys	Gly	Leu	Pro	Glu	Pro	Thr	Val	405	410	415
Thr	Asn	Glu	Glu	Tyr	Val	Glu	Glu	Leu	Lys	Lys	Gly	Ile	Leu	Asp	Met	420	425	430
Gly	Ile	Lys	Leu	Leu	Phe	Ser	Lys	Val	Lys	Ser	Leu	Leu	Lys	Lys	Leu	435	440	445
Lys	Asn	Lys	Ile	Phe	Pro	Lys	Lys	Lys	Glu	Asp	Asn	Gln	Ala	Val	Asp	450	455	460
Thr	Lys	Ser	Met	Glu	Glu	Pro	Lys	Val	Lys	Ala	Gln	Pro	Ala	Leu	Arg	465	470	475
Gly	Val	Glu	Pro	Thr	Glu	Asp	Ser	Asn	Ile	Met	Asn	Ser	Ile	Asn	Asn	485	490	495
Val	Met	Asp	Glu	Ile	Asp	Phe	Phe	Glu	Lys	Glu	Leu	Ile	Glu	Asn	Asn	500	505	510
Asn	Thr	Pro	Asn	Val	Val	Pro	Pro	Thr	Gln	Ser	Lys	Lys	Lys	Asn	Lys	515	520	525
Asn	Glu	Thr	Val	Ser	Gly	Met	Asp	Glu	Asn	Phe	Asp	Asn	His	Pro	Glu	530	535	540
Asn	Tyr	Phe	Lys	Glu	Glu	Tyr	Tyr	Tyr	Asp	Glu	Asn	Asp	Asp	Met	Glu	545	550	555

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Val Lys Val Lys Lys Ile Gly Val Thr Leu Lys Lys Phe Glu Pro Leu
565 570 575

Lys Asn Gly Asn Val Ser Glu Thr Ile Lys Leu Ile His Leu Gly Asn
580 585 590

Lys Asp Lys Lys His Ile Glu Ala Ile Asn Asn Asp Ile Gln Ile Ile
595 600 605

Lys Gln Glu Leu Gln Ala Ile Tyr Asn Glu Leu Met Asn Tyr Thr Asn
610 615 620

Gly Asn Lys Asn Ile Gln Gln Ile Phe Gln Gln Asn Ile Leu Glu Asn
625 630 635 640

Asp Val Leu Asn Gln Glu Thr Glu Glu Glu Met Glu Lys Gln Val Glu
645 650 655

Ala Ile Thr Lys Gln Ile Glu Ala Glu Val Asp Ala Leu Ala Pro Lys
660 665 670

Asn Lys Glu Glu Glu Glu Lys Glu Lys Glu Lys Glu Glu Lys Glu Lys
675 680 685

Glu Glu Lys Glu Lys Glu Lys Glu Glu Lys Glu Lys Glu Glu Lys Glu
690 695 700

Lys Glu Glu Lys Glu Lys Glu Glu Lys Glu Glu Glu Lys Lys Glu Lys
705 710 715 720

Glu Glu Glu Gln Glu Glu Glu Glu Glu Glu Glu Ile Val Pro Glu Asn
725 730 735

Leu Thr Thr Glu Glu Ser Lys
740

<210> 54

<211> 1137

<212> DNA

<213> Plasmodium falciparum

<400> 54

ggagaagcag taactccttc cgtaattgat aacatacttt ctaaaattga aaatgaatat 60

gagggttttat attttaaacc tttagcaggt gtttatagaa gtttaaaaaa acaattagaa 120

aataacgtta tgacatttaa tgtaaatggt aaggatattt taaattcacg atttaataaa 180

cgtgaaaatt tcaaaaatgt tttagaatca gatttaattc catataaaga tttaacatca 240

agtaattatg ttgtcaaaga tccatataaa tttcttaata aagaaaaaag agataaattc 300

ttaagcagtt ataattatat taaggattca atagatacgg atataaattt tgcaaatgat 360

gttcttggat attataaaat attatccgaa aaatataaat cagatttaga ttcaattaaa 420

aaatatatca acgacaaaca aggtgaaaat gagaaatacc ttcccttttt aaacaatatt 480

gagaccttat ataaaacagt taatgataaa attgatttat ttgtaattca tttagaagca 540

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aaagttctaa attatacata tgagaaatca aacgtagaag ttaaaataaa agaacttaat    600
tacttaaaaa caattcaaga caaattggca gatttttaaaa aaaataacaa tttcggttga    660
attgctgatt tatcaacaga ttataaccat aataacttat tgacaaagtt ccttagtaca    720
ggtatgggtt ttgaaaatct tgctaaaacc gttttatcta atttacttga tggaaacttg    780
caaggatatgt taaacatttc acaacaccaa tgcgtaaaaa aacaatgtcc acaaaattct    840
ggatgtttca gacattttaga tgaaagagaa gaatgtaaat gtttattaaa ttacaaacaa    900
gaaggatgata aatgtgttga aaatccaaat cctacttgta acgaaaataa tgggtggatgt    960
gatgcagatg ccaaagtac cgaagaagat tcaggtagca acggaagaa aatcacatgt    1020
gaatgtacta aacctgattc ttatccactt ttcgatggta ttttctgcag ttcctctaac    1080
ttcttaggaa tatcattctt attaatactc atgttaatat tatacagttt catttaa    1137

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<210> 55

<211> 1080

<212> DNA

<213> Plasmodium falciparum

<400> 55

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caggataaac ccgaagtaag tgcaaatgat gatacatcac attctacaaa tttgaataat    60
agttttaaatt tatttgaaaa catattgagt cttggaaaaa acaaaaatat ataccaagaa    120
ttaataggtc aaaaaagtag tgaaaacttt tatgaaaaga tattaaga tagtgatata    180
ttttataatg aatcttttac aaattttgta aaatctaaag ctgatgatat taattcattg    240
aatgatgaat caaaaaggaa gaaattagaa gaagatatta ataaattaaa aaaaacttta    300
cagttatcat ttgatattata taataaatat aaattaaaat tagaaagatt atttgataaa    360
aagaaaacag ttggtaaata taaaatgcaa attaaaaaac ttactttatt aaaagaacaa    420
ttagaatcaa aattgaattc acttaataac ccaaagcatg tattacaaaa cttttctggt    480
ttctttaaca aaaaaaaga agctgaaata gcagaaactg aaaacacatt agaaaacaca    540
aaaatattat tgaacatta taaaggactt gttaaattat ataattggtga atcatctcca    600
ttaaaaactt taagtgaaga atcaattcaa acagaagata attatgccag tttagaaaac    660
tttaaagtat taagttaatt agaaggaaaa ttaaaggata atttaaattt agaaaagaaa    720
aaattatcat acttatcaag tggattacat catttaattg ctgaattaaa agaagtaata    780
aaaaataaaa attatacagg taattctcca agtgaaaata atacggatgt taacaatgca    840
ttagaatctt acaaaaaatt tctcccagaa ggaacagatg ttgcaacagt tgtaagtga    900
agtggatccg acacattaga acaaagtcaa ccaaagaaac cagcatcaac tcatgtagga    960

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gcagagtcta acacaataac aacatcacaa aatgtcgatg atgaagtaga tgacgtaatc 1020
 atagtaccta tatttggaga atccgaagaa gattatgatg atttaggaca agtagtaaca 1080

<210> 56
 <211> 660
 <212> DNA
 <213> Plasmodium falciparum

<400> 56
 caggataaac ccgaagtaag tgcaaatgat gatacatcac attctacaaa tttgaataat 60
 agttttaaata tatttgaaaa catattgagt cttggaaaaa acaaaaatat ataccaagaa 120
 ttaataggtc aaaaaagtag tgaaaacttt tatgaaaaga tattaaga tagtgatata 180
 ttttataatg aatcttttac aaattttgta aaatctaaag ctgatgatata taattcattg 240
 aatgatgaat caaaaaggaa gaaattagaa gaagatatata ataaattaaa aaaaacttta 300
 cagttatcat ttgatttata taataaatat aaattaaaat tagaaagatt atttgataaa 360
 aagaaaacag ttggtaata taaaatgcaa attaaaaaac ttactttatt aaaagaacaa 420
 ttagaatcaa aattgaattc acttaataac ccaaagcatg tattacaaaa cttttctggt 480
 ttctttaaca aaaaaaaga agctgaaata gcagaaactg aaaacacatt agaaaacaca 540
 aaaatattat tgaaacatta taaaggactt gttaaatatt ataatggtga atcatctcca 600
 ttaaaaactt taagtgaaga atcaattcaa acagaagata attatgccag ttagaaaac 660

<210> 57
 <211> 1080
 <212> DNA
 <213> Plasmodium falciparum

<400> 57
 caggataaac ccgaagtaag tgcaaatgat gatacatcac attctacaaa tttgaataat 60
 agttttaaata tatttgaaaa catattgagt cttggaaaaa acaaaaatat ataccaagaa 120
 ttaataggtc aaaaaagtag tgaaaacttt tatgaaaaga tattaaga tagtgatata 180
 ttttataatg aatcttttac aaattttgta aaatctaaag ctgatgatata taattcattg 240
 aatgatgaat caaaaaggaa gaaattagaa gaagatatata ataaattaaa aaaaacttta 300
 cagttatcat ttgatttata taataaatat aaattaaaat tagaaagatt atttgataaa 360
 aagaaaacag ttggtaata taaaatgcaa attaaaaaac ttactttatt aaaagaacaa 420
 ttagaatcaa aattgaattc acttaataac ccaaagcatg tattacaaaa cttttctggt 480
 ttctttaaca aaaaaaaga agctgaaata gcagaaactg aaaacacatt agaaaacaca 540
 aaaatattat tgaaacatta taaaggactt gttaaatatt ataatggtga atcatctcca 600

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ttaaaaactt taagtgaaga atcaattcaa acagaagata attatgccag tttagaaaac	660
tttaaagtat taagtaaatt agaaggaaaa ttaaaggata atttaaattt agaaaagaaa	720
aaattatcat acttatcaag tggattacat catttaattg ctgaattaaa agaagtaata	780
aaaaataaaa attatacagg taattctcca agtgaaaata atacggatgt taacaatgca	840
ttagaatctt acaaaaaaatt tctcccagaa ggaacagatg ttgcaacagt tgtaagtga	900
agtggatccg acacattaga acaaagtcaa ccaaagaaac cagcatcaac tcatgtagga	960
gcagagtcta acacaataac aacatcacia aatgtcgatg atgaagtaga tgacgtaatc	1020
atagtaccta tatttggaga atccgaagaa gattatgatg atttaggaca agtactaaca	1080

<210> 58

<211> 1131

<212> DNA

<213> Plasmodium falciparum

<400> 58

gcagtaactc cttccgtaat tgataacata ctttctaaaa ttgaaatga atatgaggtt	60
ttatatattaa aaccttttagc aggtgtttat agaagttaa aaaaacaatt agaaaataac	120
gttatgacat ttaatgttaa tgtaaggat attttaaatt cagatttaa taaacgtgaa	180
aatttcaaaa atgttttaga atcagattta attccatata aagatttaa atcaagtaat	240
tatgttgtca aagatccata taaatttctt aataaagaaa aaagagataa attcttaagc	300
agttataatt atattaagga ttcaatagat acggatataa attttgcaaa tgatgttctt	360
ggatattata aaatattatc cgaaaaatat aaatcagatt tagattcaat taaaaatat	420
atcaacgaca aacaagggtga aaatgagaaa taccttcctt ttttaacaa tattgagacc	480
ttatataaaa cagttaatga taaaattgat ttatttgtaa ttcatttaga agcaaaagtt	540
ctaaattata catatgagaa atcaaacgta gaagttaaaa taaaagaact taattactta	600
aaaacaattc aagacaaatt ggcagatttt aaaaaaata acaatttcgt tggaattgct	660
gatttatcaa cagattataa ccataataac ttattgacaa agttccttag tacaggtag	720
gtttttgaaa atcttgctaa aaccgtttta tctaatttac ttgatggaaa cttgcaaggt	780
atgttaaaca tttcacaaca ccaatgcgta aaaaaacaat gtccacaaaa ttctggatgt	840
ttcagacatt tagatgaaag agaagaatgt aaatgtttat taaattacaa acaagaaggt	900
gataaatgtg ttgaaatcc aaatcctact tgtaacgaaa ataatgggtg atgtgatgca	960
gatgccaaat gtaccgaaga agattcaggt agcaacggaa agaaaatcac atgtgaatgt	1020
actaaacctg attcttatcc acttttcgat ggtattttct gcagttcctc taacttctta	1080
ggaatatcat tcttattaat actcatgtta atattataca gtttcattta a	1131

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<210> 59
<211> 343
<212> DNA
<213> Plasmodium falciparum

<400> 59
catttcacaa caccaatgcg taaaaaaaca atgtccacaa aattctggat gtttcagaca 60
tttagatgaa agagaagaat gtaaatgttt attaaattac aaacaagaag gtgataaatg 120
tggtgaaaat ccaaataccta cttgtaacga aaataatggg ggatgtgatg cagatgccaa 180
atgtaccgaa gaagattcag gtagcaacgg aaagaaaatc acatgtgaat gtactaaacc 240
tgattcttat ccacttttcg atgggtatatt ctgcagttcc tctaacttct taggaatatc 300
attcttatta atactcatgt taatattata cagtttcatt taa 343